

Standard Paper

A new *Calicium* on *Ramboldia*

Leif Tibell¹ , Stephen R. Clayden² , Maria Prieto^{3,4}  and Mats Wedin⁵ 

¹Department of Organismal Biology, Uppsala University, 75236 Uppsala, Sweden; ²Department of Natural History, New Brunswick Museum, Saint John, NB E2K 1E5, Canada; ³Área de Biodiversidad y Conservación, Departamento de Biología y Geología, Física y Química Inorgánica, Escuela Superior de Ciencias Experimentales y Tecnología (ES CET), Universidad Rey Juan Carlos, 28933 Móstoles, Madrid, Spain; ⁴Instituto de Investigación en Cambio Global de la Universidad Rey Juan Carlos, 28933 Móstoles, Madrid, Spain and ⁵Department of Botany, Swedish Museum of Natural History, SE-10405 Stockholm, Sweden

Abstract

A lichenicolous species, *Calicium ramboldiicola*, growing on *Ramboldia elabens* is described. In phylogenetic analyses with 22 *Calicium* taxa, based on 121 sequences from five DNA regions (mtSSU, *Mcm7*, nuITS, nuLSU, β -tubulin), the new species formed a strongly supported clade with *C. abietinum* and *C. verrucosum*. Although the ascomata of the new species resemble miniatures of those of *C. abietinum*, no morphological synapomorphies for this clade were found. *Calicium ramboldiicola* is known from boreal and hemiboreal areas of northern Europe and north-eastern North America. It is the second known lichenicolous species of *Calicium* and the first found on a lichen in the *Lecanorales*.

Keywords: Ascomycota; biodiversity; ecology; lichen; lichenicolous; phylogeny; taxonomy

(Accepted 3 October 2024)

Introduction

The genus *Calicium* Ach. in *Caliciaceae* Chevall. includes mazaediate fungi (Prieto *et al.* 2013) with stalked ascomata and mostly 1-septate, dark, thick-walled spores with distinctive surface ornamentation. A recent molecular study supports a wider circumscription of the genus to also include some species with sessile ascomata that were previously placed in *Cyphelium* Ach. (Prieto & Wedin 2017). However, *Cyphelium sessile* (Pers.) Trevis. and *C. marcianum* B. de Lesd., which are lichenicolous on species of *Pertusaria* and *Lepora*, were transferred to *Acolium* (Prieto & Wedin 2017).

Regional revisions of *Calicium* species in Australasia (Tibell 1987), Europe (Tibell 1999a), North America (Selva *et al.* 2023a, b), South America (Tibell 1996, 1998) and the Himalayas (Tibell 2006) have contributed descriptions of numerous species, and *Calicium* has thus been rather well investigated on a global scale. With the new circumscription and the recent description of additional species, it is known to include 36 species (Tibell & Knutsson 2016; Prieto & Wedin 2017; Selva *et al.* 2023a). Among these is the recently described *Calicium episcalaris* Tibell & Knutsson, parasitic on *Hypocenomyce scalaris* (Ach. ex Lilj.) M. Choisy (Tibell & Knutsson 2016; Selva *et al.* 2023b). All other described species of *Calicium* have a distinct, autonomous thallus that varies from thick and verrucose in some species to immersed in others.

Three of the authors were therefore surprised to encounter, independently of each other, what looked like a perfectly ‘classical’

small *Calicium* growing on the thallus of *Ramboldia elabens* (Fr.) Kantvilas & Elix in boreal and hemiboreal areas of northern Europe and north-eastern North America. The first collection was made by one of us (LT) in 1977 in Sweden. Since then, we have found the species at additional localities in Sweden, Norway and Canada, and we have also detected several other specimens in herbaria.

Material and Methods

Microscopy

Squash preparations and hand-cut sections of apothecia were mounted in water, 10% KOH (K), 1.5% Lugol’s solution (I), and lactophenol cotton blue (LCB), and examined with light microscopy (LM). Measurements of ascospores and anatomical characters were made in water and are reported as (minimum–)a–b(–maximum) (n = number of measurements), where ‘a’ is the arithmetic mean minus 1 standard deviation (SD) and ‘b’ is the arithmetic mean plus 1 SD. Apothecial dimensions were measured in air-dried specimens examined with a stereomicroscope. For scanning electron microscopy (SEM), air-dried specimens were mounted on stubs, coated with gold in an Edwards S150A sputter coater, and examined at 15 kV in a JEOL2 JSM 6400 instrument at the University of New Brunswick.

Taxon sampling

Five specimens of the new species and a wide selection of species of the genus *Calicium* were included in the phylogenetic analysis (Table 1). Based on previous studies (Prieto & Wedin 2017), three

Corresponding author: Leif Tibell; Email: leif.tibell@gmail.com

Cite this article: Tibell L, Clayden SR, Prieto M and Wedin M (2024) A new *Calicium* on *Ramboldia*. *Lichenologist* 56, 301–307. <https://doi.org/10.1017/S0024282924000343>



members of the *Caliciaceae* (*Diplotomma alboatrum* (Hoffm.) Flot., *Pyxine cocoes* (Sw.) Nyl. and *Pyxine soreliata* (Ach.) Mont.) were used as outgroups.

