Recent advances in the nutritional biochemistry of trivalent chromium

John B. Vincent

Department of Chemistry and Coalition for Biomolecular Products, The University of Alabama, Tuscaloosa, AL 35487-0336, USA

The nutritional biochemistry of trivalent Cr has been a poorly understood field of study; investigations of the biochemistry of the other essential transition metals have not proven as problematic. Despite over four decades of endeavour, only recently has a picture of the role of Cr potentially started to be defined. The biologically-relevant form is the trivalent ion. Cr^{3+} appears to be required for proper carbohydrate and lipid metabolism in mammals, although fortunately Cr deficiency is difficult to achieve. Conditions that increase circulating glucose and insulin concentrations increase urinary Cr output. Cr is probably excreted in the form of the oligopeptide chromodulin. Chromodulin may be the key to understanding the role of Cr at a molecular level, as the molecule has been found to bind to activated insulin receptor, stimulating its kinase activity. A mechanism for the action of chromodulin has recently been proposed; this mechanism can serve as a potential framework for further studies to test the role of Cr in metabolism. An examination of the nutritional supplement chromium picolinate illustrates some of the difficulties associated with these biochemical studies.

Chromium: Chromodulin: Chromium picolinate

In the last decade Cr has become amazingly popular as a nutritional supplement, weight-loss agent and muscledevelopment agent. Among mineral supplements products containing Cr are second in sales only to Ca-containing products (Nielsen, 1996). However, this popularity is not reflected in the level of understanding of how Cr functions in the body or even of whether the element is essential. With one exception, the first-row transition elements from V to Zn (and additionally the heavier transition elements Mo and W) have been shown to be essential for at least one form of life. Additionally, a biomolecule containing each of these metals, except one, has been crystallized and its three-dimensional structure determined. The exception is Cr.

While it is generally accepted that Cr is an essential element, the evidence is strongly supportive, but not definitive (Vincent, 2001). Four types of evidence have been presented to support the role for Cr as a trace nutrient for mammals: (1) five patients on total parenteral nutrition (before supplemental Cr was added to total parenteral nutrition solutions) developed symptoms of adult-onset diabetes that were reversed by the addition of Cr to the total parenteral nutrition solution (Jeejeebhoy, 1999); (2) rats fed a low-Cr sucrose-based diet have increased areas under the curve for insulin in glucose tolerance tests, suggesting the development of insulin resistance (Striffler *et al.* 1995, 1999); (3) Cr absorption is inversely proportional to dietary intake in human subjects (Anderson & Kozlovsky, 1985); (4) increases in serum glucose are accompanied by increases in urinary Cr excretion, while conditions that alter glucose metabolism (including pregnancy, type 2 diabetes and other metabolic stresses) are associated with alterations in urinary Cr output (Kozlovsky *et al.* 1986; Anderson *et al.* 1990; Morris *et al.* 1993). These associations suggest a relationship between normal glucose metabolism and Cr, probably associated with insulin action.

However, each of these sets of evidence is problematic. For example, Cr absorption in rats (Anderson & Polansky, 1995), in contrast to human subjects, is not inversely proportional to intake. Increased urinary excretion of Cr may only reflect the effects of changes in the levels of Fe mobilization in response to insulin. Incidences of diagnosed potential Cr deficiency in human subjects are limited to five cases that lack consistent relationships between the Cr in the total parenteral nutrition, time on total parenteral nutrition before symptoms, serum Cr levels and symptoms

Abbreviation: Cr(pic)₃, chromium picolinate.

Corresponding author: Professor John B. Vincent, fax +1 205 348 9203, email jvincent@bama.ua.edu

(Stearns, 2000). For these reasons the reversal of symptoms associated with Cr supplementation is the only generally accepted indicator of Cr deficiency. A biomarker of Cr status is urgently needed. Studies undertaken before 1990 that examined the effects of Cr-deficient diets are flawed by methodological concerns (Vincent, 2001).

Fortunately, the data suggest that generating Cr deficiency in human subjects is extremely difficult. The dietary guidelines for Cr intake recommended by the Food and Nutrition Board of the US National Academy of Sciences have been lowered from $50-200 \,\mu\text{g/d}$ for an adult to $35 \mu g/d$ for an adult male and $25 \mu g/d$ for an adult female (Trumbo et al. 2001). The new guideline values for Cr are adequate intakes, a recommended intake based on the intakes of groups of healthy individuals that are assumed to be adequate when there is sufficient data to establish a recommended daily allowance. Thus, these limits are set at the average intake for Americans (for data on average intake and Cr balance, see Bunker et al. 1984; Gibson & Scythes, 1984; Anderson & Kozlovsky, 1985; Offenbacher et al. 1986; Anderson et al. 1993), indicating that few Americans should be Cr deficient.

Chromium picolinate

Although the first studies that suggested that chromium picolinate (Cr(pic)₃) could have beneficial effects on body mass and composition were only published in 1989 (Evans, 1989), the nutritional supplement has become amazingly popular. For example, products containing the supplement generated approximately US 0.5×10^9 in sales in 1999 (Mirasol, 2000). However, studies of $Cr(pic)_3$ in healthy individuals conducted since the initial reports have failed to support the early findings (Hellerstein, 1998). A recent review has examined human studies investigating the effects of Cr(pic)₃ on body composition and concluded that 'the supplement has no demonstrated effects on healthy individuals, even when taken in combination with an exercise program' (Vincent, 2003). Other researchers have come to similar conclusions (Clarkson, 1997; Kreider, 1999; Lukaski, 1999), as have recent meta-analyses (Nissen & Sharp, 2003; Pittler et al. 2003). There have also been claims that suggest that $Cr(pic)_3$ has beneficial effects on plasma glucose and insulin concentrations and other blood variables in healthy subjects. These claims have not been substantiated, as shown by another recent review (Vincent, 2001) and meta-analysis (Althuis et al. 2002).

