Newborn Screening for Spinal Muscular Atrophy: Ontario Testing and Follow-up Recommendations

Hugh J. McMillan[®]*, Kristin D. Kernohan^{*}, Ed Yeh, Kim Amburgey, Jennifer Boyd, Craig Campbell, James J. Dowling, Hernan Gonorazky, Janet Marcadier, Mark A. Tarnopolsky[®], Jiri Vajsar, Alex MacKenzie, Pranesh Chakraborty

ABSTRACT: *Background:* Spinal muscular atrophy (SMA) is characterized by the progressive loss of motor neurons causing muscle atrophy and weakness. Nusinersen, the first effective SMA therapy was approved by Health Canada in June 2017 and has been added to the provincial formulary of all but one Canadian province. Access to this effective therapy has triggered the inclusion of SMA in an increasing number of Newborn Screening (NBS) programs. However, the range of disease-modifying *SMN2* gene copy numbers encountered in survival motor neuron 1 (*SMN1*)-null individuals means that neither screen-positive definition nor resulting treatment decisions can be determined by *SMN1* genotype alone. We outline an approach to this challenge, one that specifically addresses the case of SMA newborns with four copies of *SMN2*. *Objectives:* To develop a standardized post-referral evaluation pathway for babies with a positive SMA NBS screen result. *Methods:* An SMA NBS pilot trial in Ontario using first-tier MassARRAY and second-tier multi-ligand probe amplification (MLPA) was launched in January 2020. Prior to this, Ontario pediatric neuromuscular disease and NBS experts met to review the evidence regarding the diagnosis and treatment of children with SMA as it pertained to NBS. A post-referral evaluation algorithm was developed, outlining timelines for patient retrieval and management. *Conclusions:* Ontario's pilot NBS program has created a standardized path to facilitate early diagnosis of SMA and initiation of treatment. The goal is to provide timely access to those SMA infants in need of therapy to optimize motor function and prolong survival.

RÉSUMÉ : Dépistage de l'amyotrophie spinale chez le nouveau-né : recommandations sur le dépistage et le suivi de la maladie en Ontario. *Contexte :* L'amyotrophie spinale (AS) se caractérise par une perte progressive des neurones moteurs, ce qui entraîne une atrophie et une faiblesse musculaires. Nusinersen, le premier traitement efficace de l'AS a été approuvé par Santé Canada en juin 2017, et ajouté à la liste des médicaments assurés dans toutes les provinces, sauf dans une seule. L'arrivée de ce traitement efficace a eu pour effet d'ajouter l'AS dans un nombre croissant de programmes de dépistage néonatal (PDN). Toutefois, comme le nombre de copies du gène *SMN2* chez les personnes « nulles » à l'égard de *SMN1* est modifié par plus d'une maladie, ni la définition d'un test de dépistage positif ni les décisions relatives au traitement qui en découle ne peuvent reposer sur le seul génotype *SMN1*. Aussi avons-nous élaboré une démarche qui vise spécifiquement l'AS à 4 copies du gène *SMN2* chez le nouveau-né (N.-N.). *Objectif :* L'étude visait à tracer le chemin à suivre après la consultation pour l'évaluation des N.-N. ayant obtenu un résultat positif au dépistage de l'AS dans le cadre du PDN. *Méthode :* Un essai pilote de dépistage néonatal de l'AS réalisé à l'aide de MassARRAY au premier tour et de l'amplification multiplex de sondez dépendant d'une ligation (MLPA) au second tour été lancé en Ontario, en janvier 2020. Auparavant, des spécialistes des maladies neuromusculaires chez les enfants et des experts du PDN en Ontario se sont rencontrés afin d'examiner les données probantes sur le diagnostic et le traitement de l'AS chez les enfants, dans le cadre du PDN. A suivi l'élaboration d'un algorithme d'évaluation après consultation, qui établissait les différentes étapes à suivre en vue de l'aiguillage du patient et de la prise en charge de la maladie. *Conclusion :* Le programme pilote de dépistage néonatal de l'Ontario a permis d'uniformiser la démarche à suivre et ainsi de faciliter la pose précoce du

Keywords: Spinal muscular atrophy, Neonatal screening, Mass screening, Genetics

doi:10.1017/cjn.2020.229

Can J Neurol Sci. 2021; 48: 504-511

From the Children's Hospital of Eastern Ontario Research Institute, Department of Pediatrics, University of Ottawa, Ottawa, Ontario, Canada (HJMM, KDK, EY, AMK, PC); Newborn Screening Ontario, Ottawa, Ontario, Canada (KDK, EY, JM, PC); Hospital for Sick Children, Department of Pediatrics, University of Toronto, Toronto, Ontario, Canada (KA, JB, JJD, HG, JV); Children's Hospital Western Ontario, Department of Pediatrics, Bridemiology and Clinical Neurological Sciences, Schulich School of Medicine, University of Western Ontario, London, Ontario, Canada (CC); and McMaster Children's Hospital, Department of Pediatrics, McMaster University, Hamilton, Ontario, Canada (MAT) RECEIVED MAY 26, 2020. FINAL REVISIONS SUBMITTED SEPTEMBER 3, 2020. DATE OF ACCEPTANCE OCTOBER 11, 2020.