Molecular techniques

Sequences of *Calicium* were either downloaded from GenBank or produced in the laboratory for the new species. DNA was extracted using the DNeasy Plant Mini Kit (Qiagen), according to the manufacturer's instructions. Five regions were amplified: nuITS, nuLSU, mtSSU, β -tubulin and *Mcm7*. The nuITS was amplified with the primers ITS1F (Gardes & Bruns 1993) and ITS4 (White *et al.* 1990), and the nuLSU with LR0R (Rehner & Samuels 1994), LR3, LR3R, LR6 and LR7 (Vilgalys & Hester 1990). We used the primers mtSSU1 and mtSSU3R for amplification of the mtSSU region (Zoller *et al.* 1999), *Mcm7*-709for and *Mcm7*-1348rev (Schmitt *et al.* 2009) for amplification of the *Mcm7* region and Bt3-LM and Bt10-LM for the protein coding β -tubulin (Myllys *et al.* 2001).

PCR amplifications were carried out using Illustra™ Hot Start Mix RTG PCR beads (GE Healthcare, UK) in 25 μ l, adding 3–6 μ l of diluted genomic DNA, 10 μ M of each primer and distilled water. The following amplification programme was used: initial denaturation at 95 °C for 15 min, 35 cycles of 95 °C for 45 s, 54–56 °C for 50 s, 72 °C for 1 min, and a final extension at 72 °C for 5 min. PCR products were purified with ExoSAP-IT (USB Corporation, Santa Clara, California, USA) and sequenced using the same amplification primers.

Sequences were assembled and edited using Sequencher v. 4.10.1. (Genes Codes Corporation, Ann Arbor, Michigan, USA) and subsequently aligned using MacClade v. 4.01 (Maddison & Maddison 2001). Ambiguous regions (*sensu* Lutzoni *et al.* 2000) and introns were delimited manually and excluded from phylogenetic analyses.

Phylogenetic analyses

Each individual gene region was analyzed using maximum likelihood (ML) implemented in RAxML v. 8.2.12 (Stamatakis 2014) with a GTRGAMMA model for tree inference and bootstrapping with a GTRCAT model and 1000 replicates. Gene-tree incongruence was checked by comparing maximum likelihood bootstrap values (ML-BS) between the individual gene trees. After checking there was no conflict among clades, data were combined into a single concatenated data matrix. The combined maximum likelihood (ML) analysis was run with seven distinct partitions (nuITS, nuLSU, mtSSU, first and second codon positions of the *Mcm7* and β -tubulin and the third codon position of the *Mcm7* and β -tubulin) as individual analyses. All analyses were run on the CIPRES Science Gateway v. 3.3 (Miller *et al.* 2010). The resulting trees were visualized and edited in Figtree v. 1.4.4. (Rambaut 2010).

Results

The concatenated data matrix consisted of 25 taxa, 22 being *Calicium* species, and three outgroups. *Calicium* was represented by 109 sequences, of which 15 are new. In the analysis (Fig. 1), *Calicium* was strongly supported, and within the genus two major clades are apparent. *Calicium ramboldiicola*, together with *C. abietinum* and *C. verrucosum* Tibell, belongs to a strongly supported subclade. The other lichenicolous *Calicium* species, *C. episcalaris*, belongs to the same major clade as *C. ramboldiicola*.

Taxonomy

Calicium ramboldiicola Tibell, S. R. Clayden & Wedin

Mycobank No.: MB 856766

Resembling *Calicium abietinum* but occurring on *Ramboldia elabens*, lacking a distinct thallus, and with smaller apothecia, shorter asci, and ascospores with more coarsely and irregularly cracked ornamentation.

Type: Sweden, Lule Lappmark, Jokkmokk parish, River Pärälven, W of Lake Närkejaure, valley of the brook Laddonbäcken, on decorticated fallen *Pinus*, 66°36'22"N, 19°16'54"E, alt. c. 360 m, 2021, *Wedin* 9898 (S—holotype; CANL—iso-type). The type has a full representation of all five DNA regions used in this study, and was chosen for this reason.

(Fig. 2)

Thallus not evident, immersed in the thallus of *Ramboldia elabens*, not causing obvious necrosis or discoloration of the host lichen.

Apothecia on the areoles and rarely the apothecia of *R. elabens*, stalked to nearly sessile, with a black capitulum and dark olive-brown stalk, shiny except for the black mazaedium, lacking pruina, (0.25–)0.30–0.44(–0.55) mm tall ($n = 30$), all parts K–, I–. *Stalk* 0.08–0.20 mm diam., consisting of densely interwoven thick-walled hyphae with lumina 0.7–2 μ m wide, central part dark olive-brown, grading to pale brown to hyaline in outer part, with an outermost clear gelatinous layer 5–10 μ m wide. *Capitulum* initially globose to obovoid, becoming flat-topped-cupulate, (0.15–)0.17–0.27(–0.35) mm diam. ($n = 30$). *Excipulum* to c. 50 μ m thick at base, narrowing to c. 20 μ m laterally, structure continuous with that of the stalk, of \pm isodiametric to somewhat elongated cells with strongly thickened walls, inner part dark olive-brown, outermost hyaline gelatinous layer thinner than on the stalk. *Hypothecium* c. 30–60 μ m tall, slightly concave at top, upper part concolorous with inner part of exciple and stalk, lower part hyaline to very pale brown. *Asci* cylindrical to narrowly clavate, (34–)35.8–41.8(–43) \times (4–)4.2–5.4(–6) μ m ($n = 10$), with uniseriate but somewhat irregularly orientated spores. *Ascospores* initially greenish grey, soon becoming dark brown, ellipsoid, 1-septate, constricted or not at septum, surface initially smooth to finely roughened, developing coarse irregular cracks, (9.5–)10.8–13.9(–16.5) \times (5–)5.7–7.2(–8) μ m ($n = 73$).