Only 6 years after the initial report of potentially beneficial effects from $Cr(pic)_3$ supplementation, Wetterhahn and coworkers (Stearns *et al.* 1995) reported the first chemical evidence for concerns over the use of $Cr(pic)_3$. The complex generated chromosome damage in a Chinese hamster ovary cell model. Subsequently, damage was demonstrated in murine macrophages (Bagchi *et al.* 1997), and another study using the same cell line observed oxidative damage associated with $Cr(pic)_3$ (Bagchi *et al.* 2002). Stearns and colleagues (Stearns *et al.* 1995; Manygoats *et al.* 2002) in continuing work with the Chinese hamster ovary model have observed mitochondrial

damage and apoptosis generated by the supplement (Manygoats et al. 2002) and have found that the supplement is mutagenic (Stearns et al. 2002). The effects have been postulated to arise from the released picolinate ligand (Stearns et al. 1995; Manygoats et al. 2002) or from reactive oxygen species catalytically generated by the intact complex (Speetjens et al. 1999a; Sun et al. 2000). Physiologically-relevant concentrations of Cr as Cr(pic)₃ (e.g. 120 nm) and of biological reducing agents such as ascorbic acid and thiols have been shown by the author's group to result in catalytic production of reactive oxygen species, which cleaved DNA (Spectiens et al. 1999a; Sun et al. 2000). Most forms of Cr do not generate such species in the absence of a strong oxidant such as peroxide. Hence, these studies are consistent with investigations that demonstrated that mutagenic forms of trivalent Cr possessed chelating ligands containing pyridine-type N or other imine-N (e.g. 2,2'-bipyridine, phenanthroline and Schiff bases) coordinated to the metal and that they generated reactive oxygen species (Sugden et al. 1992). Cr compounds that do not have imine ligands lack the DNA cleaving activity in the presence of biological reducing agents (Speetjens et al. 1999b). Alternatively, neutral Cr(pic)₃ could serve as a vehicle for the transport of picolinate to cells. Consequently, for these mechanisms to lead to marked cell damage Cr(pic)₃ needs to enter cells intact and remain intact long enough to produce a substantial quantity of reactive oxygen species or then degrade releasing picolinate. Recently, Vincent and coworkers (Hepburn & Vincent, 2002, 2003) have shown that $Cr(pic)_3$ is able to pass rapidly from the bloodstream and enter cells intact, although the lifetime of the complex in cells is short. In these studies the appearance and distribution of Cr in tissues after intravenous injections of $Cr(pic)_3$ are very similar to those found when $Cr(pic)_3$ is administered orally (Olin et al. 1994). In hepatocytes Cr from Cr(pic)₃ first appears in the nucleus and mitochondria, then the cytosol, and finally the lysosomes and microsomes; however, the complex has no propensity to bind to DNA. Kelley and coworkers (Kareus et al. 2001) have found that hepatocyte microsomes can catalytically modify the picolinate ligands, which would result in Cr release. While Cr from administration of the supplement can accumulate in the kidneys and liver (Anderson et al. 1997*a*), it does not accumulate as $Cr(pic)_3$ (Hepburn & Vincent, 2002, 2003).

Isolated incidents of deleterious effects of $Cr(pic)_3$ supplementation have been reported (Huszonek, 1993; Wasser & D'Agati, 1997; Cerulli *et al.* 1998; Martin & Fuller, 1998; Fowler, 2000). The nature of these incidents makes their importance difficult to ascertain. In contrast, no acute toxic effects were observed in rats fed diets containing ≤ 100 mg Cr as $Cr(pic)_3/kg$ diet for 24 weeks (Anderson *et al.* 1997*a*). However, the potential effects of oxidative damage were not investigated. No toxic effects of $Cr(pic)_3$ supplementation were noted in any of the studies covered by the review articles mentioned previously, which in total monitored hundreds of subjects (Vincent, 2001, 2003). There have not yet been any studies that have examined the effects, positive or negative, of long-term (>1 year) use of $Cr(pic)_3$.

