The Annual Meeting of the Nutrition Society and BAPEN was held at Cardiff International Arena, Cardiff on 13–14 October 2009

Conference on 'Malnutrition matters'

Satellite symposium: Throw another fish on the fire: the role of *n*-3 in inflammation Rationale and use of *n*-3 fatty acids in artificial nutrition

Philip C. Calder

Institute of Human Nutrition, School of Medicine, University of Southampton, IDS Building, MP887 Southampton General Hospital, Tremona Road, Southampton SO16 6YD, UK

> Lipids traditionally used in artificial nutrition are based on n-6 fatty acid-rich vegetable oils like soyabean oil. This may not be optimal because it may present an excessive supply of linoleic acid. One alternative to the use of soyabean oil is its partial replacement by fish oil, which contains n-3 fatty acids. These fatty acids influence inflammatory and immune responses and so may be useful in particular situations where those responses are not optimal. Fish oil-containing lipid emulsions have been used in parenteral nutrition in adult patients postsurgery (mainly gastrointestinal). This has been associated with alterations in patterns of inflammatory mediators and in immune function and, in some studies, a reduction in length of intensive care unit (ICU) and hospital stay. Perioperative administration of fish oil may be superior to post-operative. Parenteral fish oil has been used in critically ill adults. Here the influence on inflammatory processes, immune function and clinical endpoints is not clear, since there are too few studies and those that are available report contradictory findings. Fish oil is included in combination with other nutrients in various enteral formulas. In post-surgical patients and in those with mild sepsis or trauma, there is clinical benefit from a formula including fish oil and arginine. A formula including fish oil, borage oil and antioxidants has demonstrated marked benefits on gas exchange, ventilation requirement, new organ failures, ICU stay and mortality in patients with acute respiratory distress syndrome, acute lung injury or severe sepsis.

Fish oil: Soyabean oil: Fatty acid: Parenteral nutrition: Enteral nutrition

This review presents the rationale for the inclusion of marine *n*-3 fatty acids, in the form of fish oil, in artificial nutrition regimens for use in various patient groups, including both parenteral and enteral applications. The review goes on to describe and interpret studies using parenteral or enteral fish oil in various patient groups with a focus on immune, inflammatory and clinical outcomes. The material described is largely based upon that presented in previous review articles of this subject⁽¹⁻³⁾, but here this information is updated with respect to additional studies of relevance⁽⁴⁾, new meta-analyses⁽⁵⁾ and newly published guidelines for parenteral nutrition^(6,7).

Fatty acids

Fatty acids are hydrocarbon chains with a carboxyl group at one end and a methyl group at the other⁽⁸⁾. The carboxyl group is reactive and readily forms ester links with alcohol groups, for example those on glycerol or cholesterol, in turn forming acylglycerols (e.g. TAG and phospholipids) and cholestryl esters. Fatty acid chain lengths vary from two to 30 or more and the chain may contain double bonds. Fatty acids containing double bonds in the hydrocarbon chain are referred to as unsaturated fatty acids; a fatty acid containing one double bond is called a monounsaturated

Abbreviations: ICU, intensive care unit; MCT, TAG containing predominantly medium-chain fatty acids. Corresponding author: Philip C. Calder, fax 44-23-8079 5255, email pcc@soton.ac.uk

 Table 1. Common names, shorthand nomenclature and typical sources of fatty acids used in artificial nutrition. Modified from Calder⁽³⁾

Common name	Shorthand nomenclature	Typical source
Caprylic acid	8:0	Coconut oil
Capric acid	10:0	Coconut oil
Myristic acid	14:0	Coconut oil
Palmitic acid	16:0	Olive oil, soyabean oil and fish oi
Oleic acid	18:1 <i>n</i> -9	Olive oil and soyabean oil
Linoleic acid	18:2 <i>n</i> -6	Soyabean oil
γ-Linolenic acid	18:3 <i>n</i> -6	Borage oil
α -Linolenic acid	18:3 <i>n</i> -3	Soyabean oil
EPA	20:5 <i>n</i> -3	Fish oil
DHA	22:6 <i>n</i> -3	Fish oil

fatty acid, while one containing two or more double bonds is called a PUFA. Fatty acids have common names (Table 1) and systematic names. They are also referred to by a shorthand nomenclature that denotes the number of carbon atoms in the chain, the number of double bonds and the position of the first double bond relative to the methyl (n) carbon (Table 1). n-3 and n-6 fatty acids are so-called because the first double bond is on carbon number 3 or 6, respectively, counting the methyl carbon as carbon number 1. The simplest n-6 fatty acid is linoleic acid (18:2n-6) and the simplest *n*-3 fatty acid is α -linolenic acid (18:3*n*-3). Linoleic and α -linolenic acids cannot be synthesised in animals, including human subjects. They are the classical essential fatty acids. In contrast, saturated and monounsaturated fatty acids can be synthesised de novo in human subjects⁽²⁾.

Although mammalian cells cannot synthesise linoleic and α -linolenic acids, they can metabolise them by further desaturation and elongation. Linoleic acid can be converted to γ -linolenic acid (18:3*n*-6), then to dihomo- γ -linolenic acid (20:3*n*-6) and then to arachidonic acid (20:4*n*-6). Using the same series of enzymes, α -linolenic acid is converted to EPA (20:5*n*-3). A complex pathway for further conversion of EPA to DHA (22:6*n*-3) exists⁽⁸⁻¹⁰⁾. Fatty acids that are important in artificial nutrition and their typical sources for this purpose are listed in Table 1.

Desirable properties for lipids to be used in artificial nutrition

Lipids used in artificial nutrition should provide:

a source of energy as an alternative to glucose;

building blocks, since patients requiring artificial nutrition will typically be undergoing processes involving cell replication and tissue repair;

essential fatty acids in order to avoid deficiency symptoms;

a 'good' fatty acid balance, although the precise definition of this balance is still lacking;

fatty acids with desirable biological activities; it is now recognised that fatty acids can affect cell membrane properties, cell signalling, gene expression and the production of bioactive mediators^(8,11,12), and it would

be desirable if the fatty acids provided as a component of artificial nutrition, at worst, did not exacerbate inappropriate cellular responses and, at best, modulated these in a manner that would improve patient outcome.