Etymology. Named after the genus of the host lichen, *Ramboldia elabens*.

Distribution. *Calicium ramboldiicola* is currently known from Norway and Sweden in northern Europe, and from the eastern Canadian provinces of New Brunswick and Québec in north-eastern North America. The occurrences in Canada are in intermediate oceanic–continental and continental (OC and C₁) sectors of the hemiboreal bioclimatic zone (*sensu* Tuhkanen 1984; Clayden 2010). In Scandinavia, *C. ramboldiicola* has a more distinctly boreal distribution. To date, it has been found at eight localities, spanning c. 10° of latitude, from the southern to northern sub-zones of the boreal zone and their elevational counterparts as defined by Ahti *et al.* (1968). These occurrences are in areas with intermediate oceanic–continental (OC, O₁) climates, not the more highly oceanic sectors (O₂, O₃) of westernmost Scandinavia.

Further study is needed to determine whether the distribution of *C. ramboldiicola* conforms more fully to that of its host lichen.

Table 1. Details on taxa used in the phylogenetic analyses, including voucher information and GenBank Accession numbers. New species and sequences are in bold.

Species name	Voucher information	GenBank Accession number				
		mtSSU	<i>Mcm7</i>	nuITS	nuLSU	β -tubulin
<i>Calicium abietinum</i>	Tibell 25061 (UPS)	KX512971	KX529041	KX512905	KX512872	KX529003
<i>C. adspersum</i>	Prieto 3037 (S)	KX512949	KX529055	KX512907	KX512895	KX529022
<i>C. chlorosporum</i> 1	Tibell 25012 (UPS)	KX512956	KX529059	-	KX512892	-
<i>C. chlorosporum</i> 2	Thor 20859 (UPS)	KX512955	-	-	-	-
<i>C. corynellum</i>	Prieto CC1 (S)	KX512985	KX529048	KX512908	KX512855	KX528989
<i>C. denigratum</i>	Prieto (S) F209596	KX512965	KX529044	KX512909	KX512878	-
<i>C. episcalaris</i> 1	Knutsson (UPS)	-	-	KX228552	-	-
<i>C. episcalaris</i> 2	Soun 1013 (ROK)	-	-	OQ717343	-	-
<i>C. episcalaris</i> 3	Vondrák 24110 (PRA)	OQ646151	-	OQ717763	-	-
<i>C. glaucellum</i>	Wedin 8563 (S)	KX512980	KX529032	KX512910	KX512864	-
<i>C. lecideinum</i>	Prieto (S) F473050	KX512961	KX529046	KX512911	KX512882	KX529009
<i>C. lenticulare</i>	Tibell 23284 (UPS)	KX512979	KX529033	KX512912	-	KX528997
<i>C. montanum</i>	van den Boom 23445 (UPS)	-	-	KX529069	KX512853	-
<i>C. nobile</i> 1	Tibell 21968 (UPS)	KX512988	KX529060	KX512913	KX529070	-
<i>C. nobile</i> 2	Tibell 23396 (UPS)	KX512987	KX529061	KX512914	KX529071	-
<i>C. notarisii</i>	Prieto 3007 (S)	KX512960	KX529047	KX512915	KX512883	KX529011
<i>C. parvum</i> 1	Vondrák 26019a (PRA)	OQ682893	-	OQ717345	-	-
<i>C. parvum</i> 2	Vondrák 23767 (PRA)	OQ646152	-	OQ717765	-	-
<i>C. pinastri</i>	Vondrák 21874/ Vondrák 24287 (PRA)	OQ682894	-	OL396663	-	-
<i>C. pinicola</i> 1	Lendemmer & Knudsen 14982 (UPS)	KX512972	KX529040	KX512916	KX512871	KX529015
<i>C. pinicola</i> 2	Thor 19856 (UPS)	KX512991	KX529066	KX512917	KX512887	KX529014
<i>C. quercinum</i>	Tibell 22287 (UPS)	-	-	KX512918	KX512854	-
<i>C. ramboldiicola</i> 1	Wedin 9898 (S)	PQ308401	PQ161417	PQ304346	PQ304346	PQ150070
<i>C. ramboldiicola</i> 2	Johannesson & Wadstein (UPS)	PQ308402	-	PQ304347	-	PQ150071
<i>C. ramboldiicola</i> 3	Clayden 28675 (NBM)	PQ308403	PQ161418	-	-	-
<i>C. ramboldiicola</i> 4	Wedin 7234 (UPS)	PQ308405	-	PQ304348	-	-
<i>C. ramboldiicola</i> 5	Hermansson 13315 (UPS)	PQ308404	PQ161419	PQ304349	-	-
<i>C. salicinum</i>	Prieto (S) F209764	KX512982	KX529027	KX512919	KX512861	KX528991
<i>C. tigillare</i>	Prieto 3038 (S)	JX000123	JX000162	JX000104	JX000088	KX529002
<i>C. trabinellum</i>	Wedin 8517 (S)	-	KX529026	KX512920	KX512858	KX528995
<i>C. verrucosum</i>	Tibell 23198 (UPS)	-	KX529030	-	-	-
<i>C. viride</i>	Wedin 24/4 2000	AY584696	JX000153	HQ650703	AY340538	KX529013
<i>Diplotomma albostrum</i>	Prieto 3034 (S)	KX512966	KX529043	KX512924	KX512877	KX529007
<i>Pyxine coccoes</i>	Prieto (S) F473049	KX512964	-	KX512936	-	KX529010
<i>P. soledata</i>	Wetmore 91254 (S)	KX512973	KX529039	KX512937	KX512870	KX529001