However, human and animal studies that have looked for DNA damage and oxidative damage have started to be published. The level of 5-hydromethyluracil in the blood serum is a marker of the extent of oxidation of the DNA base thymine; after oxidation, the modified base is enzymically removed and subsequently appears in the serum. The repair of this damage has a considerable error rate, giving rise to mutations. There was no observed effect on 5-hydroxymethyluracil levels in ten obese women given $400 \,\mu\text{g}$ Cr as Cr(pic)₃/d for 8 weeks (Kato *et al.* 1998). In contrast, Vincent and coworkers (Hepburn et al. 2003a) found that rats given an intravenous injection of $Cr(pic)_3$ daily for 60 d had elevated levels of urinary 8-hydroxydeoxyguanosine and elevated levels of peroxidized lipids. Both direct DNA oxidation and indirect DNA damage via lipid peroxidation resulting from $Cr(pic)_3$ administration provide potential pathways for the chromosome damage (Stearns et al. 1995) and, more recently, the mutations (Stearns et al. 2002) seen in cell culture studies. The quantities of Cr used in this study were high, but establish that such damage is possible in vivo. Also, a recent preliminary report (Mahboob et al. 2002) has indicated that Cr(pic)₃ causes oxidative damage when given orally to rats in quantities equivalent to the intakes of human subjects taking commercial supplements. Potentially-deleterious in vivo effects of Cr(pic)₃ have been examined recently by O'Donnell and Vincent and coworkers using Drosophila melanogaster (Hepburn et al. 2003b). $Cr(pic)_3$, but not $CrCl_3$, at $\leq 260 \,\mu g \, Cr/kg$ food (approximately equivalent to that received by a human subject taking daily Cr(pic)₃ supplement containing 200 µg Cr) was found to lower the success rate of pupation and eclosion and to arrest development of pupae in a concentration-dependent manner. X-linked lethal analysis has indicated that the supplement greatly enhances the rate of appearance of lethal mutations and dominant female sterility. Subsequently, Vincent and coworkers (DDD Hepburn and JB Vincent, unpublished results) examined polytene chromosome arms of nuclei from cells of salivary glands from third instar larval Drosophila; these larvae were the progeny of male and female Drosophila maintained on Cr(pic)₃-containing food (260 µg Cr/kg food). Although the progeny were never exposed to the supplement, >50% of the chromosome arms had chromosomal aberrations and rearrangements. In contrast, no aberrations or rearrangements were observed in chromosome arms of progeny from adults on food without the supplement.

In March 2003 the Expert Group on Vitamins and Minerals of the UK Joint Food Standards and Safety Group requested that the health supplement industry should voluntarily withdraw $Cr(pic)_3$ -containing products, while also consulting on a ban on the use and sale of $Cr(pic)_3$ in the UK. Currently, the US Food and Drug Administration, working with the US National Academy of Sciences, is studying the potential regulation of $Cr(pic)_3$.

Chromium and type 2 diabetes

While Cr supplementation of the diet of healthy individuals appears to have no statistically significant effects, the situation with supplementation of subjects with type 2 diabetes may be very different. As Cr is probably an essential trace element, supplementary levels of Cr would be expected to have little if any effect on variables such as body mass or body composition if subjects consume Cr-sufficient diets. However, the administration of pharmacological amounts of Cr could result in effects that potentially could lead to altered Cr status, especially for subjects with altered metabolisms. Type 2 diabetes (Morris et al. 1999) and pregnancy (Morris et al. 1995) are examples of conditions that lead to increased urinary Cr loss, which with time could result in a decrease in Cr status, although this outcome has not been proven. Thus, individuals with these conditions could potentially benefit from Cr supplementation. Additionally, because these individuals have lowered insulin sensitivity and could potentially benefit from increased Cr loading of chromodulin (thought to be the naturally-occurring biologically-active form of Cr; see p. 44), resulting in increased insulin signalling, administration of pharmacological amounts of Cr could be beneficial. Similarly, a therapeutic agent (such as the trinuclear biomimetic complex; see p. 45) that mimics chromodulin's action could be particularly useful in treating their insulin insensitivity.

The effects of Cr in subjects with type 2 diabetes are less certain than those in healthy subjects (in whom no significant effects are observed). A meta-analysis of studies involving diabetic subjects revealed that 'A study of 155 diabetic subjects ... showed that chromium reduced glucose and insulin concentrations; the combined data from ... the other studies did not' (Althuis et al. 2002). The placebo-controlled study of Anderson et al. (1997b), which involved 155 subjects in China, is the largest reported study of diabetic subjects. The subjects received 0, 200 or 1000 µg Cr daily for 4 months. At the higher levels of Cr subjects had reduced fasting serum glucose, insulin and total cholesterol, and lower 2h insulin and glucose concentrations after a glucose challenge. Subjects also possessed lower glycated Hb A1 levels. Follow-up studies in China (Cheng et al. 1999) yielded results in accord with the initial study. The original study has been analysed in detail by Hellerstein (1998). Before 2002 the consensus from double-blind placebo-controlled studies of the effects of Cr supplementation for 6-16 weeks in subjects with type 2 diabetes was that supplementation had no effect (Vincent, 2001). In all these studies $\leq 200 \,\mu g$ Cr was administered daily. However, a recent double-blind crossover study in India of subjects with type 2 diabetes indicated that Cr supplementation (400 µg Cr/d) for 12 weeks lowered serum insulin and glucose levels (Ghosh et al. 2002). Anderson (1998, 2000) has reviewed studies of the effects of Cr supplementation in subjects with type 2 diabetes; generally, only studies using <200 µg Cr daily reported no effects from supplementation, leading the reviewer to postulate that larger quantities of Cr can have beneficial effects in subjects with type 2 diabetes. Thus, positive effects from Cr supplementation of subjects with type 2 diabetes potentially have only been seen in very recent studies (after 1996) that utilized large pharmacologically-relevant quantities of Cr.