Use of fish oil in parenteral nutrition in surgical and critically ill patients

Rationale for the use of fish oil in parenteral nutrition

Lipids were first introduced into parenteral nutrition formulas in the 1960s in order to provide a more balanced supply of energy, along with $glucose^{(13-15)}$. The lipid typically used in parenteral nutrition is soyabean oil, in which linoleic acid comprises about 50% of fatty acids present. A meta-analysis of total parenteral nutrition suggested that inclusion of lipids might be detrimental $(P = 0.09 \text{ for lipids } v. \text{ no lipids})^{(16)}$, at least in very ill patients; most of the studies included in the meta-analysis used soyabean oil-based lipid emulsions. A more recent study in patients following major gastrointestinal surgery identified that the amount of *n*-6 PUFA (i.e. linoleic acid) infused was one of two predictors of the length of hospital stay (increased by 1.6 d/100 g of *n*-6 PUFA infused), the other being the delay in the onset of initiating nutritional support⁽¹⁷⁾. A number of *in vitro* experiments have shown that soyabean oil-based lipid emulsions can exert immunosuppressive effects (see Calder *et al.*⁽¹⁸⁾ for references), which would clearly be detrimental in patients at risk of infection and sepsis. Clinical trials with soyabean oil-based lipid emulsions provide conflicting evidence, some show-ing selective immunosuppressive effects^(19–21), perhaps linked to poorer patient outcomes⁽²⁰⁾. However, other studies do not show such effects on the immune system⁽²²⁻²⁴⁾ or on clinical outcomes⁽²⁵⁾. These studies have been described and discussed previously^(1,3). Despite the inconsistencies of the outcomes of such studies, a view has developed that the use of lipid emulsions based solely upon soyabean oil may not be optimal or may even be harmful. The concern about potential harm, based mainly on the idea that n-6 PUFA might be 'pro-inflammatory, immunosuppressive and pro-coagulatory', has led to the development of alternative lipid emulsions for parenteral applications. Two alternative philosophies towards reducing the amount of linoleic acid have been adopted. The first has been to simply dilute soyabean oil with another oil that is fairly inert. Examples of this strategy include the use of so-called medium-chain TAG (MCT; TAG containing predominantly medium-chain fatty acids) and olive oil. This approach is not discussed further here, but has been described in detail elsewhere⁽²⁶⁻²⁸⁾. The second approach has been to partially replace soyabean oil with another oil that is believed to exert benefits in its own right. An example of this strategy is the use of fish $\operatorname{oil}^{(29-32)}$.

Fish oil contains the very long-chain *n*-3 PUFA EPA and DHA. There is strong evidence for health benefits of these fatty acids, especially with regard to $\text{CVD}^{(33-36)}$. They act to modify tissue and blood lipid metabolism, blood lipid concentrations, blood coagulation, immune function, inflammation and endothelial function⁽³⁷⁻⁴¹⁾.

EPA and DHA are readily incorporated into cells and tissues and act to modify membrane properties, eicosanoid profiles, signal transduction processes and gene expression^(11,12). Through these mechanisms they result in improved cell and tissue function. Thus, using fish oil to partly replace soyabean oil in parenteral nutrition offers the possibility to both decrease the amount of linoleic acid present and increase the amount of biologically active *n*-3 PUFA.

Animal feeding studies have demonstrated that fish oil decreases the production of inflammatory eicosanoids and cytokines in endotoxaemia^(42–44) or sepsis^(45–47) and that this is associated with a decreased metabolic response^(48–51), improved organ function^(43,52–57) and improved survival^(45–47,58–60).

Three lipid emulsions that include fish oil as a component are currently available. Omegaven[®] (Fresenius Kabi, Bad Homberg, Germany) is a pure fish oil emulsion (100 g lipid/l) that will typically contain about 3 g EPA plus DHA/100 ml. It is recommended that Omegaven is used in combination with other emulsions (e.g. those based on soyabean oil) such that Omegaven contributes 10-20% of infused emulsion. Lipoplus[®] (also known as Lipidem[®]; B. Braun, Melsungen, Germany) is an emulsion (200 g lipid/l) with the lipid being a mix of 50% MCT, 40% soyabean oil and 10% fish oil. Each 100 ml of Lipoplus will typically contain about 0.6 g EPA plus DHA. SMOFLipid[®] (Fresenius Kabi, Bad Homberg, Germany) is an emulsion (200 g lipid/l) with the lipid being a mix of 30% MCT, 30% soyabean oil, 25% olive oil and 15% fish oil. Each 100 ml of SMOFLipid[®] will typically contain about 1 g EPA plus DHA.

Studies of parenteral fish oil in surgical patients

Intravenous infusion of lipid emulsions containing fish oil into patients following gastrointestinal surgery altered the fatty acid composition of $plasma^{(61-64)}$, $platelet^{(65)}$ and erythrocyte⁽⁶⁴⁾ phospholipids: typically EPA content was increased. Intravenous infusion of a lipid emulsion containing fish oil into patients for 5 d following gastrointestinal surgery also altered the fatty acid composition of leucocytes: EPA content was increased 2.5-fold⁽⁶⁶⁾. This would be expected to impact on the profile of eicosanoids produced from arachidonic acid and EPA. Indeed, several studies have demonstrated that intravenous infusion of lipid emulsions containing fish oil into patients who had undergone major gastrointestinal surgery results in lower production of arachidonic acid-derived eicosanoids and higher production of EPA-derived eicosanoids by blood leucocytes stimulated *ex vivo*^(61,62,66,67). Plasma TNF- α concentrations were lower at day 6 post-surgery, while plasma IL-6 concentrations were lower at day 10 postsurgery in patients who had undergone major gastrointestinal surgery and then received a mix of MCT, soyabean oil and fish oil (50:30:20 by vol.; this was a prototype version of Lipoplus) for 5 d post-surgery compared with those who received an MCT-soyabean oil mix⁽⁶⁶⁾. The study did not report clinical outcomes. Another study infused Omegaven, providing 10 g fish oil/d, on the day before abdominal surgery and on days 1-5

