An update on diet and nutritional factors in systemic lupus erythematosus management

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Abstract

Systemic lupus erythematosus (SLE) is a chronic inflammatory and autoimmune disease characterised by multiple organ involvement and a large number of complications. SLE management remains complicated owing to the biological heterogeneity between patients and the lack of safe and specific targeted therapies. There is evidence that dietary factors can contribute to the geoepidemiology of autoimmune diseases such as SLE. Thus, diet therapy could be a promising approach in SLE owing to both its potential prophylactic effects, without the side effects of classical pharmacology, and its contribution to reducing co-morbidities and improving quality of life in patients with SLE. However, the question arises as to whether nutrients could ameliorate or exacerbate SLE and how they could modulate inflammation and immune function at a molecular level. The present review summarises preclinical and clinical experiences to provide the reader with an update of the positive and negative aspects of macro- and micronutrients and other nutritional factors, including dietary phenols, on SLE, focusing on the mechanisms of action involved.

Key words: Diet: Immunomodulation: Lupus: Nutrients: Systemic lupus erythematosus

Introduction

Systemic lupus erythematosus (SLE) can be defined as a chronic inflammatory and autoimmune disease that can affect multiple organ systems, including skin, joints, kidneys and the brain, among others⁽¹⁾. The clinical heterogeneity of this remarkable and challenging disorder has required the establishment of eleven criteria with four needed for the formal diagnosis of SLE⁽²⁾. The action of pathogenic factors results in the generation of autoantibodies, immune complexes, autoreactive or inflammatory T cells and inflammatory cytokines that may initiate and amplify inflammation and damage to various organs, contributing to the clinical manifestations of SLE. Moreover, heritable, hormonal and environmental factors contribute to the expression of organ damage. SLE is characterised by a deposition of immune complexes, formed in large amounts as antinuclear antibodies bind to the abundant nuclear material in blood and tissues, along with disturbances in both innate and adaptive immunity manifest by disorders in cytokines, apoptotic cell clearance, B-cell immunity and T-cell signalling^(3,4).

It is also characterised by its clinical and pathogenic complexity, difficult diagnosis and the high number of complications that can affect the patients' quality of life. The high morbidity and mortality associated with patients with SLE may be related to late diagnosis, problems in access to care, less effective treatments and poor adherence to therapeutic regimens⁽⁵⁾.

SLE incidence has been estimated to range between 1 and 10 cases per 100 000 individuals per year, and the prevalence has been reported to range between 20 and 150 cases per 100 000 individuals worldwide with large regional and ethnic variations⁽⁶⁾. SLE is a disease that can affect both sexes, but more than 90% of new patients presenting with SLE are women in the childbearing years. In addition, although the disease may begin at any age, it appears most often at the end of the patients' second decade of life and at the beginning of the third decade⁽⁷⁾. SLE affects multiple systems and its presentation and course are highly variable, ranging from indolent to fulminant. The most common clinical manifestations include fatigue, loss of appetite and weight, cutaneous lesions (mainly malar rash), arthritis, serositis (pleuritis and/or pericarditis), renal or central nervous system involvement and haematological manifestations (cytopenias) associated with several autoantibodies, particularly antinuclear antibodies⁽⁵⁾.

Additionally, patients with SLE show an increased risk of atherosclerosis and vascular events, which contribute to increase the morbidity and mortality of these patients. In SLE, autoantibodies and cytokines are able to modulate and decrease lipoprotein lipase activity, a key enzyme in lipid metabolism, producing the 'lupus pattern' of dyslipoproteinaemia characterised by elevated levels of VLDL and TAG and low HDL-cholesterol levels, which are directly correlated with SLE disease activity index (SLEDAI) scores⁽⁸⁾. The mechanisms underlying this enhanced risk are still not clear, but some

- Abbreviations: 2-OH, 2-hydroxyestrone; 16α-OH, 16α-hydroxyestrone; Akt, protein kinase B; BAFF, B-cell activating factor; dsDNA, double-stranded DNA; EGCG, epigallocatechin gallate; ER, oestrogen receptor; EVOO, extra virgin olive oil; FOXP3, forkhead box P3; I3C, indole-3-carbinol; IFN, interferon; LPS, lipopolysaccharide; MRL, Murphy Roths large; Nrf-2, nuclear factor E2-related factor 2; NZB/W, New Zealand black/white; ppm, parts per million; SLE, systemic lupus erythematosus; Th, T-helper; Treg, regulatory T-cell.
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theories include excess of monocyte activation, dysregulation of the complement system, oxidative stress and production of different antibodies to endothelial cells, anti-atherogenic HDL, anti-lipoprotein lipase and oxidised LDL among others⁽⁴⁾.