Ramboldia elabens occurs widely in boreal and temperate-montane areas of the Northern Hemisphere (Kantvilas & Elix 2007). In North America, it has a mainly Appalachian–Great Lakes distribution, with outliers as far west as the Black Hills of South Dakota (Wetmore 1967). Reports of *R. elabens* from areas of North America west of the Black Hills are based on mis-identifications of other lichens (Pérez-Ortega *et al.* 2010). In

Eurasia, it occurs discontinuously from Scandinavia and sub-alpine western and central Europe (Hafellner 1993) to Japan (Inoue 1982) and the Russian Far East (e.g. Konoreva *et al.* 2018).

Ecology. *Calicium ramboldiicola* has been found exclusively on the crustose lichen *Ramboldia elabens* growing on hard exposed wood ('lignum') of conifers, including standing snags and fallen

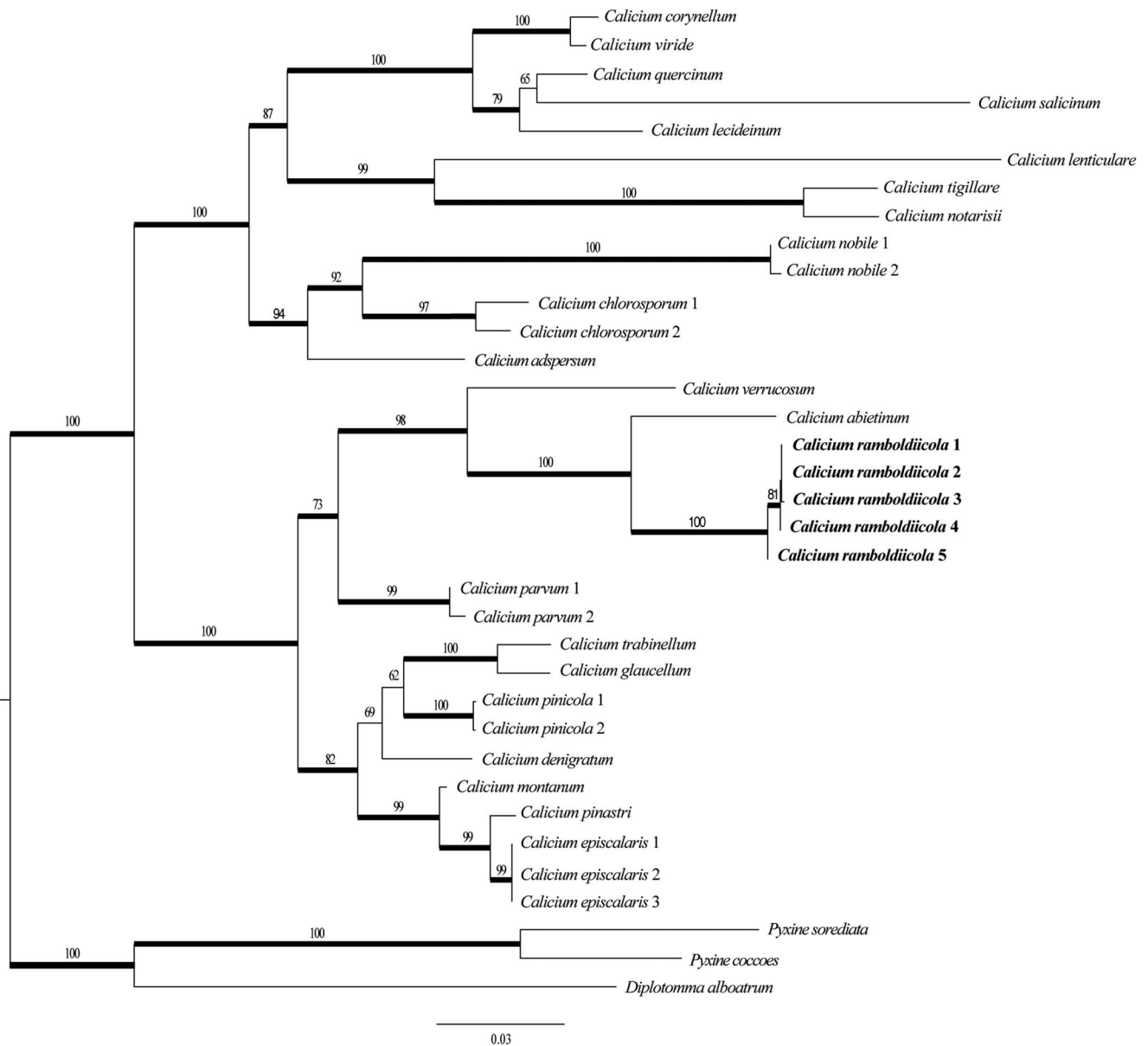


Figure 1. Phylogeny of *Calicium* using a combination of five loci (nuITS, nuLSU, mtSSU, *Mcm7* and β -tubulin) resulting from maximum likelihood (ML) in RAxML, with bootstrap values shown above nodes. Strongly supported branches (ML \geq 70) and *C. ramboldiicola* are shown in bold.