following abdominal surgery⁽⁶⁸⁾. On days 4 and 5 the patients also received standard total parenteral nutrition, which included 50 g fat/d as soyabean oil. TNF- α production by endotoxin-stimulated whole blood tended to be lower at post-operative day 5 in the fish oil group, but this was not significant. Serum IL-6 concentrations were significantly lower at days 0, 1 and 3 in the fish oil group than in controls. Monocyte expression of human leucocyte antigen-DR, an indication of ability to present antigen and so to mount an immune response, was preserved in the fish oil group, but declined at post-surgery days 3 and 5 in the control group. No differences in infection rates or mortality were observed. However, post-operative stay in intensive care tended to be shorter in the fish oil group (4.1 v. 9.1 d)in the control group) as did total hospital stay (17.8 v. 23.5 d). Post-operative stay on medical wards was significantly shorter in the fish oil group (P < 0.05). Another study compared the effects of lipid-free total parenteral nutrition or parenteral nutrition including soyabean oil or a mix of 83% soyabean oil and 17% fish oil from Omegaven for 5 d after large bowel surgery⁽⁶⁹⁾. There were no dif-</sup> ferences between the groups with respect to the numbers of circulating lymphocytes, B lymphocytes, helper T lymphocytes, cytotoxic T lymphocytes or natural killer cells before surgery or at days 3 and 6 post-surgery, although these were affected by surgery itself. There were no differences between groups with respect to T lymphocyte proliferation, but IL-2 production was increased in the fish oil group and the post-surgery decline in interferon- γ production was prevented by fish oil. Liang *et al.*⁽⁴⁾ compared soyabean oil with a mixture of soyabean oil and Omegaven (5:1, v/v) over 7 d in patients who had undergone radical colorectal cancer resection. The decline in serum IL-6 concentration between post-operative day 1 and day 8 was greater in the group receiving fish oil, while the increase in the ratio of CD4⁺ to CD8⁺ cells in the bloodstream, believed to be a marker of the cell-mediated immune response, was greater in that group. Length of hospital stay tended to be shorter in the fish oil group (17.5)v. 19.6d in the control group) and infectious complications and mortality were not different between groups. Wichmann et al.⁽⁶³⁾ reported length of hospital stay in post-gastrointestinal surgery patients receiving soyabean oil or Lipoplus: length of stay was significantly shorter (P = 0.006) in patients receiving fish oil (17.2 d) than in the control group (21.9 d). In another study in post-surgical patients, SMOFLipid for 6d resulted in significantly shorter hospital stay (13.4 v. 20.4 d; P < 0.05) than soyabean oil⁽⁶²⁾. Taken together, these studies indicate that inclusion of fish oil in parenteral nutrition regimens for gastrointestinal surgical patients modulates the generation of inflammatory eicosanoids^(61,62,66,67) and cytokines^(4,66,68) and may help to counter the surgery-induced decline in antigen presenting cell activity⁽⁶⁸⁾ and T lymphocyte cytokine production⁽⁶⁹⁾. Importantly, these studies do not reveal any deleterious effects of fish oil infusion in these patients. Furthermore, studies that have examined the hard endpoint of length of hospital stay suggest a real clinical benefit from fish oil infusion in these patients^(62,63,68). Another report from a cohort of patients receiving parenteral nutrition post-surgery also indicates benefit from

the inclusion of fish oil in the regimen $^{(70)}$. There were no differences between the control group (MCT-soyabean oil) and the patients receiving fish oil (a mix of Omegaven with a 50:50 MCT-soyabean oil mix, where a maximum of one-third of the mix was as fish oil) with respect to the proportion of patients who developed wound infections (6% in the fish oil group v. 11% in the control group) or who died (12% v. 15%), or in the length of hospital stay (25 v. 29 d). However, the proportion of patients in the fish oil group who were readmitted to intensive care (5%) was significantly lower (P < 0.05) than in the control group (17%). A group of patients also received the fish oil-containing emulsion for 2 d preoperatively. Here there were a number of very significant benefits. This group showed a significantly decreased need for mechanical ventilation (17% v. 31% in the control group; P < 0.05), a significantly shorter length of hospital stay (22 v. 29 d; P < 0.05), significantly less need for readmission to intensive care (5% v. 17%; P < 0.05) and significantly lower mortality $(3\% \ v. \ 15\%; \ P < 0.05)^{(70)}$. Another study revealed that intravenous infusion of a lipid emulsion containing 80% soyabean oil and 20% Omegaven into patients for 5 d following major gastrointestinal surgery accelerated normalisation of liver and pancreatic function compared with soyabean oil $alone^{(71)}$. Overall, there was no difference between the groups with respect to length of stay in the intensive care unit (ICU) or in hospital. However, in a subgroup of patients at risk of sepsis, there was a reduced ICU stay in patients receiving fish oil (4.0 v.)5.3 d in the control group; P = 0.01)⁽⁷¹⁾. In a recently published study, a mixed group of over 650 patients including about 230 post-surgical patients received parenteral nutrition including fish oil (Omegaven) for at least 3 d (mean 8.7 d); there was a significantly lower rate of infections (P < 0.0005), fewer complications (P < 0.005) and a shorter length of hospital stay (P = 0.05) in post-surgery patients receiving fish oil compared with those receiving the control emulsion⁽¹⁷⁾. These authors identified that infusion of about 0.15 g fish oil/kg per d decreased mean ICU stay from 8.7 to 5.3 d and hospital stay from 27.4 to 25.5 d. Thus, findings available from published studies in gastrointestinal surgical patients fairly clearly demonstrate clinical benefit from the inclusion of very long-chain n-3PUFA in the form of fish oil in parenteral nutrition regi-mens^(17,62,63,68,70,71). However, the study of Tsekos et al.⁽⁷⁰⁾ also demonstrates a greater benefit if these fatty acids are additionally provided pre-surgery, which, of course, is only possible in elective surgery. The greater benefit of pre-operative infusion of long-chain n-3 PUFA most likely relates to better incorporation of the fatty acids into leucocytes and other tissues.

Recently, a study using MCT–soyabean oil or Lipoplus in ICU patients having undergone abdominal aorta aneurysm repair surgery was published⁽⁷²⁾. There were no differences in glucose metabolism or in inflammatory markers. Clinical outcomes were not affected either, but there was a trend towards shorter ICU stay (1.6 v. 2.3 d) and shorter hospital stay (9.9 v. 11.3 d).

Thus, all three available fish oil-containing lipid emulsions have been used in adult post-surgery (mainly gastrointestinal) patients. No adverse effects of the use of fish oil have been reported, indicating that it is safe to use in such patients. The use of fish oil is associated with altered patterns of inflammatory eicosanoids and cytokines in post-gastrointestinal surgery patients, and immune function may be better maintained by fish oil in these patients. Two studies reported that the use of fish oil is associated with a trend towards reduced length of ICU stay and three studies reported that fish oil significantly reduced length of hospital stay (three more studies reported a trend to reduced length of hospital stay). Lack of significance in studies that reported favourable trends may be due to the small sample size of those studies. Perioperative administration of fish oil may be superior to post-operative. Taken together the studies in post-surgery patients present a fairly consistent and positive view of the efficacy of intravenous fish oil administration post-surgery. However, in these studies patients who would not normally require parenteral nutrition have frequently been included. Furthermore, the lengths of ICU and hospital stay reported in both control and fish oil groups are frequently much longer than typically seen in many clinical settings. Thus, although the data presently available are highly supportive of the inclusion of fish oil, translation of the findings to the real clinical situation requires further studies designed to mimic current clinical practice; clearly such studies need to be properly designed and adequately powered.