Correspondence to: Pranesh Chakraborty, Newborn Screening Ontario, and Alex MacKenzie, Molecular Biomedicine Program), Children's Hospital of Eastern Ontario Research Institute, 401 Smyth Road, Ottawa, Ontario K1H 8L1, Canada. Emails: pchakraborty@cheo.on.ca, mackenzie@cheo.on.ca *Both authors contributed equally to this work.

INTRODUCTION

Spinal muscular atrophy (SMA) is an autosomal recessive, neurodegenerative disorder typically resulting from biallelic deletions of the survival motor neuron 1 (*SMN1*) gene. Patients demonstrate a loss of motor neurons resulting in progressive skeletal muscle atrophy and weakness. SMA carrier frequency is 1 in 50 resulting in a disease prevalence of approximately 1 in 10,000 live births¹ making SMA the most common genetic cause of childhood death.

The contiguous *SMN1* paralog, *SMN2*, encodes amino acids that are identical to *SMN1*; however, an exonic point mutation in a putative exonic splice enhancer leads to the exclusion of exon 7 in about 90% of *SMN2* mRNA transcripts. As such, each *SMN2* copy only produces about 10% of the functional SMN protein ordinarily produced from a single, functional *SMN1* copy.²

Historically, SMA has been categorized into three clinical types based upon the onset of clinical symptoms and the maximum motor milestone achieved. SMA type I is characterized by symptom onset before 6 months of age and an inability to sit or stand independently.³ This severe infantile-onset form of SMA accounts for up to 60% of cases with a mean survival of 8-10-1/2 months of age.^{4,5} SMA type II shows symptom onset from 6 months to 18 months old with infants achieving the ability to sit but not walk independently. It has been estimated that 25% of children with SMA will have this form of the disease. SMA type III is characterized by symptom onset after 18 months old and children achieving the ability to walk at least some point during their lives.⁶ Rarer forms of SMA can include a congenital form (type 0) or a late, adult-onset form (type IV) with each accounting for <1% of all cases of this disease.^{6,7}

SMN2 copy number has some predictive value with respect to the type of SMA. For example, approximately 80% of SMA type I infants have two *SMN2* copies or less with a remainder having three *SMN2* copies.⁸ Children diagnosed with SMA type II are most likely to have three *SMN2* copies (80%) with approximately 16% having two *SMN2* copies and 4% having four *SMN2* copies. For SMA type III, 90% of children will have three or four *SMN2* copies.⁸ Individuals with adult-onset SMA type IV have four or more *SMN2* copies. Notwithstanding these associations, the *SMN2* genotype–phenotype correlation is insufficient to allow unequivocal *SMN2* genotype-based prediction of discrete SMA I, II, and III diagnoses.

Therapies have emerged for the treatment of SMA. In June 2017, nusinersen (Spinraza[®]) was approved by Health Canada and, at the time of writing, reimbursement criteria have been established in all but one Canadian province. An antisense oligonucleotide, nusinersen is administered at set intervals via intrathecal injection and binds to a segment of SMN2 pre-mRNA, altering its splicing and promoting the inclusion of exon 7.9 As a result, it augments the amount of full-length SMN protein produced enhancing survival of motor neurons in affected individuals with resulting significant clinical improvement. Nusinersen has been shown to be particularly effective when administered to SMA babies with two or three SMN2 copies before symptom onset.¹⁰ Children with four SMN2 copies have not been studied in these pivotal clinical trials. Additional treatments are emerging for SMA with a beneficial response reported from preliminary clinical trials of onasemnogene abeparvovec (Zolgensma[®]), a gene replacement therapy¹¹ as well as RO7034067 (Risdaplam[®]) a small molecule that also alters *SMN2* splicing increasing full-length *SMN2* mRNA production.¹² These treatments have received regulatory approval in other jurisdictions and will require review by regulatory bodies in Canada.

Newborn Screening Ontario (NSO), based at CHEO in Ottawa, began operations in 2006. Prior to this, screening in the province of Ontario was based out of the provincial public health laboratories and screening targets were limited to phenylketonuria and congenital hypothyroidism. NSO has expanded the provincial NBS panel to now include over 25 conditions and the resulting identification and treatment of over 2500 infants affected by one of these conditions. As outlined in a seminal 1968 WHO publication by Wilson and Jungner,¹³ "the object of screening for disease is to discover those among the apparently well who are in fact suffering from disease". Newborn screening (NBS) specifically is a public health population-based system, which involves testing all infants shortly after birth to identify those at risk for an increasing number of treatable conditions. which are not clinically evident in the newborn period.¹⁴ Wilson and Jungner also delineated several fundamental principles of screening, which have been interpreted over time as "criteria" to be used in the consideration of the appropriateness of a given disease for inclusion in a screening program. These principles/ criteria include characteristics of the disease itself (e.g. severity, knowledge of natural history), the screening test (e.g. test performance, robustness), the treatment (e.g. effectiveness, acceptability, and accessibility), and societal considerations (e.g. costeffectiveness, harms to those with false-positive result including the risk of overtreatment). The emergence of access to nusinersen as a transformative therapy for SMA, along with the availability of robust and accurate DBS screening tests, the detailed understanding of the natural history of the disease, and the availability of a system of care for screen-positive infants involving pediatric academic health science centers has made SMA particularly appealing for inclusion in NBS programs. Potential concerns about the inclusion of SMA as a target of NBS include the high cost of treatment, challenges in predicting the severity of disease in infancy (with the associated risk of overtreatment of infants with less severe forms of SMA), and equitable and timely access to care across an area as large and, in places, as sparsely populated as Ontario. The NSO Advisory Council reviewed the evidence and recommended that SMA be added to the provincial NBS program. However, in order to maximize benefits and minimize potential harms of SMA NBS, and ensure consistency across the province, it was recognized that a strong consensus was needed on the approach to SMA screening, clinical evaluation, and treatment. Therefore, a group of Ontario screening and pediatric neuromuscular experts met prior to the initiation of pilot screening to define which individuals should be reported as positive (i.e. SMN1 deletion with or without consideration of SMN2 copy number), and for whom, among these, immediate treatment versus careful follow-up should be recommended. Shortly thereafter, on January 13, 2020, an SMA pilot program was initiated at NSO. The SMA test was multiplexed with a recently established hearing impairment and severe combined immunodeficiency mutation screening test, allowing a transition with relative ease, no additional blood sample requirements, and minimal additional testing cost.