Importantly, the findings of the studies using $>200 \,\mu g$ Cr/d are supported by the findings of studies that used

model rats. A trinuclear biomimetic complex (see p. 45) was found to have beneficial effects on Zucker obese rats, models for the early stages of type 2 diabetes (Sun *et al.* 1999, 2002), while smaller, but significant ($P \le 0.05$), effects were also noted in healthy rats. Cefalu *et al.* (2002) observed beneficial effects of Cr(pic)₃ administration (18 µg Cr/kg body mass daily, equivalent to 540 µg for a 60 kg subject) on insulin sensitivity in a rat model for type 2 diabetes and CVD. Thus, the continuation of rat and human studies is required in order to further elucidate the potential use of Cr administration as an adjuvant therapy for type 2 diabetes.

Additionally, one study of the effects of Cr on gestational diabetes has been reported (Jovanovic et al. 1999). In women (average body mass 82-84 kg) who received 0, 4 or 8µg Cr/kg body mass daily it was found that both Cr-supplemented groups had significantly ($P \le 0.05$) lower plasma insulin and glucose levels. If the findings of this single study are confirmed by additional studies, this result would have important implications for the treatment of this condition. Recently, studies of the effects of Cr supplements on steroid-induced diabetes have generated interesting results (Ravina et al. 1999a,b), including improvements in plasma glucose levels. Although these studies still need to be followed up by larger investigations, the findings are supported by those of a recent study of rats treated with dexamethasone (Kim et al. 2002). The dexamethasone-treated rats receiving approximately 4 mg Cr daily had lower fasting serum insulin levels and lower insulin, triacylglycerol and glucose areas under the curve in glucose or insulin challenges.

A mechanism for the action of chromium and an active chromium-containing biomolecule

Investigations over the last two decades have suggested that there is a naturally-occurring biomolecule that binds trivalent Cr that can explain how Cr is involved in carbohydrate and lipid metabolism. This molecule, chromodulin (originally termed low-molecular-weight Cr-binding substance), is a naturally-occurring oligopeptide composed of glycine, cysteine, aspartate and glutamate with the carboxylates comprising more than half the total amino acid residues (Yamamoto et al. 1987; Davis & Vincent, 1997b). Despite its small size (molecular weight approximately 1438 for the bovine liver material), the molecule tightly binds four equivalents of Cr^{3+} . The binding is quite tight (association constant approximately 10^{21} M^4) and highly cooperative (Hill coefficient, n 3.47; Sun et al. 2000). Spectroscopic studies suggest that the Cr³⁺ comprise an anion-bridged multinuclear assembly supported by carboxylates from the oligopeptide (Davis & Vincent, 1997b; Jacquamet et al. 2003). Chromodulin has been isolated from the liver or kidney of several mammals and purified. A related Cr-containing oligopeptide (M-lowmolecular-weight Cr-binding substance) has been isolated from bovine colostrum, which comprises the same amino acids, but in distinctly different ratios, and also stimulates insulin-dependent glucose metabolism in rat adipocytes (Yamamoto et al. 1988). The relationship between the milk and liver oligopeptides is deserving of additional

study and suggests the possibility that other carboxylaterich oligopeptides might exist and play a role in Cr transport or function.

The oligopeptide is maintained *in vivo* in the apo form (Yamamoto *et al.* 1987; Davis & Vincent, 1997*b*). This observation has resulted in the suggestion that chromodulin may play a role in Cr detoxification. However, injection of Cr^{3+} or chromate into mice does not stimulate the production of apochromodulin (Yamamoto *et al.* 1984). Chromodulin does carry Cr into the urine after the intake of large dosages of trivalent and hexavalent Cr (Wada *et al.* 1983; Clodfelder *et al.* 2001) and can, therefore, assist in Cr detoxification.

Another potential function for chromodulin has been identified by the author's laboratory. Chromodulin has been shown to activate the tyrosine kinase activity of insulin-activated insulin receptor (Davis et al. 1997; Davis & Vincent, 1997a) and to activate a membrane phosphotyrosine phosphatase in adipocyte membranes (Davis et al. 1996). For example, the addition of bovine liver chromodulin to rat adipocytic membranes in the presence of 100 nm-insulin results in an up to eight-fold stimulation of insulin-dependent protein tyrosine kinase activity that is concentration dependent, while no activation of kinase activity is observed in the absence of insulin (Davis & Vincent, 1997a). The dependence of the kinase activation on the concentration of chromodulin can be fitted to a hyperbolic curve to give dissociation constants of approximately 875 pm. Cr plays a crucial role in the activation of insulin receptor kinase activity by chromodulin (Davis & Vincent, 1997a). Apochromodulin does not activate insulin-dependent tyrosine kinase activity in rat adipocyte membranes. Titration of apochromodulin with Cr³⁺ results in the total restoration of the ability to activate kinase activity; approximately four ${\rm Cr}^{3+}$ per oligopeptide are required for maximal activity. This reconstitution of chromodulin's activation potential is specific to Cr; addition of other biologically-relevant metals does not restore activity to the apo-oligopeptide.