Studies of parenteral fish oil in critically ill patients

Septic patients who were intolerant of enteral nutrition received a standard soyabean oil-based emulsion or an emulsion containing fish oil (Omegaven) for $5^{(73)}$ or $10^{(74)}$ d. Blood leucocyte counts and serum C-reactive protein concentration tended to be lower, and production of leukotriene B₅ by stimulated neutrophils was significantly higher in patients receiving fish oil⁽⁷³⁾. Production of TNF- α , IL-1 β , IL-6, IL-8 and IL-10 by endotoxin-stimulated mononuclear cells did not increase during infusion of the fish oil-containing emulsion, whereas production of the four pro-inflammatory cytokines was markedly elevated during the first 2d of soyabean oil infusion⁽⁷⁴⁾. These studies establish that infusion of long-chain n-3 PUFA into patients with sepsis can modulate inflammatory mediator production and related inflammatory processes. It has been demonstrated that this might be associated with clinical improvements. Heller et al.⁽⁷⁵⁾ included patients with abdominal sepsis, multiple trauma and severe head injury in their study of parenteral n-3 PUFA (in the form of Omegaven) infusion. They found a significantly lower rate of infection and shorter lengths of ICU and hospital stay in those patients receiving more than 0.05 g fish oil/kg per d than in those receiving less than this. Mortality was significantly decreased in those patients who received more than 0.1 g fish oil/kg per d. The survival advantage was greater in some patient groups than others (severe head injury>multiple trauma>abdominal sepsis>nonabdominal sepsis>post-surgery), but small numbers of patients in some groups make the interpretation of these data difficult. Furthermore, this study was not controlled or blinded. Nevertheless, these recent data are strongly suggestive of genuine clinical benefit from the inclusion of

long-chain n-3 PUFA in parenteral nutrition regimens given to critically ill patients. This conclusion is in part supported by a study in patients with severe acute pancreatitis⁽⁷⁶⁾. The patients received soyabean oil or a mixture of soyabean oil and Omegaven for 5 d. Although there were no differences between the groups with regard to inflammatory markers, number of infections, or lengths of ICU (27.5 d in the control group v. 21.4 d in the fish oil group) and hospital stay, there was better gas exchange (P < 0.05) and a reduced requirement for continuous renal replacement therapy (P < 0.05) in those patients receiving fish oil. In contrast to the generally positive findings from the above studies, Friesecke et $al.^{(77)}$ reported no differences between MCT-soyabean oil and MCT-soyabean oil-Omegaven given over 7 d in medical ICU patients in several outcomes, including immune markers, inflammatory markers, bleeding, ventilation requirement, number of infections, length of ICU stay or mortality.

Thus, of the three available fish oil-containing lipid emulsions, only Omegaven has been used in critically ill adults. No adverse effects of the use of fish oil have been reported in these studies, indicating that it is safe to use in such patients. The influence of fish oil on inflammatory processes and on immune function in critically ill patients is not yet clear. Similarly, the impact of fish oil on clinical endpoints like infections, length of ICU and hospital stay, and mortality is not clear, since there are too few studies and those that are available (75-77) report contradictory findings or do not have a satisfactory design. One important factor, highlighted by the study of Heller et al., is the dose of fish oil required to influence clinical outcomes. Overall the data available are suggestive of some clinical benefit from the inclusion of long-chain n-3 PUFA in parenteral nutrition regimens given to critically ill patients. However, only limited studies have been published and the inconsistency of findings limits translation to the clinic. Thus, further studies are required; clearly such studies need to be properly designed and adequately powered.

Guidelines with regard to the use of fish oil in parenteral nutrition

Very recently new guidelines for parenteral nutrition were issued by the European Society for Clinical Nutrition and Metabolism (ESPEN)^(6,7). The guidelines on parenteral nutrition in surgery state '... there is some evidence that inclusion of n-3 fatty acids in parenteral nutrition may benefit organ function and reduce length of stay in patients undergoing major surgery or admitted to the surgical ICU. However, these trends will need to be substantiated in adequately powered randomised trials' and recommend 'The optimal parenteral nutrition regimen for critically ill surgical patients should probably include supplemental n-3fatty acids (Grade C)⁽⁶⁾. The guidelines on parenteral nutrition in intensive care recommend 'Addition of EPA and DHA to lipid emulsions has demonstrable effects on cell membranes and inflammatory processes (Grade B). Fish oil-enriched lipid emulsions probably decrease length of stay in critically ill patients (Grade B),⁽⁷⁾.

Use of fish oil in enteral nutrition in post-surgical and critically ill patients

Enteral formulas combining fish oil and arginine

Several enteral formulas using a combination of nutrients have been developed, typically including arginine, nucleotides and long-chain n-3 fatty acids. The majority of trials in surgical and critically ill patients have used IMPACT® (Nestle Nutrition, Gland, Switzerland) and a number of these studies reported immune and/or inflammatory outcomes (see Calder⁽⁷⁸⁾ for references). Most studies reporting circulating lymphocyte numbers and subsets, and circulating Ig concentrations showed little difference between IMPACT-treated patients and controls, although some studies reported benefits on phagocytosis, respiratory burst, lymphocyte proliferation, human leucocyte antigen-DR expression on monocytes and cytokine production⁽⁷⁸⁾. These effects could be due to any single specified nutrient (i.e. arginine, nucleotides, long-chain n-3 fatty acids) or to a combination of nutrients. Metaanalyses of controlled, randomised clinical studies using IMPACT or similar formulas have identified significant reductions in infections and length of hospital stay but these effects are more evident in surgical rather than critically ill patients⁽⁷⁹⁻⁸³⁾, and none of the meta-analyses shows a significant effect on mortality. Despite some clear statements to the contrary in the earlier meta-analyses⁽⁷⁹⁻⁸¹⁾, concern has been raised that these formulas may actually be detrimental in the seriously ill⁽⁸⁴⁻⁸⁶⁾. This is because some studies of these formulas in critically ill patients reported increased mortality $^{(79-81)}$. The source of the concern is the high arginine content, which is thought to drive excessive production of nitric oxide^(86,87).