trees. *Ramboldia elabens* also occurs less frequently on hard, weathered wood of built structures such as old barns or fences, but no occurrences of *C. ramboldiicola* are known on host thalli occupying such substrata. In Scandinavia, all samples are from very old *Pinus sylvestris* L. snags in old-growth forest localities. In New Brunswick, the habitat at one of the localities is an open, south-west-facing, granitic boulder talus slope with patches of *Picea rubens* Sarg. and *Pinus strobus* L. At the other, the habitat is an open, wet, ombrotrophic bog, with scattered low thickets of *Larix laricina* (Du Roi) K. Koch and *Picea mariana* (Mill.) Britton et al. Associated lignicolous lichens at one or both localities in New Brunswick include *Imshaugia aleurites* (Ach.) S. L. F. Mey., *Mycoblastus alpinus* (Fr.) Kernst., *M. sanguinarius* (L.) Norman, *Ochrolechia mahluensis* Räsänen, *O. pseudopallescens* Brodo, *Pertusaria sulcata* Dibben, *Xylographa disseminata* Willey, and *X. vitiligo* (Ach.) J. R. Laundon.

Although the thallus of *R. elabens* is often moderately thick, with contiguous, somewhat lumpy areoles contrasting with the black, shiny apothecia, it can also be thin and even to mostly endoxylic towards the thallus margin or more extensively. Apothecia of *Calicium ramboldiicola* are present mainly on well-developed areoles of *R. elabens* (Fig. 2A), but also on more or less endoxylic portions of some thalli (Fig. 2C), and less commonly on the host apothecia. Because of the small amount or age of the material available for study, we have not examined areoles histologically to assess whether the hyphae of *C. ramboldiicola* are distinguishable from those of *R. elabens*, or whether there are differences in the abundance or appearance of photobiont cells in areoles of *R. elabens* with or without apothecia of *C. ramboldiicola*. Older apothecia of *R. elabens* sometimes develop uneven or wrinkled discs, but these changes occur in thalli with or without *C. ramboldiicola*. Apothecia of *C. ramboldiicola* are sporadically present on