METHODS

Newborn Screening Testing Methodology

NBS dried blood samples (DBS) are collected on specially designed filter paper (also known as Guthrie paper) according to published criteria ideally from infants between 24 and 48 h of age. An insert outlining the SMA NBS pilot project was given to all parents with details on where to find further information on the NSO website.

The SMA NBS pilot in Ontario includes a laboratorydeveloped first-tier MassARRAY test for the presence of SMN1 and a second-tier multi-ligand probe amplification (MLPA) test for both SMN1 and SMN2 copy numbers (MRC Holland P021). The MassARRAY (Agena) test involves initial PCR amplification of the relevant SMN1 genomic region followed by annealing of primers overlapping or adjacent to sites of interest, with a single-base extension. SMA genotyping was added to a large multiplex MassARRAY assay assessing 22 mutations associated with early-onset hearing loss (GJB2/GJB6 and SLC26A4) as well as mutations within two genes causing severe combined immunodeficiency (SCID; IKBKB, and ZAP70) already being performed at NSO. Although many screening laboratories have multiplexed SMA PCR with TREC (SCID) qPCR,¹⁵ these tests had already been multiplexed with an assay for congenital CMV at NSO; additional targets in this qPCR assay were not possible. The first-tier screen performed at NSO, therefore, assesses for the presence of an SMN1 exon 7 single-nucleotide variant (SNV), and of exon 8 SMN1 and SMN2 SNVs. In traditional MassARRAY design, one primer would anneal to both SMN1 and SMN2, however, we noted during assay development that SMN2 copy number impacted SMN1 genotyping; specifically, individuals with \geq 5 copies of SMN2 were often incorrectly genotyped as SMN1 null. To address this issue, we altered our design to independently assess SMN1 exon 7 by using a primer whose 3' base is the C>T variant such that it will only anneal and produce a signal from SMN1. The exon 8 assay follows the traditional design where the primer sits adjacent to the exon 8 SNV and the signal is produced from both SMN1 and SMN2 (Figure 1). Given SMN1 and SMN2 dual null individuals are not viable, an exon 8 signal is anticipated in every live-born individual; if none is observed then a failed reaction is likely and the reaction is repeated. This method does not identify SMN1 +/- SMA carriers who would produce a signal indistinguishable from SMN1 + /+ individuals.

First-tier-positive samples (*SMN1* null) are immediately analyzed by MLPA (MRC Holland P021) for *SMN1* and *SMN2* copy numbers and if confirmed, a screen-positive report which includes *SMN2* copy number is issued if $\leq 4 \ge 3MN2$ copy numbers are detected. Samples with first-tier inconclusive results (exon 8 SMN1 null, exon 7 SMN1 present) are most likely due to high SMN2 copy number and thus analyzed by MLPA on a weekly basis.

Neuromuscular Disease Expert Consensus

In a series of teleconferences culminating in a 1-day face-toface meeting, Ontario-based Pediatric Neuromuscular disease experts reviewed and discussed the evidence, expert consensus statements, provincial and national treatment reimbursement guidelines, and clinical practice regarding diagnosis and treatment of children with SMA as it pertained to NBS. The definition of a screen-positive result (specifically as related to *SMN2* copy number) and details of a post-referral evaluation and management plan (including timelines) were then discussed and a post-referral evaluation algorithm was established.

RESULTS

Newborn Screening Evaluation Algorithm

NSO medical staff (PC) in consultation with Ontario Pediatric Neuromuscular experts (CC, JD, HG, AM, HM, and MT) created a post-referral evaluation algorithm (Figure 2) comprising the following key points:

It was decided that *SMN1*-null infants with four or fewer *SMN2* copies would be classified "screen positive". The group agreed that while the natural history of infants with 5x*SMN2* copies or more was not wholly predictable, adult-onset disease or potentially remaining completely asymptomatic throughout his or her life was the most likely outcomes. As such, reporting this condition when there is a chance that disease manifestation may not occur was deemed to be unethical and not in patients' best interest given the potential psychosocial impact, exclusion from insurability, and other potential ramifications associated with this disclosure.