Enteral formulas combining fish oil, borage oil and antioxidants

Work in experimental animals indicated a benefit of the combination of fish oil and borage oil (the latter contains γ -linolenic acid) with regard to lung inflammation and damage in endoxaemia^(56,57). $OxEPA^{\mathbb{R}}$ (Abbott Nutrition, Columbus, OH, USA) is an enteral formula that contains a high level of fat (55% of energy), with the fat being a combination of 32% canola (i.e. low-erucic acid rapeseed oil), 20% fish oil, 20% borage oil, 25% MCT and 3% soyabean phospholipid. OxEPA also contains β -carotene, taurine and carnitine, and more vitamin C and vitamin E than standard enteral formulas. OxEPA was trialled for 7 d in patients with acute respiratory distress syndrome^(88,89). By 4 d of treatment the numbers of total leucocytes and of neutrophils in the alveolar fluid declined significantly in the *n*-3 fatty acid group and were lower than in controls⁽⁸⁸⁾. Alveolar fluid IL-8 was lower in the n-3 fatty acid group compared with controls and LTB₄ and TNF- α tended to be lower⁽⁸⁹⁾. Arterial oxygenation and gas exchange were also improved and the n-3 fatty acid-treated patients had a decreased requirement for supplemental oxygen, decreased time on ventilation support (11 v. 16·3 d; P = 0.011) and a shorter length of stay in intensive care (12.8 v. 17.5 d;P = 0.016⁽⁸⁸⁾. The total length of hospital stay tended to be shorter in the n-3 fatty acid group (29.4 v. 34.6 d; NS)

NS Proceedings of the Nutrition Society

and fewer patients developed new organ failure (4/51 v. 13/47; P = 0.015)⁽⁸⁸⁾. Mortality was 12% in the *n*-3 fatty acid group and 19% in the control group, but this difference was not statistically significant⁽⁸⁸⁾. Since OxEPA not only provided fish and borage oils but also MCT, β -carotene, taurine and carnitine, and more vitamin C and vitamin E than the control formula, it is not possible to ascribe the benefits to any particular nutrient, although the antiinflammatory effects seen are consistent with those of fish oil n-3 PUFA. Two more recent studies also report benefits from OxEPA in acutely ill patients^(90,91). In one of these studies patients with acute lung injury received a control formula or OxEPA for 14d⁽⁹⁰⁾. By days 4 and 7 patients receiving OxEPA showed improved oxygenation, a reduction in length of ventilation (160 v. 167 h; P < 0.03) and a shorter ICU stay (12.8 v. 17.5 d; P = 0.016), although there was no difference between the groups in mortality $\overset{(90)}{}$. In the second trial, OxEPA was used in ventilated patients with severe sepsis and septic shock⁽⁹¹⁾. Patients receiving OxEPA had better oxygenation, more ventilator-free days (13.4 v. 5.8 d; P < 0.001), fewer days in the ICU (4.6 v. 10.8 d; P < 0.001), less development of new organ dysfunctions (38% v. 81%; P<0.001) and reduced 28-d mortality (33% v. 52%; P = 0.037).

These three studies were recently combined in a metaanalysis⁽⁵⁾. This demonstrated a significant overall benefit of OxEPA *v*. control on requirement for ventilation (P < 0.0001), ICU stay (P < 0.0001), new organ failures (P < 0.0001) and mortality (P = 0.001).

Guidelines with regard to the use of fish oil in enteral nutrition

The ESPEN guidelines for use of enteral nutrition in surgical patients recommend 'Use enteral nutrition with immuno-modulating substrates (arginine, n-3 fatty acids and nucleotides) perioperatively for patients undergoing major neck surgery for cancer, undergoing major abdominal cancer surgery or after severe trauma (Grade A)'⁽⁹²⁾. The guidelines for use of enteral nutrition in intensive care recommend 'Immune modulating formulae (formulae enriched with arginine, nucleotides and n-3 fatty acids) are superior to standard enteral formulae in elective upper gastrointestinal surgical patients (Grade A), in patients with mild sepsis (Grade B), in patients with trauma (Grade A), in patients with acute respiratory distress syndrome (formulae containing n-3 fatty acids and antioxidants) (Grade B)'⁽⁹³⁾. The latter recommendation was based upon a single study⁽⁸⁸⁾, and as mentioned earlier two further supportive studies have now been published^(90,91). The guidelines for enteral nutrition in intensive care also recommend 'ICU patients with very severe illness who do not tolerate more than 700 ml enteral formula per day should not receive an immune-modulating formula enriched with arginine, nucleotides and n-3 fatty acids (Grade B)'⁽⁹³⁾

Summary and conclusions

Lipids traditionally used in parenteral nutrition are based on n-6 PUFA-rich vegetable oils like soyabean oil. This may not be optimal because it may present an excessive supply of linoleic acid. Alternatives to use of soyabean oil include its partial replacement by MCT, olive oil or fish oil, either alone or in combination. Lipid emulsions containing fish oil are well tolerated and without adverse effects in a wide range of adult patients. Fish oil-containing lipid emulsions have been used in parenteral nutrition in adult patients post-surgery (mainly gastrointestinal). This has been associated with alterations in patterns of inflammatory mediators and in immune function and, in some studies, a reduction in length of ICU and hospital stay. Perioperative administration of fish oil may be superior to post-operative. Parenteral fish oil has been used in critically ill adults. Here the influence on inflammatory processes, immune function and clinical endpoints is not clear, since there are too few studies and those that are available report contradictory findings. Fish oil is included in combination with other nutrients in various enteral formulas. In post-surgical patients and in those with mild sepsis or trauma, there is clinical benefit from a formula including fish oil and arginine. A formula including fish oil, borage oil and antioxidants has demonstrated marked benefits on gas exchange, ventilation requirement, new organ failures, ICU stay and mortality in patients with acute respiratory distress syndrome, acute lung injury or severe sepsis.

Acknowledgements

The author has received speaking fees from B. Braun, Fresenius Kabi, Baxter Healthcare, Abbott Nutrition and Nestle, participated in the Baxter Healthcare Global Advisory Board in 2008, and has received research funding from B. Braun.