Based on these results, it has been proposed by the author's laboratory that chromodulin functions as part of a unique autoamplification system for insulin signalling (Vincent, 2000b,c). In this mechanism apochromodulin is stored in insulin-sensitive cells. In response to increases in blood insulin concentrations insulin binds to its receptor, bringing about a conformation change that results in the autophosphorylation of tyrosine residues on the internal side of the receptor. This process transforms the receptor into an active tyrosine kinase and transmits the signal from insulin into the cell. In response to insulin Cr is moved from the blood to insulin-sensitive cells. Here, the Cr flux results in the loading of apochromodulin with Cr. The holochromodulin then binds to the receptor, presumably assisting in the maintenance of the receptor in its active conformation, amplifying the receptor's kinase activity. When the signalling is to be turned off, a drop in blood insulin levels facilitates relaxation of the conformation of the receptor, and the holochromodulin is excreted from the cell into the blood. Ultimately, chromodulin is efficiently excreted in the urine. The Fe-transport protein transferrin has recently been shown to be responsible for maintaining

 Cr^{3+} levels in the blood plasma and for transporting Cr to tissues in an insulin-responsive manner (Vincent, 2000*a*; Clodfelder *et al.* 2001). The basis of the name chromodulin is the similarity of this proposed mechanism of action to that of the Ca-binding protein calmodulin (Vincent, 2000*b*). Both molecules bind four equivalents of metal ions in response to a metal ion flux; however, the four Ca²⁺ that bind to the larger protein calmodulin lie in mononuclear sites. Both holoproteins selectively bind to kinases and phosphatases, thus stimulating their activity.

A potential chromium therapeutic

The existence of a multinuclear trivalent Cr-carboxylate assembly in an active biomolecule has spurred an interest in the synthesis and characterization of multinuclear oxo(hydroxo)-bridged trivalent Cr-carboxylate assemblies (Vincent, 2000b). Well-characterized water-soluble assemblies have been tested by the author's laboratory for the ability to stimulate insulin receptor's tyrosine kinase ability in a manner similar to chromodulin. The trinuclear cation $[Cr_3O(O_2CCH_2CH_3)_6(H_2O)_3]^+$ was found in vitro to mimic the ability of chromodulin to stimulate this activity (Davis et al. 1997); thus, the cation is a functional biomimetic of chromodulin. Consequently, the trinuclear cation has been proposed as a potential therapeutic agent to increase insulin sensitivity. The synthetic complex has other several potential benefits over the natural material. For example, it is inexpensive to synthesize and can be readily prepared in bulk. Also, while chromodulin is susceptible to hydrolysis, especially in the presence of acid, the synthetic material can be recrystallized from dilute mineral acid (Johnson et al. 1981) and should consequently survive oral ingestion. The biomimetic cation remains intact after injection into the bloodstream and is subsequently taken up intact by cells (Shute & Vincent, 2001, 2002). When given intravenously to healthy rats at a level equivalent to 20 µg Cr/kg per body mass per d for 12 weeks the biomimetic has been shown to lower fasting serum triacylglycerol and total cholesterol levels; administration of similar quantities of propionate by itself had no effect (Sun et al. 1999). Subsequently, in healthy rats 24 weeks of intravenous administration of the cation (0-20µg Cr/kg body mass), resulted in a concentrationdependent lowering of levels of fasting blood plasma LDL-cholesterol, total cholesterol, triacylglycerols and insulin, and of 2h plasma insulin and glucose levels after a glucose challenge (Sun et al. 2002). The cation had little, if any, effect on rats with streptozotocin-induced diabetes (a type 1 diabetes model). However, after 24 weeks of supplementation (20µg/kg) Zucker obese rats (a model of the early stages of type 2 diabetes) had lower fasting plasma total cholesterol, HDL- and LDL-cholesterol, triacylglycerol and insulin levels and lower 2h plasma insulin levels (Sun et al. 2002). The lowering of plasma insulin concentrations with little effect on glucose concentrations suggests that the cation increases insulin sensitivity. The author's laboratory, as part of an ongoing study, has found that the trinuclear biomimetic complex, when given orally at amounts equivalent to those used in

early intravenous studies, has beneficial effects after just 8 weeks of administration (BJ Clodfelder, B Gullick and JB Vincent, unpublished results).

Conclusions

While the evidence is not overwhelming, it suggests that Cr is an essential trace element in mammals, including man, and affects carbohydrate and lipid metabolism through the action of insulin. The daily requirement for human subjects is small, i.e. approximately $30 \mu g$, such that it is difficult for healthy individuals to develop Cr deficiency. Thus, the use of Cr supplements is probably unnecessary for the general public. However, the use of certain Cr supplements, such as Cr(pic)₃, is probably harmful. While nutritional supplement levels of Cr do not appear to have beneficial effects, pharmacological quantities of Cr may increase insulin sensitivity in both healthy subjects and subjects with type 2 diabetes. The biomolecule chromodulin may be part of an insulin-potentiating pathway and may explain the requirement for Cr.

Acknowledgements

Research on Cr biochemistry in the author's laboratory is funded by the American Diabetes Association and the National Institutes of Health (DK62094–01). The author notes that the University of Alabama holds four patents resulting from his research and dealing with isolated oligopeptides or trivalent Cr-containing complexes and their potential use as nutritional supplements or drugs. However, to date neither the author nor the University of Alabama have marketed or formed a company to market the Cr-containing species.