The target time for the initial screen result was 7–10 days of age, acknowledging that samples may be taken at several days of age and/or shipped from remote sites within the province. All *SMN1*-null infants from the initial assay would undergo timely reflex MLPA testing, both confirming *SMN1* –/– genotype and delineating *SMN2* copy number.

SMN1 -/- infants with four SMN2 copies or less would be referred to a regional treatment center. A trained genetic counselor or nurse would contact the infant's family by telephone and they would either be directed to the closest pediatric hospital or have blood sent for confirmatory SMA genetic testing to be performed and to meet with a pediatric neuromuscular specialist to discuss the potential implications of the NSO test result. One tube of whole blood (EDTA, 3 ml) would also be sent at the same time to the NSO laboratory for quality assurance purposes. Confirmatory testing would be performed at either SickKids Hospital (Toronto) or the Children's Hospital of Eastern Ontario (Ottawa) aiming to return results within 7-10 days from receipt of the sample. One tube of whole blood (EDTA, 3 ml) would also be sent at the same time to the NSO laboratory for quality assurance purposes and rapid resolution of any discordant results (i.e. between the original screening results, diagnostic lab results, and second sample results at NSO). The rationale for the rapid investigation would be to determine if there was any evidence for the misidentification of specimens at the referring center or the receiving NBS laboratory to ensure there was no further delay in diagnosis and treatment initiation for another infant.

Following diagnostic confirmation, and determination of *SMN2* copy number, infants, and their families are assessed by a pediatric neuromuscular specialist at which time the family would have an opportunity to discuss treatment options and standard of care guidelines that are followed at all Ontario Pediatric Neuromuscular clinics.^{16,17} Baseline functional assessments (CHOP-INTEND, HINE) would be performed by a trained physiotherapist or clinical evaluator at or around that time. Establishing a definitive diagnosis between 16 and 27 days of age was determined to be feasible, and would allow initiation of

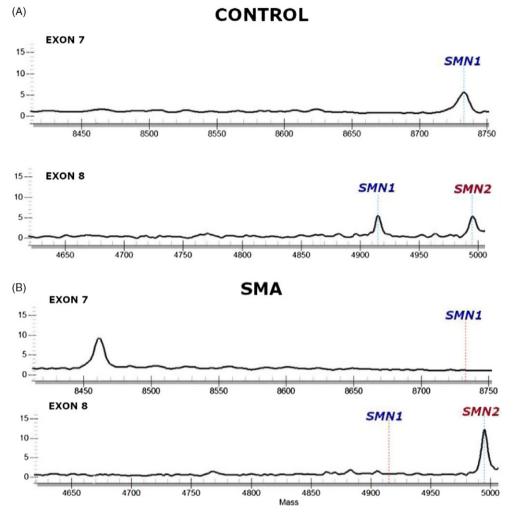


Figure 1: SMN1 and SMN2 MassARRAY chromatograms. (A) Infants not affected with SMA ("control") with one or two SMN1 copies will show a peak at SMN1 in both the exon 7 and exon 8 assays. (B) Infants affected with SMA (bottom) will not show SMN1 peaks in either exon 7 or exon 8 reflecting homozygous SMN1 deletion. The presence of SMN2 is seen just downstream (right) of where the SMN1 signal is seen (controls) or would have been expected (in affected infants), though this is not a target of the initial assay.

disease-modifying therapy in the majority of SMA patients with 2 or 3x*SMN*2 copies by 30 days of life.

Treatment and Surveillance Recommendations: 2 or 3 SMN2 Copies

All infants with two or three *SMN2* copies, given the evidence for rapid and irreversible loss of motor neurons, were recommended for immediate initiation of disease-modifying therapy prior to any clinical symptom onset (Figure 2). This recommendation is concordant with Ontario's Exceptional Access Program (EAP) reimbursement criteria for nusinersen.¹⁸ Application for access to nusinersen is to be placed with the family's private insurance (if applicable) and/or the EAP. Neurophysiological testing (i.e. nerve conduction studies, electromyography (EMG)) was not recommended for children in this cohort as it would not alter treatment decisions. Although extremely uncommon, *SMN1*-null infants with only 1 *SMN2* copy (i.e. predictive of SMA type 0) would be evaluated immediately. Given the potential severity of this congenital-onset form of SMA which could include the need for mechanical ventilation, the pediatric neuromuscular physician and family would discuss potential treatment options. Details of the recommended ongoing surveillance of patients with two or three copies of *SMN2* are outlined in Table 1.

Treatment and Surveillance Recommendations: 4x SMN2

Once confirmatory testing identifies four *SMN2* copies in an SMA infant, an assessment is to be conducted by a Pediatric Neuromuscular expert since a proportion of infants with four *SMN2* copies develop type I (<2%) or type II SMA (11%).⁸ Any clinical sign of SMA on neuromuscular examination (i.e. weakness, hypotonia, hyporeflexia, etc.) would prompt initiation of disease-modifying therapy. Motor nerve studies were recommended from the ulnar nerve (to abductor digiti minimi) and common peroneal nerve (to tibialis anterior). If the compound motor action potential (CMAP) was found to be <80% the lower-

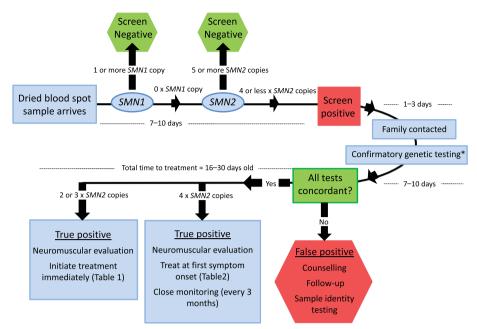


Figure 2: Post-referral evaluation algorithm.