References

- 1. Calder PC (2006) Use of fish oil in parenteral nutrition: rationale and reality. *Proc Nutr Soc* **65**, 264–277.
- Calder PC (2007) Immunonutrition in surgical and critically ill patients. Br J Nutr 98, S133–S139.
- 3. Calder PC (2009) Rationale for using new lipid emulsions in parenteral nutrition and a review of the trials performed in adults. *Proc Nutr Soc* **68**, 252–260.
- 4. Liang B, Wang S, Ye YJ *et al.* (2008) Impact of postoperative omega-3 fatty acid-supplemented parenteral nutrition on clinical outcomes and immunomodulations in colorectal cancer patients. *World J Gastroenterol* **14**, 2434– 2439.
- 5. Pontes-Arruda A, Demichele S, Seth A & Singer P (2008) The use of an inflammation-modulating diet in patients with acute lung injury or acute respiratory distress syndrome: a meta-analysis of outcome data. *J Parenter Enteral Nutr* **32**, 596–605.
- Braga M, Ljungqvist O, Soeters P *et al.* (2009) ESPEN guidelines on parenteral nutrition: surgery. *Clin Nutr* 28, 378–386.
- 7. Singer P, Berger MM, Van den Berghe G *et al.* (2009) ESPEN guidelines on parenteral nutrition: intensive care. *Clin Nutr* **28**, 387–400.
- Calder PC & Burdge GC (2004) Fatty acids. In Bioactive Lipids, pp. 1–36 [A Nicolaou and G Kafatos, editors]. Bridgwater, Somerset: The Oily Press.

- 9. Gurr MI, Harwood JL & Frayn KN (2002) *Lipid Biochemistry*. Oxford: Blackwell Science.
- Sprecher H (2002) The roles of anabolic and catabolic reactions in the synthesis and recycling of polyunsaturated fatty acids. *Prostaglandins Leukot Essent Fatty Acids* 67, 79–83.
- Yaqoob P & Calder PC (2007) Fatty acids and immune function: new insights into mechanisms. Br J Nutr 98, S41–S45.
- 12. Calder PC (2008) The relationship between the fatty acid composition of immune cells and their function. *Prostaglandins Leukot Essent Fatty Acids* **79**, 101–108.
- 13. Edgren B & Wrtelind A (1963) The theoretical background of the intravenous nutrition with fat emulsions. *Nutr Dieta Eur Rev Nutr Diet* **13**, 364–386.
- Hallberg D, Schuberth O & Wretlind A (1966) Experimental and clinical studies with fat emulsion for intravenous nutrition. *Nutr Dieta Eur Rev Nutr Diet* 8, 245–281.
- Wretlind A (1972) Complete intravenous nutrition. Theoretical and experimental background. *Nutr Metab* 14, Suppl., 1–57.
- Heyland DK, MacDonald S, Keefe L & Drover JW (1998) Total parenteral nutrition in the critically ill patient: a metaanalysis. J Am Med Assoc 280, 2013–2019.
- Koch T & Heller AR (2005) Auswirkungen einer parenteralen ernahrung mit n-3-fettsauren auf das therapieergebnis – eine multizentrische analyse bei 661 patienten. Akt Ernahrungs 30, 15–22.
- Calder PC, Sherrington EJ, Askanazi J & Newsholme EA (1994) Inhibition of lymphocyte proliferation *in vitro* by two lipid emulsions with different fatty acid compositions. *Clin Nutr* 13, 69–74.
- 19. Monson JRT, Sedman PC, Ramsden CW *et al.* (1988) Total parenteral nutrition adversely influences tumour-directed cellular cytotoxic responses in patients with gastrointestinal cancer. *Eur J Surg Oncol* **14**, 435–443.
- 20. Battistella FD, Widergren JT, Anderson JT *et al.* (1997) A prospective, randomized trial of intravenous fat emulsion administration in trauma victims requiring total parenteral nutrition. *J Trauma* **43**, 52–58.
- 21. Furukawa K, Yamamori H, Takagi K *et al.* (2002) Influences of soybean oil emulsion on stress response and cell-mediated immune function in moderately or severely stressed patients. *Nutrition* **18**, 235–240.
- 22. Dionigi P, Dionigi R, Prati U *et al.* (1985) Effect of Intralipid[®] on some immunological parameters and leukocyte functions in patients with esophageal and gastric cancer. *Clin Nutr* **4**, 229–234.
- Gogos CA, Kalfarentzos FE & Zoumbos NC (1990) Effect of different types of total parenteral nutrition on T-lymphocyte subpopulations and NK cells. *Am J Clin Nutr* 51, 119–122.
- 24. Sedman PC, Somers SS, Ramsden CW *et al.* (1991) Effects of different lipid emulsions on lymphocyte function during total parenteral nutrition. *Br J Surg* **78**, 1396–1399.
- 25. Lenssen P, Bruemmer BA, Bowden RA *et al.* (1998) Intravenous lipid dose and incidence of bacteremia and fungemia in patients undergoing bone marrow transplantation. *Am J Clin Nutr* **67**, 927–933.
- Ulrich H, McCarthy Pastores S *et al.* (1996) Parenteral use of medium-chain triglycerides: a reappraisal. *Nutrition* 12, 231– 238.
- 27. Adolph M (1999) Lipid emulsions in parenteral nutrition. Ann Nutr Metab 43, 1–13.
- Sala-Vila A, Barbosa VM & Calder PC (2007) Olive oil in parenteral nutrition. *Curr Opin Clin Nutr Metab Care* 10, 165–174.
- 29. Furst P & Kuhn KS (2000) Fish oil emulsions: what benefits can they bring? *Clin Nutr* **19**, 7–14.