Systemic lupus erythematosus aetiology

The aetiology of SLE is not fully known. Genetics is thought to play a crucial role in the development of SLE, and multiple genes that contribute to the predisposition and susceptibility to SLE have been identified, including IRF5, STAT4, osteopontin, IRAK1, TREX1 and TLR8, and genes related to interferon (IFN) production or implicated in the signalling mechanisms mediated by T (PTPN22, TNFSF4, PDCD1) or B (BANK1, BLK, LYN) cells⁽⁹⁾. In addition, recent studies have shown that several regions located in the major histocompatibility complex as well as BsmI and FokI polymorphisms contribute to increased SLE risk^(10,11). Environmental factors, including extreme stress, exposure to UV light, smoking, infections, administration of certain drugs (some antidepressants and antibiotics) and hormonal factors (oestrogen) can also contribute to the disease progression^(12–14). Nevertheless, over recent years, multiple lines of evidence have supported a major role of specific dietary factors, including vitamins, mineral elements, fatty acids and polyphenols, in the modulation of immune responses. Thus, an inadequate diet could constitute an important risk factor in SLE epidemiology⁽¹⁵⁻¹⁸⁾.

The pathological hallmark of SLE is altered immune response with loss of B and T lymphocyte self-tolerance resulting in hyperactivity, autoantibody production (antinuclear antibodies, anti-double-stranded DNA (anti-dsDNA), anti-Ro, anti-histone, anti-Smith, anti-ribonucleoprotein and anti-ribosomal P protein, among others), aberrant immune complex formation and generation of a systemic inflammatory response in which multiple organs are involved: kidneys, heart, joints, skin, lungs, blood vessels, liver and nervous system⁽¹⁹⁾. The course of the disease is unpredictable, with periods of exacerbations and remissions⁽³⁾. Autoantibodies, immune complexes and an imbalance of T-helper-cell subsets (Th1/Th2/Th17) and regulatory T-cells (Tregs) contribute to tissue damage and could be responsible for an increased proinflammatory response, especially in the active form of the disease⁽²⁰⁾. In particular, patients with active SLE exhibit high cytokine levels, including IFN-y, TNF, IL-4, IL-6, IL-10, IL-12, IL-17 and IL-18 in serum and plasma; by contrast, IL-2 levels are lower in comparison with healthy controls⁽²¹⁾. In addition, recent research has associated SLE development with changes in the dendritic cell compartment $^{(22)}$. Likewise, it has been suggested that $\gamma\delta$ T cells may be involved in the regulation of SLE owing to their functions in cytokine secretion, antigen presentation and supporting B cells in antibody production⁽²³⁾. On the other hand, many studies have demonstrated quantitative and/or qualitative defects of Tregs (phenotype $CD4^+$ CD25 high forkhead box P3 (FOXP3)⁺) in patients with SLE as well as a reduction in the number of natural killer (NK) cells. Moreover, an enhanced Th17 cell response correlates with disease activity in patients with SLE, suggesting a role for IL-17 in the pathogenesis of the disease owing to its ability to amplify local inflammation by recruiting innate immune system cells and to stimulate the adaptive immune response in conjunction with B-cell activating factor (BAFF)^(24,25). The molecular signalling pathways involved in SLE include NF-KB, FOXP3, mitogen-activated protein kinase (MAPK). Janus kinase and signal transducer and activator of transcription (JAK/STAT) and, more recently, NLR family pyrin domain-containing 3 (NLRP3) inflammasome and nuclear factor E2-related factor 2 (Nrf-2) pathways⁽²⁶⁻³⁰⁾. The processes of adhesion, extravasation and subsequent activation of neutrophils in SLE are due to increases in the expression of intercellular adhesion molecule-1 (ICAM-1), endothelial adhesins, selectins and integrins. The secretion of proteases, reactive oxygen species and myeloperoxidase by the neutrophils then contributes to the cell damage⁽³¹⁾. Finally, an increased apoptotic burden that determines the recognition of apoptotic-derived autoantigens and hyperactivation of innate and adaptive immune system cells has been described in human SLE and murine models of the disease⁽³²⁾.

Aim of systemic lupus erythematosus treatment

Nowadays, the overall aim of SLE therapy is to control disease activity. Nevertheless, SLE management remains complicated owing to the biological heterogeneity between patients and the lack of safe and specific targeted therapies. Thus, the search for new therapeutic targets and strategies that can act more selectively on certain routes or biological processes and improve the course of disease or reverse the outbreak phase without generating collateral damage to unaffected tissues and organs is the pillar underlying current research in SLE.