All SMN1-null infants with four (or less) SMN2 copies will be reported. Patients' families will be contacted by a genetic counselor or nurse within 3 days who will relay the test result and arrange for confirmatory genetic testing and an urgent meeting with a pediatric neuromuscular specialist. *Denotes the need for confirmatory multi-ligand probe amplification (MLPA) to be sent to a clinical laboratory (either SickKids Hospital or the Children's Hospital of Eastern Ontario) as well as repeat sample to NSO laboratory. Patients with 2 or 3xSMN2 copies will receive immediate initiation of disease-modifying treatment. Patients with 4xSMN2 copies will be followed every 3 months by a pediatric neuromuscular specialist, with treatment initiation at the earliest sign of symptom onset. The goal for initiation of treatment is within the first 16–30 days of life.

Assessments:	Months of age													
	0	3	6	10	14	18	22	24	28	32	36	40	44	48
NMSK assessment	Х	X	X	Х	Х	X	Х	Х	X	X	X	X	X	Х
HINE	Х		X	X	X	X	X		Stop when HFMSE initiated					
CHOP-INTEND			X	X	X	X	X	Х	Continue if score <50					
HFMSE					Start CHOP-INTEND score ≥50			Х	X	X	X	X	X	Х
6MWT														X*
RULM														X*
Approach for CNDR enrollment	Х													
Treatment initiation	Х													

Table 1: Surveillance recommendations for infants with two or three SMN2 copies

X denotes the age (in months) at which each assessment is performed. NMSK = pediatric neuromuscular assessment. Treatment will be initiated at the first visit and administered at 16-30 days of life. The Hammersmith Infant Neurological Examination (HINE) will be discontinued when the HFMSE = Hammersmith Functional Motor Scale Extended is initiated. CHOP-INTEND scoring will be discontinued at age 24 months old unless the score is <50 (maximum CHOP-INTEND score is 64). X* denotes that 6-min walk test (6MWT) and Revised Upper Limb Module (RULM) will be at 4 years of age if the child is developmentally capable of cooperating with this test.

limit for age or if needle EMG noted any sign of denervation this would also confirm neurophysiological evidence of disease onset and prompt treatment initiation. If no clinical or neurophysiological evidence of disease was noted, it was recommended that treatment not be initiated and the child be seen every 3 months until 12 months of age. Surveillance physical examinations would be performed by a pediatric neuromuscular specialist and validated functional assessments (CHOP-INTEND, HINE) would be

508

Assessments:	Months of age											
	0	3	6	9	12	18	24	36	48	60	72	
NMSK assessment	Х	X	Х	X	X	X	X	Х	Х	х	X	
HINE	Х	Х	Х	X	X	Х	Stop when HFMSE initiated					
CHOP-INTEND	Х	Х	Х	X	X	Х	Continue if score <50					
HFMSE							Х	X	Х	Х	X	
6MWT									X*	X*	X*	
RULM									X*	X*	X*	
СМАР	Х	Repeat if clinically indicated										
EMG	Х	X Repeat if clinically indicated										
Approach for CNDR enrollment												
Treatment initiation	X [#] ; initiate treatment if clinical or neurophysiological signs of disease appear											

Table 2: Surveillance recommendations for infants with four SMN2 copies

X denotes the age (in months) at which each assessment is performed. NMSK = pediatric neuromuscular assessment. Infants with 4xSMN2 copies who show clinical or neurophysiological signs of disease onset (i.e. compound muscle action potential (CMAP) amplitudes <80% lower limit of normal and/or needle electromyography (EMG) demonstrating an evidence of denervation) will prompt initiation of treatment (denoted by $X^{\#}$). Declining motor function scores will prompt initiation of treatment. Infants not showing evidence of disease onset will have a repeat examination and motor function scoring every 3 months (until 12 months old) where after they will be seen at 18 months old, 24 months old, and then annually. The Hammersmith Infant Neurological Examination (HINE) will be performed every 3 months until the HFMSE = Hammersmith Functional Motor Scale Extended is initiated. CHOP-INTEND scoring will be discontinued at age 24 months old unless the score is <50 (maximum CHOP-INTEND score is 64). X* denotes that 6-min walk test (6MWT) and Revised Upper Limb Module (RULM) will be performed beginning at 4 years of age if the child is developmentally capable of cooperating with this test. CMAP or EMG will be repeated only if needed to guide decision-making regarding treatment initiation.

completed by a trained clinical evaluator until at least 2 years of age. Asymptomatic children with four *SMN2* copies beyond 2 years of age are to be regularly assessed with the Hammersmith Functional Motor Scale Extended (HFMSE). The 6-min walk test (6MWT) and Revised Upper Limb Module (RULM) would be initiated at 4 years of age if the child was deemed developmentally capable of cooperating with this test. The complete recommended schedule of assessment is summarized in Table 2. Treatment would be recommended at the earliest clinical or electrophysiological sign of disease symptoms in this cohort.