- Adolph M (2001) Lipid emulsions in total parenteral nutrition – state of the art and future perspectives. *Clin Nutr* 20, Suppl. 4, 11–14.
- Grimble R (2005) Fatty acid profile of modern lipid emulsions: scientific considerations for creating the ideal composition. *Clin Nutr Suppl* 1, 9–15.
- Grimm H (2005) A balanced lipid emulsion a new concept in parenteral nutrition. *Clin Nutr Suppl* 1, 25–30.
- Calder PC (2004) N-3 fatty acids and cardiovascular disease: evidence explained and mechanisms explored. *Clin Sci* 107, 1–11.
- 34. Kris-Etherton PM, Harris WS, Appel LJ, American Heart Association Nutrition Committee (2002) Fish consumption, fish oil, omega-3 fatty acids, and cardiovascular disease. *Circulation* **106**, 2747–2757.
- 35. Bucher HC, Hengstler P, Schindler C & Meier G (2002) N-3 polyunsaturated fatty acids in coronary heart disease: a meta-analysis of randomized controlled trials. *Am J Med* **112**, 298–304.
- Studer M, Briel M, Leimenstoll B *et al.* (2005) Effect of different antilipidemic agents and diets on mortality: a systematic review. *Arch Intern Med* 165, 725–730.
- 37. Wang C, Harris WS, Chung M *et al.* (2006) n-3 Fatty acids from fish or fish-oil supplements, but not alpha-linolenic acid, benefit cardiovascular disease outcomes in primary- and secondary-prevention studies: a systematic review. *Am J Clin Nutr* **84**, 5–17.
- Calder PC (2006) N-3 polyunsaturated fatty acids, inflammation, and inflammatory diseases. *Am J Clin Nutr* 83, 1505S–1519S.
- 39. Tagawa H, Shimokawa H, Tagawa T *et al.* (1999) Long-term treatment with eicosapentaenoic acid augments both nitric oxide-mediated and non-nitric oxide-mediated endothelium-dependent forearm vasodilatation in patients with coronary artery disease. *J Cardiovasc Pharmacol* **33**, 633–640.
- Calder PC (2007) Immunomodulation by omega-3 fatty acids. Prostaglandins Leukot Essent Fatty Acids 77, 327– 335.
- Calder PC (2008) Danone Chair Monograph: Omega-3 Fatty Acids – The Good Oil? Brussels: Institut Danone.
- 42. Utsunomiya T, Chavali SR, Zhong WW & Forse RA (1994) Effects of continuous tube feeding of dietary fat emulsions on eicosanoid production and on fatty acid composition during an acute septic shock in rats. *Biochim Biophys Acta* 1214, 333–339.
- Sane S, Baba M, Kusano C *et al.* (2000) Eicosapentaenoic acid reduces pulmonary edema in endotoxemic rats. *J Surg Res* 93, 21–27.
- 44. Sadeghi S, Wallace FA & Calder PC (1999) Dietary lipids modify the cytokine response to bacterial lipopolysaccharide in mice. *Immunology* **96**, 404–410.
- 45. Barton RG, Wells CL, Carlson A *et al.* (1991) Dietary omega-3 fatty acids decrease mortality and Kupffer cell prostaglandin E2 production in a rat model of chronic sepsis. *J Trauma* **31**, 768–774.
- 46. Rayon JI, Carver JD, Wyble LE *et al.* (1997) The fatty acid composition of maternal diet affects lung prostaglandin E2 levels and survival from group B Streptococcal sepsis in neonatal rat pups. *J Nutr* **127**, 1989–1992.
- Lanza-Jacoby S, Flynn JT & Miller S (2001) Parenteral supplementation with a fish oil emulsion prolongs survival and improves lymphocyte function during sepsis. *Nutrition* 17, 112–116.
- 48. Mulrooney HM & Grimble RF (1993) Influence of butter and of corn, coconut and fish oils on the effects of recombinant human tumour necrosis factor- α in rats. *Clin Sci* **84**, 105–112.

- 49. Pomposelli J, Mascioli EA, Bistrian BR & Flores SM (1990) Attenuation of the febrile response in guinea pigs by fish oil enriched diets. *J Parenter Enteral Nutr* **13**, 136–140.
- Pomposelli JJ, Flores EA, Blackburn G *et al.* (1991) Diets enriched with n-3 fatty acids ameliorate lactic acidosis by improving endotoxin-induced tissue hypoperfusion in guinea pigs. *Ann Surg* 213, 166–176.
- 51. Teo TC, Selleck KM, Wan JMF *et al.* (1991) Long-term feeding with structured lipid composed of medium-chain and n-3 fatty acids ameliorates endotoxic shock in guinea-pigs. *Metabolism* **40**, 1152–1159.
- Murray MJ, Kumar M, Gregory TJ et al. (1995) Select dietary fatty acids attenuate cardiopulmonary dysfunction during acute lung injury in pigs. Am J Physiol 269, H2090–H2097.
- 53. Murray MJ, Svinger BA, Holman RT & Yaksh TL (1991) Effects of a fish oil diet on pig's cardiopulmonary response to bacteremia. *J Parenter Enteral Nutr* **15**, 152–158.
- Murray MJ, Svinger BA, Yaksh TL & Holman RT (1993) Effects of endotoxin on pigs prefed omega-3 vs. omega-6 fatty acids-enriched diets. *Am J Physiol* 265, E920–E927.
- 55. Murray MJ, Kanazi G, Moukabary K *et al.* (2000) Effects of eicosapentaenoic and γ-linolenic acids (dietary lipids) on pulmonary surfactant composition and function during porcine endotoxemia. *Chest* **117**, 1720–1727.
- 56. Mancuso P, Whelan J, DeMichele SJ *et al.* (1997) Effects of eicosapentaenoic and gamma-linolenic acid on lung permeability and alveolar macrophage eicosanoid synthesis in endotoxic rats. *Crit Care Med* 25, 523–532.
- 57. Mancuso P, Whelan J, DeMichele SJ *et al.* (1997) Dietary fish oil and fish and borage oil suppress intrapulmonary proinflammatory eicosanoids biosynthesis and attenuate pulmonary neutrophil accumulation in endotoxic rats. *Crit Care Med* 25, 1198–1206.
- Mascioli EA, Leader L, Flores E *et al.* (1988) Enhanced survival to endotoxin in guinea pigs fed iv fish oil emulsion. *Lipids* 23, 623–625.
- 59. Mascioli EA, Iwasa Y, Trimbo S *et al.* (1989) Endotoxin challenge after menhaden oil diet: effects on survival of guinea pigs. *Am J Clin Nutr* **49**, 277–282.
- Johnson JA, Griswold JA, Muakkassa FF *et al.* (1993) Essential fatty acids influence survival in stress. *J Trauma* 35, 128–131.
- 61. Morlion BJ, Torwesten E, Lessire A *et al.* (1996) The effect of parenteral fish oil on leukocyte membrane fatty acid composition and leukotriene-synthesizing capacity in post-operative trauma. *Metabolism* **45**, 1208–1213.
- 62. Grimm H, Mertes N, Goeters C *et al.* (2006) Improved fatty acid and leukotriene pattern with a novel lipid emulsion in surgical patients. *Eur J Nutr* **45**, 55–60.
- Wichmann MW, Thul P, Czarnetzki HD *et al.* (2007) Evaluation of clinical safety and beneficial effects of a fish oil containing lipid emulsion (Lipoplus, MLF541): data from a prospective, randomized, multicenter trial. *Crit Care Med* 35, 700–706.
- 64. Senkal M, Geier B, Hannemann M et al. (2007) Supplementation of omega-3 fatty acids in parenteral nutrition beneficially alters phospholipid fatty acid pattern. J Parenter Enteral Nutr 31, 12–17.
- 65. Roulet M, Frascarolo P, Pilet M & Chapuis G (1997) Effects of intravenously infused fish oil on platelet fatty acid phospholipid composition and on platelet function in postoperative trauma. J Parenter Enter Nutr 21, 296–301.
- 66. Wachtler P, Konig W, Senkal M *et al.* (1997) Influence of a total parenteral nutrition enriched with ω-3 fatty acids on leukotriene synthesis of peripheral leukocytes and systemic cytokine levels in patients with major surgery. *J Trauma* 42, 191–198.