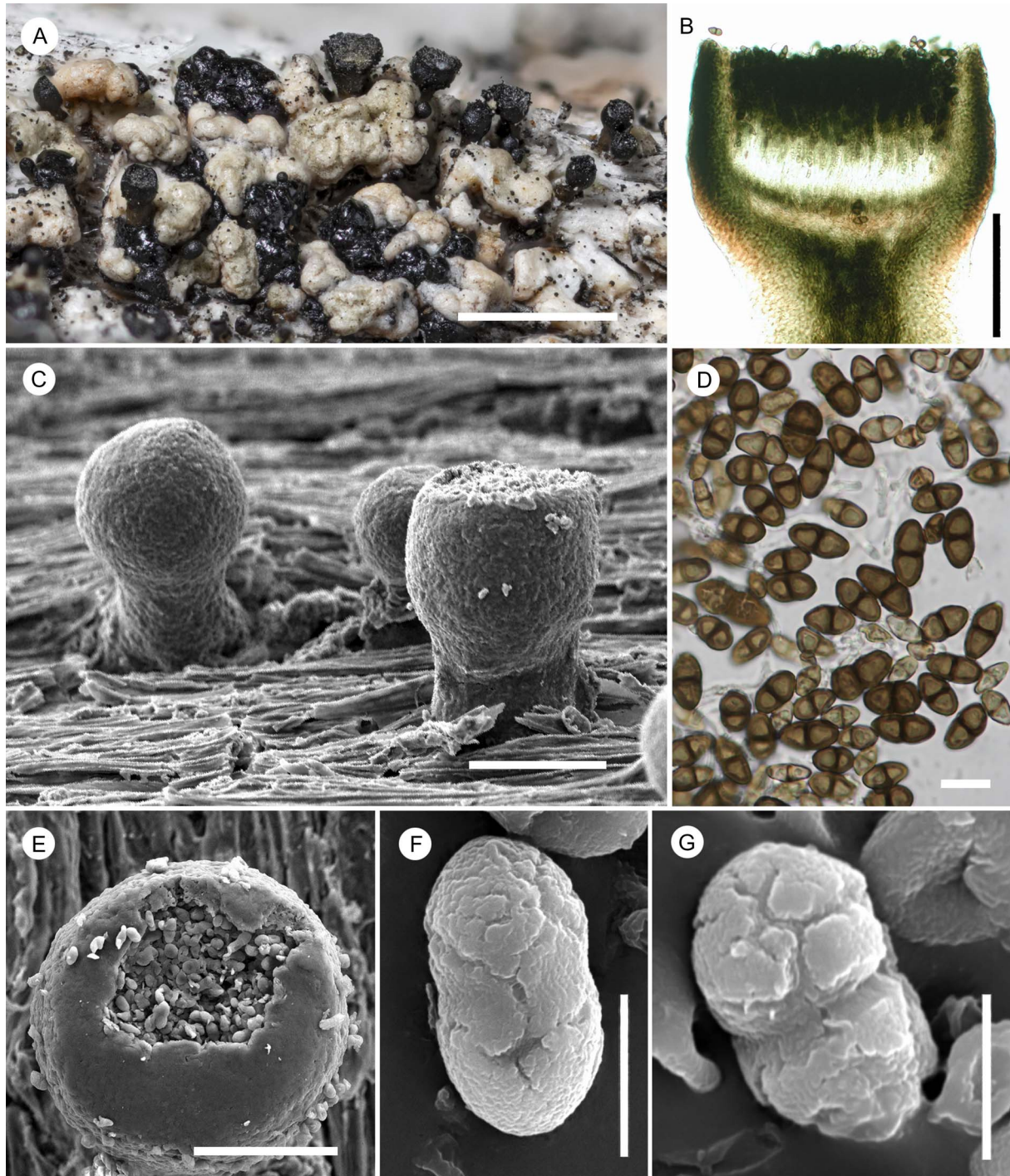


Figure 2. *Calicium ramboldiicola*. A, apothecia arising from thallus and apothecia of *Ramboldia elabens* (Clayden 21343). B, longitudinal section of mature apothecium, LM (Clayden 21343). C, immature and mature apothecia, arising from thin to endoxylic area of thallus of *R. elabens*, SEM (Clayden 24619). D, mature ascospores, liberated from asci, LM (Clayden 28675). E, capitulum in oblique view, with wall rupturing apically and exposing the mazaedium, SEM (Clayden 24619). F, mature ascospore with surface ornamentation formed by irregular cracking, SEM (Clayden 21343). G, mature ascospore constricted at septum and with coarsely ornamented surface, SEM (Clayden 21343). LM = light microscopy; SEM = scanning electron microscopy. Scales: A = 1 mm; B & C = 100 μ m; D = 10 μ m; E = 50 μ m; F & G = 5 μ m. In colour online.

eroded or senescent areoles of the host lichen, but these occurrences may be accidental. That is, the deterioration of such areoles could have been caused by agents other than the *Calicium*.

Additional specimens examined. **Canada:** *New Brunswick:* Kings Co., Canadian Forces Base Gagetown, Nerepis Hills, 1 km NE of

confluence of Askwith Brook with Nerepis River, 45.442°N, 66.298°W, alt. 140 m, on decorticated snag of *Picea rubens*, 2010, Clayden 21343 (NBM); *ibid.*, 2022, Clayden 28675 (NBM); Sunbury Co., Bull Pasture Bog Protected Natural Area, 46.0524°N, 66.3113°W, alt. 100 m, lignicolous on snag of *Larix laricina*, 2014, Clayden 24619 (NBM). *Québec:* Parc de la

Vérendrye, Réservoir Cabonga, Îles Bruyères, 47°14'30"N, 76°34'30"W, sur un vieux tronc de conifère en bordure du réservoir, 1991, *Bastien* 789 (QFA).—**Norway:** *Troms:* Storfjord, 13.5 km SSE of Skibotn, along Skibotnelva, above Gustavsvingen, 69°16'N, 20°28'E, alt. 120–200 m, 2003, *Wedin* 7234 (UPS). *Buskerud:* Nore of Uvdal, Viken, Godfarfoss S, 60.4312°N, 8.605°E, 2021, *Reiso* (O L-2289081).—**Sweden:** *Dalarna:* Los parish, Gönhammaren, on dry, still standing *Pinus*, *Forsslund* (UPS); Ore parish, Jässelåsen, Rärmyren, 61°14'N, 15°16'E, alt. 370 m, on dry, still standing *Pinus* snags, 2003, *Hermansson* 13315 (UPS); Svärdsjö parish, E of Orrtjärnen, on dry snag in a bog, 2024, *Skog* (S). *Jämtland:* Undersåker parish c. 15.5 km S of Undersåker church, south side of Lake Häckrenmagasinet, c. 570 m E of Furbergskojan, 63.19080°N, 13.42355°E, 2020, *Johannesson & Wadstein* (UPS). *Lule Lappmark:* Gällivare parish, Linaåive Nature Reserve, E of Parajaure, 67°16'N, 20°26'E, alt. 490 m, on decorticated branches of fallen, dead *Pinus*, 1977, *Tibell* 6916 (UPS); *ibid.*, Jokkmokk par., Karats area, SW of Mt Luspevarasj, 66.62713°N, 18.81745°E, elev. 440 m, 2021, *Westberg* SCN150 (UPS).

Discussion

Fungi with stalked ascomata resembling those of *Calicium* are a heterogeneous group from a phylogenetic standpoint, but they are often studied together and thus form a defined research area (Temu et al. 2024). Although lichenicolous species occur in many calicioid genera, especially *Chaenothecopsis* and *Sphinctrina* (both in the *Mycocaliciales*; Tibell & Wedin 2000), *Calicium ramboldiicola* is one of rather few lichenicolous species discovered in *Caliciaceae*. In this family, *Acolium marcianum* (B. de Lesd.) M. Prieto & Wedin and *A. sessile* (Pers.) Arnold are associated with *Pertusaria* and *Lepra* species (*Pertusariaceae* and *Variolariaceae*, respectively; both *Pertusariales*), whereas the species described here is found with *Ramboldia* (*Ramboldiaceae*, *Lecanorales*; Miadlikowska et al. 2014). This is only the second *Calicium* species known to occur on *Lecanoromycetes*, the other being *Calicium episcalaris*, on *Hypocenomyce* (*Ophioparmaceae*, *Umbilicariales*). Other genera of *Caliciaceae* including small numbers of lichenicolous species are *Diplotomma* and *Tetramelas* (Cannon et al. 2021).