References

- Althuis MD, Jordan NE, Ludington EA & Wittes JT (2002) Glucose and insulin responses to dietary chromium supplements: a meta-analysis. *American Journal of Clinical Nutrition* 76, 148–155.
- Anderson RA (1998) Chromium, glucose tolerance and diabetes. Journal of the American College of Nutrition **17**, 548–555.
- Anderson RA (2000) Chromium in the prevention and control of diabetes. *Diabetes Metabolism (Paris)* 26, 22–27.
- Anderson RA, Bryden NA & Polansky MM (1993) Dietary intake of calcium, chromium, copper, iron, magnesium, manganese and zinc: duplicate plate values corrected using nutrient intake. *Journal of the American Dietetic Association* **93**, 462–464.
- Anderson RA, Bryden NA & Polansky MM (1997*a*) Lack of toxicity of chromium chloride and chromium picolinate in rats. *Journal of the American College of Nutrition* **16**, 273–279.
- Anderson RA, Bryden NA, Polansky MM & Reisner S (1990) Urinary chromium excretion and insulinogenic properties of carbohydrates. *American Journal of Clinical Nutrition* 51, 864–868.
- Anderson RA, Cheng NC, Bryden NA, Polansky MM, Cheng N, Chi J & Feng J (1997*b*) Elevated levels of supplemental chromium improve glucose and insulin variables in individuals with type 2 diabetes. *Diabetes* **46**, 1786–1791.

Biological Trace Element Research **50**, 97–108. Bagchi D, Bagchi M, Balmoori J, Ye X & Stohs SJ (1997) Comparative induction of oxidative stress in cultured J774A.1 macrophage cells by chromium picolinate and chromium nicotinate. Research Communications in Molecular Pathology and Pharmacology **97**, 335–346.

Anderson RA & Kozlovsky AS (1985) Chromium intake,

absorption and excretion of subjects consuming self-selected

diets. American Journal of Clinical Nutrition 41, 1177-1183.

Anderson RA & Polansky MM (1995) Dietary and metabolite

- Bagchi D, Stohs SJ, Downs BW, Bagchi M & Preuss HG (2002) Cytotoxicity and oxidative mechanisms of different forms of chromium. *Toxicology* 180, 5–22.
- Bunker VW, Lawson MS, Delues HT & Clayton BE (1984) The uptake and excretion of chromium by the elderly. *American Journal of Clinical Nutrition* **39**, 797–802.
- Cefalu WT, Wang ZQ, Zhang XH, Baldor LC & Russell JC (2002) Oral chromium picolinate improves carbohydrate and lipid metabolism and enhances skeletal muscle Glut-4 translocation in obese, hyperinsulinemic (JCR-LA corpulent) rats. *Journal of Nutrition* **132**, 1107–1114.
- Cerulli J, Grabe DW, Gauthier I, Malone M & McGoldrick MD (1998) Chromium picolinate toxicity. *Annals of Pharmacotherapy* **32**, 428–431.
- Cheng N, Zhu X, Shi H, Wu W, Chi J, Cheng J & Anderson RA (1999) Follow-up survey of people in China with type 2 diabetes mellitus consuming supplemental chromium. *Journal of Trace Elements in Experimental Research* **12**, 55–60.
- Clarkson PM (1997) Effects of exercise on chromium levels: is supplementation required? *Sports Medicine* **23**, 341–349.
- Clodfelder BJ, Emamaullee J, Hepburn DD, Chakov NE, Nettles H & Vincent JB (2001) The trail of chromium(III) from the blood to the urine: the roles of transferrin and chromodulin. *Journal of Biological Inorganic Chemistry* **6**, 608–617.
- Davis CM, Royer AC & Vincent JB (1997) Synthetic multinuclear chromium assembly activates insulin receptor kinase activity: functional model for low-molecular-weight chromiumbinding substance. *Inorganic Chemistry* 36, 5316–5320.
- Davis CM, Sumrall KH & Vincent JB (1996) The biologically active form of chromium may activate a membrane phosphotyrosine phosphatase (PTP). *Biochemistry* 35, 12963–12969.
- Davis CM & Vincent JB (1997*a*) Chromium oligopeptide activates insulin receptor tyrosine kinase activity. *Biochemistry* **36**, 4382–4385.
- Davis CM & Vincent JB (1997*b*) Isolation and characterization of a biologically active chromium oligopeptide from bovine liver. *Archives of Biochemistry and Biophysics* **339**, 335–343.
- Evans GW (1989) The effect of chromium picolinate on insulin controlled parameters in humans. *International Journal of Biosocial Medicine and Research* **11**, 163–180.
- Fowler JF Jr (2000) Systemic contact dermatitis caused by oral chromium picolinate. *Cutis* **65**, 116.
- Ghosh D, Bhattacharya B, Mukherjee B, Manna B, Sinha M, Chowdhury J & Chowdhury S (2002) Role of chromium supplementation in Indians with type 2 diabetes mellitus. *Journal of Nutritional Biochemistry* **13**, 690–697.
- Gibson RS & Scythes CA (1984) Chromium, selenium, and other trace element intakes of a selected sample of Canadian premenopausal women. *Biological Trace Element Research* 6, 105–116.
- Hellerstein MK (1998) Is chromium supplementation effective in managing type II diabetes? *Nutrition Reviews* 56, 302–306.
- Hepburn DDD, Burney JM, Woski SA & Vincent JB (2003*a*) The nutritional supplement chromium picolinate generates oxidative DNA damage and peroxidized lipids in vivo. *Polyhedron* 22, 455–463.