- 67. Koller M, Senkal M, Kemen M et al. (2003) Impact of omega-3 fatty acid enriched TPN on leukotriene synthesis by leukocytes after major surgery. Clin Nutr 22, 59–64.
- Weiss G, Meyer F, Matthies B *et al.* (2002) Immunomodulation by perioperative administration of n-3 fatty acids. *Br J Nutr* 87, S89–S94.
- 69. Schauder P, Rohn U, Schafer G *et al.* (2002) Impact of fish oil enriched total parenteral nutrition on DNA synthesis, cytokine release and receptor expression by lymphocytes in the postoperative period. *Br J Nutr* **87**, S103–S110.
- Tsekos E, Reuter C, Stehle P & Boeden G (2004) Perioperative administration of parenteral fish oil supplements in a routine clinical setting improves patient outcome after major abdominal surgery. *Clin Nutr* 23, 325–330.
- Heller AR, Rossel T, Gottschlich B *et al.* (2004) Omega-3 fatty acids improve liver and pancreas function in postoperative cancer patients. *Int J Cancer* 111, 611–616.
- Berger MM, Tappy L, Revelly JP *et al.* (2008) Fish oil after abdominal aorta aneurysm surgery. *European J Clin Nutr* 62, 1116–1122.
- 73. Mayer K, Fegbeutel C, Hattar K *et al.* (2003) W-3 vs. w-6 lipid emulsions exert differential influence on neutrophils in septic shock patients: impact on plasma fatty acids and lipid mediator generation. *Intensive Care Med* 29, 1472–1481.
- Mayer K, Gokorsch S, Fegbeutel C *et al.* (2003) Parenteral nutrition with fish oil modulates cytokine response in patients with sepsis. *Am J Respir Crit Care Med* 167, 1321– 1328.
- Heller AR, Rössler S, Litz RJ *et al.* (2006) Omega-3 fatty acids improve the diagnosis-related clinical outcome. *Crit Care Med* 34, 972–979.
- 76. Wang X, Li W, Li N & Li J (2008) Omega-3 fatty acids-supplemented parenteral nutrition decreases hyperinflammatory response and attenuates systemic disease sequelae in severe acute pancreatitis: a randomized and controlled study. J Parenter Enteral Nutr 32, 236–241.
- Friesecke S, Lotze C, Köhler J *et al.* (2008) Fish oil supplementation in the parenteral nutrition of critically ill medical patients: a randomised controlled trial. *Intensive Care Med* 34, 1411–1420.
- Calder PC (2003) Long-chain n-3 fatty acids and inflammation: potential application in surgical and trauma patients. *Braz J Med Biol Res* 36, 433–446.
- Beale RJ, Bryg DJ & Bihari DJ (1999) Immunonutrition in the critically ill: A systematic review of clinical outcome. *Crit Care Med* 27, 2799–2805.
- Heys SD, Walker LG, Smith I & Eremin O (1999) Enteral nutritional supplementation with key nutrients in patients with critical illness and cancer – a meta-analysis of randomized controlled clinical trials. *Ann Surg* 229, 467–477.
- Heyland DK, Novak F, Drover JW *et al.* (2001) Should immunonutrition become routine in critically ill patients? A systematic review of the evidence. *JAMA* 286, 944–953.
- Montejo JC, Zarazaga A, López-Martínez J *et al.* (2003) Immunonutrition in the intensive care unit. A systematic review and consensus statement. *Clin Nutr* 22, 221–233.
- Waitzberg DL, Saito H, Plank LD *et al.* (2006) Postsurgical infections are reduced with specialized nutrition support. *World J Surg* 30, 1592–1604.
- Heyland DK, Dhaliwal R, Drover JW *et al.* (2003) Canadian clinical practice guidelines for nutrition support in mechanically ventilated, critically ill adult patients. *J Parenter Enteral Nutr* 27, 355–373.
- 85. Heyland DK, Samis A (2003) Does immunonutrition in patients with sepsis do more harm than good? *Intensive Care Med* **29**, 669–671.

- Suchner U, Heyland DK & Peter K (2002) Immunemodulatory actions of arginine in the critically ill. *Br J Nutr* 87, S121–S132.
- 87. Zhou M & Martindale RG (2007) Arginine in the critical care setting. *J Nutr* **137**, 1687S–1692S.
- 88. Gadek JE, DeMichele SJ, Karlstad MD *et al.* of the Enteral Nutrition in ARDS Study Group (1999) Effect of enteral feeding with eicosapentaenoic acid, γ-linolenic acid, and antioxidants in patients with acute respiratory distress syndrome. *Crit Care Med* 27, 1409–1420.
- 89. Pacht ER, DeMichele SJ, Nelson JL et al. (2003) Enteral nutrition with eicosapentaenoic acid, gammalinolenic acid, and antioxidants reduces alveolar inflammatory mediators and protein influx in patients with acute respiratory distress syndrome. Crit Care Med 31, 491– 500.
- 90. Singer P, Theilla M, Fisher H *et al.* (2006) Benefit of an enteral diet enriched with eicosapentaenoic acid and gammalinolenic acid in ventilated patients with acute lung injury. *Crit Care Med* 34, 1033–1038.
- 91. Pontes-Arruda A, Aragão AM & Albuquerque JD (2006) Effects of enteral feeding with eicosapentaenoic acid, gamma-linolenic acid, and antioxidants in mechanically ventilated patients with severe sepsis and septic shock. *Crit Care Med* 34, 2325–2333.
- 92. Weimann A, Braga M, Harsanyi L *et al.* ESPEN (European Society for Parenteral and Enteral Nutrition) (2006) ESPEN guidelines on enteral nutrition: surgery including organ transplantation. *Clin Nutr* 25, 224–244.
- Kreymann KG, Berger MM, Deutz NE *et al.* (2006) ESPEN guidelines on enteral nutrition: intensive care. *Clin Nutr* 25, 210–223.