Typical SLE management includes the use of antimalarial drugs (mainly hydroxychloroquine and chloroquine), immunosuppressive agents, biological agents and some adjunctive therapies following international recommendations⁽³³⁾. Mild activity can be managed with non-steroidal anti-inflammatory drugs or low-dose glucocorticoids, but more severe manifestations require more advanced treatment⁽³³⁻³⁵⁾. Glucocorticoids act as anti-inflammatory and immunosuppressive agents, but they are associated with several severe side effects including osteoporosis, cataracts, hyperglycaemia and cognitive impairment, among others^(36,37). In particular, glucocorticoid therapy may increase the predisposition to obesity, leading to increased levels of proinflammatory cytokines that exacerbate the SLE symptoms and increase the risk of developing diabetes mellitus, hypertension and CHD symptoms. Moreover, the chronic use of corticosteroids in SLE is associated to an increase of total plasma cholesterol and its fractions (LDL and HDL) levels, and also TAG⁽⁸⁾. On the other hand, antimalarial drugs have shown beneficial effect on lipids and diabetes control in patients with SLE⁽³⁸⁻⁴⁰⁾. If glucocorticoid administration cannot be reduced or discontinued, the administration of immunosuppressive diseasemodifying anti-rheumatic drugs, commonly mycophenolate mofetil, cyclophosphamide, azathioprine, calcineurin inhibitors (particularly tacrolimus and cyclosporine A) and methotrexate, is recommended, with a tailored treatment regimen for every patient^(41,42). However, the continuous use of immunosuppressive drugs may increase susceptibility to infections and gastrointestinal symptoms.

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The increased knowledge about the aetiopathogenesis of SLE has enabled the use of biological agents specifically targeting the different pathways implicated in the disease⁽⁴³⁾. Since B cell deregulation is one of the SLE hallmarks, B-cell-targeted therapies have become an important focus of SLE research. At present, belimumab, a monoclonal antibody that blocks BAFF, is the only approved biological drug for SLE, but other B-cell-targeted agents, such as rituximab, epratuzumab, blisibimod and tabalumab, are currently undergoing clinical evaluation, which has produced interesting results supporting their potential role in SLE treatment⁽⁴⁴⁻⁴⁹⁾. Abatacept, which acts by blocking interactions between T and B cells, has shown efficacy in lupus mouse models, but controversial results have been described in human controlled trials^(50,51). Moreover, high serum levels of IFN in patients with SLE have been correlated with SLE disease activity. Recently, anti-IFN therapies, specifically sifalimumab, rontalizumab, anifrolumab and AMG 811, have been investigated with promising results, but several hurdles in their development must still be overcome and further studies are requested to fully determine their effects in SLE⁽⁵²⁾.

Thus, although pharmacological treatment for SLE has improved during the last decade, and many potential new agents are in development, SLE and its treatments contribute to increased mortality rates and the outcome remains unoptimistic in a considerable percentage of patients⁽⁵³⁾.

Diet therapy and nutrients in systemic lupus erythematosus

Nutritional therapy, including diet modification and the use of nutritional supplements, could be a promising way to approach SLE owing to both its potential prophylactic effects, without the side effects of the classic pharmacological therapy, and its possible contribution to reducing co-morbidities and improving the quality of life of patients with SLE⁽⁵⁴⁾.

Diet quality in patients with SLE is important since these patients, in addition to being at higher risk of CVD, are also at higher risk of low bone mineral density, high blood homocysteine levels and anaemia, which are directly influenced by diet⁽⁵⁵⁾. It is important to highlight that more than half of the patients with SLE present three or more risk factors for CVD (mostly obesity, hypertension and dyslipidaemias), been certainly more susceptible to suffer with the metabolic syndrome^(56,57). Moreover, obesity leads to increased levels of proinflammatory cytokines, which may exacerbate the inflammatory processes of SLE and increase the risk of diabetes mellitus, atherosclerosis and CHD⁽⁵⁸⁾. Additionally, a hyperlipidic diet, rich in cholesterol and saturated fat, is one of the major risk factors for maintaining dyslipidaemia in patients with SLE, perpetuating and aggravating lipid profile changes⁽⁵⁹⁾. Thus, we can suggest that the nutritional status and food intake of patients with SLE may interfere in the disease course and that an adequate diet may improve SLE prognosis and prevent several related diseases. Currently, a diet rich in vitamin- and mineral-rich foods and MUFA/PUFA with moderate energy consumption is recommended to control the inflammatory findings of the disease and the complications and co-morbidities resulting from SLE therapy^(60,61).

The restriction of energy in the diet has, in particular, shown beneficial effects in murine lupus models. An initial study revealed that a 40% food-restricted maize oil-based diet delayed the onset of autoimmunity in New Zealand black/white (NZB/W) F_1 lupus-prone mice⁽⁶²⁾. Later studies demonstrated that energy restriction prevented the decline in CD8⁺ T lymphocytes, eliminated the abnormal increase in IL-12, IFN- γ , IgA and IgG2 production, delayed the development of kidney disease and age-related immune dysfunction and prolonged the lifespan in aged NZB/W F_1 mice, probably owing to the down-regulation of mRNA expression or NF- κ B^(63,64).