- Hepburn DDD & Vincent JB (2002) In vivo distribution of chromium from chromium picolinate in rats and implications for the safety of the dietary supplement. *Chemical Research in Toxicology* **15**, 93–100.
- Hepburn DDD & Vincent JB (2003) Tissue and subcellular distribution of chromium picolinate with time after entering the bloodstream. *Journal of Inorganic Biochemistry* **94**, 86–93.
- Hepburn DDD, Xiao J, Bindom S, Vincent JB & O'Donnell J (2003b) Nutritional supplement chromium picolinate causes sterility and lethal mutations in *Drosophila melanogaster*. *Proceedings of the National Academy of Sciences USA* **100**, 3766–3771.
- Huszonek J (1993) Over-the-counter chromium picolinate. American Journal of Psychiatry **150**, 1560.
- Jacquamet L, Sun Y, Hatfield J, Gu W, Cramer SP, Crowder MW, Lorigan GA, Vincent JB & Latour J-M (2003) Characterization of chromodulin by X-ray absorption and electron paramagnetic resonance spectroscopies and magnetic susceptibility measurements. *Journal of the American Chemical Society* 125, 774–780.
- Jeejeebhoy KN (1999) Chromium and parenteral nutrition. Journal of Trace Elements in Experimental Medicine 12, 85–89.
- Johnson MK, Powell DB & Cannon RD (1981) Vibrational spectra of carboxylate complexes-III. Spectrochimica Acta 37A, 995–1006.
- Jovanovic L, Gutierrez M & Peterson CM (1999) Chromium supplementation for women with gestational diabetes mellitus. *Journal of Trace Elements in Experimental Medicine* **12**, 91–97.
- Kareus SA, Kelley C, Walton HS & Sinclair PC (2001) Release of Cr(III) from Cr(III) picolinate upon metabolic activation. *Journal of Hazardous Materials* **84**B, 163–174.
- Kato I, Vogelman JH, Dilman V, Karkoszka J, Frenkel K, Durr NP, Orentreich N & Toniolo P (1998) Effect of supplementation with chromium picolinate on antibody titers to 5-hydroxymethyl uracil. *European Journal of Epidemiology* 14, 621–626.
- Kim D-S, Kim T-W, Park I-K, Kang J-S & Om A-S (2002) Effects of chromium picolinate supplementation on insulin sensitivity, serum lipids, and body weight in dexamethasonetreated rats. *Metabolism* 51, 589–594.
- Kozlovsky AS, Moser PB, Reisner RA & Anderson RA (1986) Effects of diets high in simple sugars on urinary chromium losses. *Metabolism* 35, 515–518.
- Kreider RB (1999) Dietary supplements and the promotion of muscle growth with resistance exercise. *Sports Medicine* **27**, 97–110.
- Lukaski HC (1999) Chromium as a supplement. Annual Review of Nutrition 19, 279–302.
- Mahboob L, McNeil L, Tolliver T & Ogden L (2002) Effects of chromium picolinate on antioxidant enzyme levels in rats. *Toxicological Sciences* 66, Suppl. 1, 32.
- Manygoats KR, Yazzie M & Stearns DM (2002) Ultrastructural damage in chromium picolinate-treated cells: a TEM study. *Journal of Biological Inorganic Chemistry* 7, 791–798.
- Martin WR & Fuller RE (1998) Suspected chromium picolinateinduced rhabdomyolysis. *Pharmacotherapy* 18, 860–862.
- Mirasol F (2000) Chromium picolinate market sees robust growth and high demand. *Chemical Market Reporter* **257**, 2000.
- Morris B, MacNeil S, Fraser R & Gray T (1995) Increased urine chromium excretion in normal pregnancy. *Clinical Chemistry* 41, 1544–1545.
- Morris BW, MacNeil S, Hardisty CA, Heller S, Burgin C & Gray TA (1999) Chromium homeostasis in patients with type II (NIDDM) diabetes. *Journal of Trace Elements in Medicine and Biology* **13**, 57–61.