Limitations of Testing

As with all other SMA NBS approaches currently in use, our screening platform will only identify patients with an *SMN1* gene deletion or conversion, but not those with other pathogenic variations in *SMN1* such as point mutations or small deletions. This is predicted to miss 3-5% of children who may have point mutations on one or both alleles. As such, a level of clinical vigilance must be maintained when seeing screen-negative infants and children with an SMA phenotype including, when indicated, neurophysiological testing (i.e. EMG) and/or confirmatory Sanger sequencing.

DISCUSSION

The remarkable progress in SMA therapies, along with advances in NBS technology, has made the disease an excellent candidate for inclusion in screening programs. It is clear that early initiation of treatment, ideally in pre-symptomatic or minimally

Volume 48, No. 4 - July 2021

https://doi.org/10.1017/cjn.2020.229 Published online by Cambridge University Press

symptomatic SMA infants offers the best chance of optimizing motor function, reducing complications such as respiratory insufficiency requiring ventilation as well as indefinitely prolonging survival.¹⁰ In keeping with this, many countries and jurisdictions have adopted NBS for SMA. Following the inclusion of SMA in the American Recommended Uniform Screening Panel (RUSP) in July 2018, 37 individual states have adopted NBS for SMA with 23 of those states having implemented full screening programs as of May 2020.¹⁹ An additional three states have undertaken pilot screening programs.¹⁹

The ability to predict natural history in asymptomatic patients diagnosed during screening is vital to make rational treatment decisions and appropriately counsel families. While the overlap in observed disease severity between those with various SMN2 copy numbers means it is an imperfect predictor for this purpose, it is nevertheless an important parameter in guiding such decisions. We recommend treating all babies with two or three SMN2 copies, in alignment with provincial drug reimbursement guidelines. The recommended deferral of treatment of infants with four SMN2 copies pending clinical or electrophysiological evidence of disease onset follows provincial drug reimbursement guidelines as well as the approach used in a recent German pilot^{20,21} and previously published American treatment guidelines,²² but diverges from a revision of the latter guideline which recommends early treatment.²³ More recently, an NBS pilot program in New South Wales and the Australian Capital Territory (NSW/ ACT) did not report patients with four or more SMN2 copy patients.²⁴ In this regard, our recommendation to not treat all four SMN2 copy SMA patients was grounded in the observation that there is heterogeneity with regard to the age of symptom onset. Although patients may show symptom onset at around 3 years of age, a subset may have much later onset, not occurring until midadulthood (mean 37 years old; range 30–43 years old).²⁵ This as well as the paucity of clear evidence for the efficacy of treatment in the four *SMN2* copy groups was the basis for the decision for careful surveillance in this cohort. It is also very important to note that this decision was also guided by core screening principles including access to treatment within the system of care, avoid-ance of harm from overtreatment, and cost-effectiveness considerations.

The rapid initiation of treatment is essential, particularly for pre-symptomatic, 2xSMN2 copy patients who show a rapid decline in neurophysiological markers (e.g. CMAP amplitudes) shortly after birth.²⁶ Over 40% of SMA patients identified in an NBS program show clear evidence of clinical symptoms within the first few weeks of life.²⁴ We have established a target range of 16-30 days of age (Figure 2) within which to initiate treatment, similar to the NSW/ACT program which started diseasemodifying treatment at a median of 26.5 days of life (range: 16-37 days).²⁴ Optimally, by decreasing the time for confirmatory genetic testing and/or reducing time for EAP treatment approval, we hope to initiate treatment before 21 days of life. Currently, EAP approval cannot proceed until confirmatory genetic testing has been obtained. The demonstration of a strong concordance between NBS and confirmatory genetic test results could potentially allow a process of conditional EAP approval pending confirmatory testing results.

The profound alteration of SMA natural history observed following early treatment alters the historical concept of SMA "typing" based upon the age of symptom onset and the highest motor milestone achieved. There is strong evidence that initiation of therapy in the pre- or early symptomatic phases of disease is vital to optimizing outcomes for children with early-onset SMA.^{10,11} With NBS enabling early diagnoses and access to highly effective therapies, children with SMA should survive and achieve motor milestones that would not have previously been possible. While SMN2 copy number has been used as a surrogate for SMA type, the overlap between copy number and SMA type means that the evaluation of the effectiveness of treatment in individual patients cannot solely be based on copy number alone as it lacks sensitivity and specificity. Other genetic modifiers alter the severity of the SMA phenotype including the rare SMN2 c.859G>C mutation, which is associated with higher SMN2 protein levels.²⁷ Actin-binding protein plastin 3 (PLS3), neurocalcin delta (NCALD), and neuronal apoptosis inhibitory protein (NAIP) have been reported to have possible disease-modifying effects.^{28–30} Given that there is currently insufficient evidence pertaining to the population prevalence, strength of association, and effect upon the phenotype for these genetic modifiers, testing was not incorporated into the screening algorithm. As evidence emerges, it will be necessary to consider including such testing, which may be particularly valuable for individuals with 4xSMN2 copies since they have the potential for considerable phenotypic heterogeneity. Additional means to predict natural history of disease will be needed both to guide individual patient treatment decisions, as well as for a better understanding of the effectiveness of NBS for SMA in changing outcomes.