The ascomata of *C. ramboldiicola* are small but otherwise similar to those of other species in *Calicium*, such as *C. glaucellum*. In our analysis (Fig. 1), *C. ramboldiicola* is sister to *C. abietinum*, and indeed resembles a miniature of that species, with much smaller ascomata but different spore ornamentation: *C. ramboldiicola* has an irregularly cracked spore surface, while that of *C. abietinum* presents with minute, distinct warts. The clade including *C. abietinum*, *C. ramboldiicola* and *C. verrucosum* has high support (Fig. 1), but apart from a general similarity in ascoma morphology, we have not been able to find any specific morphological synapomorphies.


Calicium ramboldiicola also resembles *C. pinastri* in its small size, lack of pruina, short cylindrical asci, and spore ornamentation. However, the apothecial stalks in *C. pinastri* are black, lacking the brown hue that is usually present in *C. ramboldiicola*. Also, *C. pinastri* is a mainly corticolous species, in Europe occurring especially on trunks of *Pinus sylvestris* (Tibell 1999a, b). Our phylogenetic analysis (Fig. 1) indicates that these two species are not closely related to one another within *Calicium*.

Ramboldia elabens is currently red-listed in Sweden and categorized as near-threatened (SLU Artdatabanken 2020). As *C.*

ramboldiicola is known only from this host, it makes the new lichenicolous species extremely rare *per se*. Similar to *R. elabens*, it probably occurs only in localities of very high conservation value. Thus, it should be given consideration as a threatened species and probably placed in a higher category than its host.

Acknowledgements. SRC is grateful to Steven Selva for his encouragement of the studies reported here, and to Claude Roy and Philip Bell-Doyon for their generous help at the Herbarium Louis-Marie (QFA), Université Laval. Permission to enter and make collections in CFB Gagetown, New Brunswick (NB), Canada, was arranged by Deanna McCullum, Range Biologist with the Canadian Department of National Defence. Collecting in Bull Pasture Bog PNA, NB, was carried out through the BiotaNB program of the New Brunswick Museum, with the support of the NB Wildlife Trust Fund and a research permit from the NB Department of Natural Resources and Energy Development. SRC also thanks Roger Smith of Fredericton, NB, for macrophotography and preparation of Fig. 2, and Nhu Trieu and Steven Cogswell for SEM work in the Microscopy and Microanalysis Facility at the University of New Brunswick. MW was supported by funds from the Swedish Royal Academy of Sciences (KVA). We thank the Molecular Systematic Laboratory (MSL) at the Swedish Museum of Natural History, in particular Bodil Cronholm, for skilful laboratory assistance. We are also indebted to Associate Editor, Paul Diederich, and to two anonymous reviewers for their helpful comments on the manuscript.

Author Contribution. LT made the initial discovery, conceived and directed the study, and prepared the original draft of the manuscript. SRC carried out field and morphological studies. MP performed the phylogenetic analyses. MW carried out field, laboratory and morphological studies. All four authors contributed to writing and revising the manuscript.

Author ORCIDs.  Leif Tibell, 0000-0002-8629-7989; Stephen R. Clayden, 0000-0004-9031-2151; María Prieto, 0000-0002-1692-9821; Mats Wedin, 0000-0002-8295-5198.

References

- Ahti T, Hämet-Ahti L and Jalas J (1968) Vegetation zones and their sections in northwestern Europe. *Annales Botanici Fennici* 5, 169–211.
- Cannon P, Prieto M, Coppins B, Sanderson N, Scheidegger C and Simkin J (2021) *Caliciales: Caliciaceae*, including the genera *Acolium*, *Amandinea*, *Buellia*, *Calicium*, *Diploicia*, *Diplotomma*, *Endohyalina*, *Monerolechia*, *Orcularia*, *Pseudothelomma*, *Rinodina* and *Tetramelas*. *Revisions of British and Irish Lichens* 15, 1–35.
- Clayden SR (2010) Lichens and allied fungi of the Atlantic Maritime Ecozone. In McAlpine DF and Smith IM (eds), *Assessment of Species Diversity in the Atlantic Maritime Ecozone*. Ottawa: NRC Research Press, pp. 153–178.
- Gardes M and Bruns TD (1993) ITS primers with enhanced specificity for basidiomycetes – application to the identification of mycorrhizae and rusts. *Molecular Ecology* 2, 113–118.
- Hafellner J (1993) Die Gattung *Pyrrhospora* in Europa. Eine erste Übersicht mit einem Bestimmungsschlüssel der Arten nebst Bemerkungen zu einigen außereuropäischen Taxa (lichenisierte *Ascomycotina*, *Lecanorales*). *Herzogia* 9, 725–747.
- Inoue M (1982) The genera *Lecidea*, *Lecidella* and *Huilia* (Lichens) in Japan I. *Lecidea*. *Journal of Science of the Hiroshima University, Series B, Division 2 (Botany)* 18, 1–55.
- Kantvilas G and Elix J (2007) The genus *Ramboldia* (*Lecanoraceae*): a new species, key and notes. *Lichenologist* 39, 135–141.
- Konoreva IA, Tchabanenko SI, Ezhkin AK, Schumm F and Chesnokov SV (2018) New and noteworthy lichen and allied fungi records from Sakhalin Island, Far East of Russia. *Herzogia* 31, 276–292.
- Lutzoni F, Wagner P, Reeb V and Zoller S (2000) Integrating ambiguously aligned regions of DNA sequences in phylogenetic analyses without violating positional homology. *Systematic Biology* 49, 628–651.
- Maddison WP and Maddison DR (2001) *MacClade: Analysis of Phylogeny and Character Evolution, version 4.01*. Sunderland, Massachusetts: Sinauer Associates.