- Morris BW, MacNeil S, Stanley K & Gray TA (1993) The interrelationship between insulin and chromium in hyperinsulinaemic euglycaemic clamps in healthy volunteers. *Journal of Endocrinology* **139**, 339–345.
- Nielsen F (1996) Controversial chromium: does the superstar mineral of the Mountebanks receive appropriate attention from clinicians and nutritionists? *Nutrition Today* **31**, 226–233.
- Nissen SL & Sharp RL (2003) Effect of dietary supplements on lean mass and strength gains with resistance exercise: A meta-analysis. *Journal of Applied Physiology* 94, 651–659.
- Offenbacher EG, Spencer H, Dowling HJ & Pi-Sunyer FX (1986) Metabolic chromium balances in men. *American Journal of Clinical Nutrition* **44**, 77–82.
- Olin KL, Stearns DM, Armstrong WH & Keen CL (1994) Comparative retention/adsorption of ⁵¹chromium (⁵¹Cr) from ⁵¹Cr chloride, ⁵¹Cr nicotinate, and ⁵¹Cr picolinate in a rat model. *Trace Elements and Electrolytes* **11**, 182–186.
- Pittler MH, Stevinson C & Ernst E (2003) Chromium picolinate for reducing body weight: Meta-analysis of randomized trials. *International Journal of Obesity* 27, 522–529.
- Ravina A, Slezak L, Mirsky N & Anderson RA (1999a) Control of steroid-induced diabetes with supplemental chromium. *Journal* of Trace Elements in Medicine and Biology 12, 375–378.
- Ravina A, Slezak L, Mirsky N, Bryden NA & Anderson RA (1999b) Reversal of corticosteroid-induced diabetes mellitus with supplemental chromium. *Diabetic Medicine* 16, 164–167.
- Shute AA & Vincent JB (2001) The stability of the biomimetic cation triaqua-µ-oxohexapropionatotrichromium(III) in vivo in rats. *Polyhedron* **20**, 2241–2252.
- Shute AA & Vincent JB (2002) The fate of the biomimetic cation triaqua-μ-oxohexapropionatotrichromium(III) in rats. *Journal of Inorganic Biochemistry* **89**, 272–278.
- Speetjens JK, Collins RA, Vincent JB & Woski SA (1999a) The nutritional supplement chromium(III) tris(picolinate) cleaves DNA. Chemical Research in Toxicology 12, 483–487.
- Speetjens JK, Parand A, Crowder MW, Vincent JB & Woski SA (1999*b*) Low-molecular-weight chromium-binding substance and biomimetic $[Cr_3O(O_2CCH_2CH_3)_6(H_2O)_3]^+$ do not cleave DNA under physiologically-relevant conditions. *Polyhedron* **18**, 2617–2624.
- Stearns DM (2000) Is chromium a trace essential element? *Biofactors* **11**, 149–162.
- Stearns DM, Silveira SM, Wolf KK & Luke AM (2002) Chromium(III) tris(picolinate) is mutagenic at the hypoxanthine (guanine) phosphoribosyltransferase locus in Chinese hamster ovary cells. *Mutation Research* 513, 135–142.
- Stearns DM, Wise JP Sr, Patierno SR & Wetterhahn KE (1995) Chromium(III) picolinate produces chromosome damage in Chinese hamster ovary cells. *FASEB Journal* 9, 1643–1648.
- Striffler JS, Law JS, Polansky MM, Bhathena SJ & Anderson RA (1995) Chromium improves insulin response to glucose in rats. *Metabolism* 44, 1314–1320.
- Striffler JS, Polansky MM & Anderson RA (1999) Overproduction of insulin in the chromium-deficient rat. *Metabolism* 48, 1063–1068.

- Sugden KD, Geer RD & Rogers SG (1992) Oxygen radicalmediated DNA damage by redox-active Cr(III) complexes. *Biochemistry* 31, 11626–11631.
- Sun Y, Clodfelder BJ, Shute AA, Irvin T & Vincent JB (2002) The biomimetic $[Cr_3O(O_2CCH_2CH_3)_6(H_2O)_3]^+$ decreases plasma insulin, cholesterol and triglycerides in healthy and type II diabetic rats but not type I diabetic rats. *Journal of Biological Inorganic Chemistry* **7**, 852–862.
- Sun Y, Mallya K, Ramirez J & Vincent JB (1999) The biomimetic $[Cr_3O(O_2CCH_2CH_3)_6(H_2O)_3]^+$ decreases cholesterol and triglycerides in rats: towards chromium-containing therapeutics. *Journal of Biological Inorganic Chemistry* **4**, 838–845.
- Sun Y, Ramirez J, Woski SA & Vincent JB (2000) The binding of chromium to low-molecular-weight chromium-binding substance (LMWCr) and the transfer of chromium from transfer and chromium picolinate to LMWCr. *Journal of Biological Inorganic Chemistry* 5, 129–136.
- Trumbo P, Yates AA, Schlickek S & Poos M (2001) Dietary reference intakes: vitamin A, vitamin K, arsenic, boron, chromium, copper, iodine, iron, molybdenum, nickel, silicon, vanadium, and zinc. *Journal of the American Dietetic Association* **101**, 294–301.
- Vincent JB (2000a) The biochemistry of chromium. Journal of Nutrition 130, 715–718.
- Vincent JB (2000b) Elucidating a biological role for chromium at a molecular level. Accounts of Chemical Research 33, 503–510.
- Vincent JB (2000c) Quest for the molecular mechanism of chromium action and its relationship to diabetes. *Nutrition Reviews* 58, 67–72.
- Vincent JB (2001) The bioinorganic chemistry of chromium(III). Polyhedron 20, 1–26.
- Vincent JB (2003) The potential value and potential toxicity of chromium picolinate as a nutritional supplement, weight loss agent, and muscle development agent. *Sports Medicine* 33, 213–230.
- Wada O, Wu GY, Yamamoto A, Manabe S & Ono T (1983) Purification and chromium-excretory function of lowmolecular-weight, chromium-binding substances from dog liver. *Environmental Research* 32, 228–239.
- Wasser WG & D'Agati VD (1997) Chromic renal failure after ingestion of over-the-counter chromium picolinate. *Annals of Internal Medicine* **126**, 410.
- Yamamoto A, Ono T & Wada O (1987) Isolation of a biologically active low-molecular-mass chromium compound from rabbit liver. *European Journal of Biochemistry* 165, 627–631.
- Yamamoto A, Wada O & Ono T (1984) Distribution and chromium-binding capacity of a low-molecular-weight, chromium-binding substance in mice. *Journal of Inorganic Biochemistry* 22, 91–102.
- Yamamoto A, Wada O & Suzuki H (1988) Purification and properties of biologically active chromium complex from bovine colostrum. *Journal of Nutrition* **118**, 39–45.