Ongoing assessment of this program will be necessary, as will review of the current algorithms as further evidence, drug reimbursement guidelines, and clinical consensus statements emerge. Ultimately, as a better understanding of "4x*SMN2* copy" SMA disease onset emerges from the NBS-based postnatal ascertainment of affected infants, a better sense of treatment timing will also be delineated. The high levels of SMN observed in both control fetal and postnatal spinal cord tissue drop dramatically so that by 3 months of age it is indistinguishable from levels seen in SMA postmortem tissue.³¹ Thus, judiciously timed reduced doses in the first 6 months may afford long-term benefit, forestalling, and possibly even preventing disease onset for SMA patients with four copies of *SMN2*. In keeping with this, the elegant study of inducible SMA mouse models has shown minimal SMN requirement following maturation of the neuromuscular junction.³² In addition, new biomarkers are under investigation which may enable further precision related to symptom onset and need for and response to therapy.

In the end, clinical trials of such treatment approaches in the subset of SMA patients mostly at risk for late-onset disease will be needed to support individual patient treatment decisions, drug reimbursement policies, and screening approaches and policies. As multiple disease-modifying therapies emerge, there will be a need to evaluate the efficacy and tolerability of treatments at different stages of the disease. As Wilson and Jungner elegantly described in their treatise, and as illustrated in our recommended approach, screening decisions and programs must always balance individual and population benefits and harms, taking into account disease, treatment, test, and social considerations. Ultimately, our hope is that this can provide useful information for Ontario physicians regarding the current landscape of NBS in the province as well as for other provinces that may be making decisions about screening programs of their own.

ACKNOWLEDGMENTS

Biogen provided funding to AMK (Principal Investigator) to allow Newborn Screening Ontario (NSO) to carry out this pilot study.

DISCLOSURES

KDK, EY, KA, JM, MAT, JV, and PC report no disclosures relevant to the manuscript. HJM has been a member of an AveXis Advisory Board and was a site investigator for an SMA clinical trial (AveXis). JB has been a member of Advisory Boards (Biogen, Novartis). CC has been a site investigator for SMA clinical trials (Biogen, Roche). JJD has been a consultant for a company developing therapies for rare diseases. HG has been a member of Advisory Boards (Biogen, Roche). AMK was the principal investigator and received research funding for the pilot newborn screening project (Biogen).

AUTHORS CONTRIBUTIONS

HJM, KDK drafted the manuscript. AMK, PC assisted with the writing of the manuscript. EY, KA, JJD, CC, JD, HG, JM, MAT, JV, AMK, and PC provided critical review and edited the manuscript. All authors approve of the manuscript in its current form.

ABBREVIATIONS

CHEO = Children's Hospital of Eastern Ontario;

CHOP-INTEND = Children's Hospital of Philadelphia Infant Test of Neuromuscular Disorders;

CMAP = compound motor action potential;

CMV = cytomegalovirus;

DBS = dried blood spot;

HFMSE = Hammersmith Functional Motor Scale Extended;

HINE = Hammersmith Infant Neurological Examination;

mRNA = messenger ribonucleic acid;

MLPA = multi-ligand probe amplification;

NAIP = neuronal apoptosis inhibitory protein;

NBS = Newborn Screening;

NSO = Newborn Screening Ontario;

NSW/ACT = New South Wales and Australian Capital Territory;

PCR = polymerase chain reaction;

SCID = severe combined immunodeficiency;

SMA = spinal muscular atrophy;

SMN1 = survival motor neuron 1;

SMN2 = survival motor neuron 2;

SNV = single-nucleotide variant;

WHO = World Health Organization.

References

- Verhaart IEC, Robertson A, Wilson IJ et al. Prevalence, incidence and carrier frequency of 5q-linked spinal muscular atrophy – a literature review. Orphanet J Rare Dis. 2017;12(1):124.
- Burghes AHM, Beattie CE. Spinal muscular atrophy: Why do low levels of survival motor neuron protein make motor neurons sick? Nat Rev Neurosci. 2009;10(8):597–609.
- 3. Markowitz JA, Singh P, Darras BT. Spinal muscular atrophy: a clinical and research update. Pediatr Neurol. 2012;46(1):1–12.
- Finkel RS, McDermott MP, Kaufmann P, et al. Observational study of spinal muscular atrophy type I and implications for clinical trials. Neurology. 2014;83(9):810–17.
- Kolb SJ, Coffey CS, Yankey JW, et al. Natural history of infantileonset spinal muscular atrophy. Ann Neurol. 2017;82(6):883–91.
- Wadman RI, Wijngaarde CA, Stam M, et al. Muscle strength and motor function throughout life in a cross-sectional cohort of 180 patients with spinal muscular atrophy types 1c-4. Eur J Neurol. 2018;25(3):512–18.
- Juntas Morales R, Pageot N, Taieb G, Camu W. Adult-onset spinal muscular atrophy: an update. Rev Neurol (Paris). 2017;173(5):308–19.
- Calucho M, Bernal S, Alías L, et al. Correlation between SMA type and *SMN2* copy number revisited: An analysis of 625 unrelated Spanish patients and a compilation of 2834 reported cases. Neuromuscul Disord. 2018;28(3):208–15.
- 9. Corey DR. Nusinersen, an antisense oligonucleotide drug for spinal muscular atrophy. Nat Neurosci. 2017;20(4):497–99.
- De Vivo DC, Bertini E, Swoboda KJ, et al. Nusinersen initiated in infants during presymptomatic stage of spinal muscular atrophy: interim efficacy and safety results from the Phase 2 NURTURE study. Neuromuscul Disord 2019;29(11):842–56.
- Mendell JR, Al-Zaidy S, Shell R, et al. Single-dose gene replacement therapy for spinal muscular atrophy. N Engl J Med. 2017;377(18):1713–22.
- Sturm S, Günther A, Jaber B, et al. A phase 1 healthy male volunteer single escalating dose study of the pharmacokinetics and pharmacodynamics of risdiplam (RG7916, RO7034067), a SMN2 splicing modifier. Br J Clin Pharmacol. 2019;85(1):181–93.
- Wilson JMG, Jungner G. (1968) Principles and practice of screening for disease. Geneva: World Health Organization. Available at:

https://apps.who.int/iris/bitstream/handle/10665/37650/WHO_PHP_34.pdf?sequence=17 [cited 27 Aug 2020]

- Zavon MR. Principles and Practice of Screening for Disease. Arch Intern Med. 1969;123(3):349.
- Taylor JL, Lee FK, Yazdanpanah GK, et al. Newborn blood spot screening test using multiplexed real-time PCR to simultaneously screen for spinal muscular atrophy and severe combined immunodeficiency. Clin Chem. 2015;61(2):412–19.
- Mercuri E, Finkel RS, Muntoni F et al. Diagnosis and management of spinal muscular atrophy: part 1: recommendations for diagnosis, rehabilitation, orthopedic and nutritional care. Neuromuscul Disord. 2018;28(2):103–15.
- Finkel RS, Mercuri E, Meyer OH et al. Diagnosis and management of spinal muscular atrophy: part 2: pulmonary and acute care; medications; supplements and immunizations; other organ sstems and ethics. Neuromuscul Disord. 2018;28(3):197–207.
- Exceptional Access Program Reimbursement Criteria for Frequently Requested Drugs. Available at: http://www.health.gov.on.ca/en/ pro/programs/drugs/docs/frequently_requested_drugs.pdf [cited 23 May 2020].
- Newborn Screening for SMA in America. Available at: https:// curesma.org/newborn-screening-for-sma/ [cited 26 May 2020]
- Vill K, Kölbel H, Schwartz O, et al. One year of newborn screening for SMA – Results of a German pilot project. J Neuromuscul Dis. 2019;6(4):503–15.
- Müller-Felber W, Vill K, Schwartz O, et al. Infants diagnosed with spinal muscular atrophy and 4 *SMN2* copies through newborn screening - opportunity or burden? J Neuromuscul Dis. 2020;7(2):109–17.
- Glascock J, Sampson J, Haidet-Phillips AH, et al. Treatment algorithm for infants diagnosed with spinal muscular atrophy through newborn screening. J Neuromuscul Dis. 2018;5:145–58.
- Glascock J, Sampson J, Connolly AM, et al. Revised recommendations for the treatment of infants diagnosed with spinal muscular atrophy via newborn screening who have 4 copies of *SMN2*. J Neuromuscul Dis. 2020;7:97–100.
- Kariyawasam DST, Russell JS, Wiley V, et al. The implementation of newborn screening for spinal muscular atrophy: the Australian experience. Genet Med. 2020;22(3):557–65.
- Wadman RI, Stam M, Gizen M, et al. Association of motor milestones, *SMN2* copy number and outcome in spinal muscular atrophy types 0-4. J Neurol Neurosurg Psychiatry. 2017;88(4):365–67.
- Swoboda KJ, Prior TW, Scott CB, et al. Natural history of denervation in SMA: relation to age, SMN2 copy number, and function. Ann Neurol. 2005;57(5):704–12.
- Vezain M, Saugier-Veber P, Goina E, et al. A rare SMN2 variant in a previously unrecognized composite splicing regulatory element induces exon 7 inclusion and reduces the clinical severity of spinal muscular atrophy. Hum Mut. 2010;31(1): E1110-25.
- Yanyan C, Yujin Q, Jinli B, et al. Correlation of PLS3 expression with the disease severity in children with spinal muscular atrophy. J Hum Genet. 2014;59(1):24–27.
- Riessland M, Kaczmarek A, Schneider S, et al. Neurocalcin delta suppression protects against spinal muscular atrophy in humans and across species by restoring impaired endocytosis. Am J Hum Genet. 2017;100(2):297–315.
- Qu Y, Ge XS, Bai J, et al. Association of copy numbers of survival motor neuron gene 2 and neuronal apoptosis inhibitory gene protein with the natural history in a Chinese spinal muscular atrophy cohort. J Child Neurol. 2015;30(4):429–36.
- Ramos DM, d'Ydewalle C, Gabbeta V, et al. Age-dependent SMN expression in disease-relevant tissue and implications for SMA treatment. J Clin Invest. 2019;129(11):4817–31.
- Kariya S, Obis T, Garone C, et al. Requirement of enhanced Survival motoneuron protein imposed during neuromuscular junction maturation. J Clin Invest. 2014;124(2):785–800.