- Miadlikowska J, Kauff F, Högnabba F, Oliver JC, Molnár K, Fraker E, Gaya E, Hafellner J, Hofstetter V, Gueidan C, et al. (2014) A multigene phylogenetic synthesis for the class *Lecanoromycetes* (Ascomycota): 1307 fungi representing 1139 infrageneric taxa, 317 genera and 66 families. *Molecular Phylogenetics and Evolution* **79**, 132–168.
- Miller MA, Pfeiffer W and Schwartz T (2010) Creating the CIPRES Science Gateway for inference of large phylogenetic trees. In *Proceedings of the Gateway Computing Environments Workshop (GCE), 14 November 2010, New Orleans, Louisiana*, pp. 1–8.
- Myllys L, Lohtander K and Tehler A (2001) Beta-tubulin, ITS and group I intron sequences challenge the species pair concept in *Physcia aipolia* and *P. caesia*. *Mycologia* **93**, 335–343.
- Pérez-Ortega S, Spribille T, Palice Z, Elix JA and Printzen C (2010) A molecular phylogeny of the *Lecanora varia* group, including a new species from western North America. *Mycological Progress* **9**, 523–535.
- Prieto M and Wedin M (2017) Phylogeny, taxonomy and diversification events in the *Caliciaceae*. *Fungal Diversity* **82**, 221–238.
- Prieto M, Baloch E, Tehler A and Wedin M (2013) Mazaedium evolution in the Ascomycota (Fungi) and the classification of mazaediate groups of formerly unclear relationship. *Cladistics* **29**, 296–308.
- Rambaut A (2010) *FigTree v.1.3.1*. Institute of Evolutionary Biology, University of Edinburgh. [WWW resource] URL <http://tree.bio.ed.ac.uk/software/figtree/>
- Rehner S and Samuels GJ (1994) Taxonomy and phylogeny of *Gliocladium* analysed from nuclear large subunit ribosomal DNA sequences. *Mycological Research* **98**, 625–634.
- Schmitt I, Crespo A, Divakar PK, Fankhauser JD, Herman-Sackett E, Kalb K, Nelsen MP, Nelson NA, Rivas-Plata E, Shimp AD, et al. (2009) New primers for promising single-copy genes in fungal phylogenetics and systematics. *Persoonia* **23**, 35–40.
- Selva SB, Tibell L, Gordon M and McMullin RT (2023a) *Calicium sperlingiae* (Caliciaceae), a new species of calicioid lichen from Douglas County, Oregon, U.S.A. *Bryologist* **126**, 236–241.
- Selva SB, McMullin RT, Bell-Doyon P, Henderson B and Lay E (2023b) Calicioid lichens and fungi in North America: species new to science, reported as new from elsewhere and placed into synonymy. *Bryologist* **126**, 427–446.
- SLU Artdatabanken (2020) *Rödlistade arter i Sverige 2020 (Red-listed Species in Sweden 2020)*. Uppsala: SLU.
- Stamatakis A (2014) RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* **30**, 1312–1313.
- Temu SG, Tibell S, Tibuhwa DD and Tibell L (2024) *Coniocybe* Ach. revisited. *Journal of Fungi* **10**, 363.
- Tibell L (1987) Australasian *Caliciales*. *Symbolae Botanicae Upsalienses* **27**, 1–279.
- Tibell L (1996) *Caliciales*. *Flora Neotropica* **69**, 1–78.
- Tibell L (1998) Crustose mazaediate lichens and the *Mycocaliciaceae* in temperate South America. *Bibliotheca Lichenologica* **71**, 1–107.
- Tibell L (1999a) Calicioid lichens and fungi. In Ahti T, Jørgensen PM, Kristinsson H, Moberg R, Søchting U and Thor G (eds), *Nordic Lichen Flora Vol. 1*. Uddevalla, Sweden: Nordic Lichen Society, pp. 20–94.
- Tibell L (1999b) Two new species of *Calicium* from Europe. *Mycotaxon* **70**, 431–433.
- Tibell L (2006) *Calicium* in the Indian Himalayas. *Journal of the Hattori Botanical Laboratory* **100**, 809–851.
- Tibell L and Knutsson T (2016) *Calicium episcalaris* (Caliciaceae), a new lichen species from Sweden. *Symbolae Botanicae Upsalienses* **38**, 49–52.
- Tibell L and Wedin M (2000) *Mycocaliciales*, a new order for nonlichenized calicioid fungi. *Mycologia* **92**, 577–581.
- Tuhkanen S (1984) A circumboreal system of climatic-phytogeographical regions. *Acta Botanica Fennica* **127**, 1–50.
- Vilgalys R and Hester M (1990) Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. *Journal of Bacteriology* **172**, 4238–4246.
- Wetmore CM (1967) Lichens of the Black Hills of South Dakota and Wyoming. *Publications of the Museum, Michigan State University, Biology Series* **3**, 209–464.
- White TJ, Bruns T, Lee S and Taylor JW (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In Innis MA, Gelfand DH, Sninsky JJ and White TJ (eds), *PCR Protocols: a Guide to Methods and Applications*. New York: Academic Press, pp. 315–322.
- Zoller S, Scheidegger C and Sperisen C (1999) PCR primers for the amplification of mitochondrial small subunit ribosomal DNA of lichen-forming ascomycetes. *Lichenologist* **31**, 511–516.