In this regard, there is evidence that dietary factors can contribute to the geoepidemiology of autoimmune diseases⁽¹⁵⁾. Recent studies have suggested that the traditional Mediterranean diet might confer protection from certain chronic diseases related to oxidative stress, inflammation and the immune system. This diet emphasises the intake of vegetables, fruits, nuts, grains and fish, along with small amounts of wine and olive oil as the main monounsaturated fat source, and limits meat consumption. The beneficial effects of the Mediterranean diet have been proven in cancer, CVD, obesity and arthritis⁽⁶⁵⁻⁶⁷⁾. Likewise, epidemiological data have shown a lower prevalence of rheumatic diseases in Mediterranean countries when compared with Northern Europe, and some clinical trials have demonstrated that the Mediterranean diet improves rheumatic symptoms and decreases the use of anti-inflammatory drugs and classic pharmacotherapy-related side effects^(68,69).

The broad range of evidence demonstrating the antioxidant, anti-inflammatory and immunomodulatory effects of some nutrients in immunoinflammatory diseases has suggested a possible supportive role for these nutrients in the primary and secondary prevention of SLE.

However, the question arises as to whether different nutrients could ameliorate or exacerbate SLE symptoms and how they could modulate inflammation and immune function at a molecular level. To this end, the present review summarises preclinical and clinical experiences to provide the reader with an update on the effects of different macro- and micronutrients and dietary phenols in SLE, focusing on the mechanisms of action involved (Tables 1, 2 and 3).

Methods: literature search strategy

A literature survey was conducted to obtain published literature to create this review. The original search was conducted in April 2016 and a second, updated search was completed in October 2016. We searched MEDLINE and SCOPUS (January 1976, to present) databases with the terms 'systemic lupus erythematosus' and 'lupus' in combination with the terms 'aetiology', 'apigenin', 'classification', 'carbinol', 'curcumin', 'diet', 'epidemiology', 'fibre', 'isoflavone', 'flavanols', 'flavones', 'flavonoids', 'flavonol', 'lignans', 'lipids', 'management', 'melatonin', 'mineral elements', 'natural products', 'nutrition', 'olive oil', 'pathogenesis', 'polyphenol', 'protein', 'resveratrol', 'stilbene', 'treatment', 'vitamins', with no language restrictions. We also searched the references of articles identified by this strategy and selected those that were relevant.

Table 1. Beneficial effects of macronutrients in systemic lupus erythematosus (SLE)

Source	Experimental system	Mechanism(s) of action	Effective dose	Reference
Lipids				
<i>n</i> -3 PUFA Fish oil	Patients with SLE	Decreased SLEDAI, leptin and TAG Beduced enthrocyte sedimentation rate and serum II -12 levels	3 g/d Six capsules/d (2.25 g EPA	(77) (76)
		Increased serum IL-13 levels Improved endothelial function	and 2.25 g DHA) 3 g/d	(78)
		Decreased platelet 8-isoprostanes Increased EPA, DHA and HDL-cholesterol	6 g/d, washout period,	(195)
		Reduced platelet arachidonic acid, neutrophil leukotriene B ₄ , TAG and VLDL-cholesterol	18 g/d	(196)
	NZB/W E, mice	Decreased creatinine levels	5% enriched diet	(197)
		Maintained GSH:GSSG ratio		
		Increased SOD, catalase, GPx, glutathione reductase and GST activities Reduced total ROS and COX-derived ROS levels		
		Decreased co-stimulatory (CD80 and CD86) and adhesion (ICAM-1, PGP-1, LFA-1 and Mac-1) molecules in PBMC	5% enriched diet	(198)
		Increased hepatic antioxidant enzymes: liver catalase, SOD and GPx	5% enriched diet	(199)
		Decreased serum cholesterol, TAG and phospholipid and anticardiolipin antibody	180 g/kg enriched diet	(200)
		Decreased fibronectin-1, ICAM-1 and IGF-B1 mRNA levels and IGF-B1 protein in kidney	10% enriched diet	(202)
		Increased antiovidant enzymes (catalase, GPx and SOD) in kidney		
		Decreased serum levels of four COX products	25% enriched diet	(203)
	MRL/lpr mice	Increased IL-10 and IL-2 by spleen cells	9% enriched diet	(204)
		Decreased IL-12, IL-4, PGE ₂ , leukotriene B ₄ and thromboxane B ₂ production by spleen cells		
		Decreased <i>c-myc</i> oncogene in spleen		
		Normalised IGF-β1 levels in spleen cells	00 % envice ad dist	(205)
		Enhanced production of pentaene leukotrienes in supernatant fractions from peritoneal	20% enfiched diet	
		macrophages and kidney		(000)
	BXSB/MpJ and MRL/lpr mice	Decreased proteinuria and renal disease	25% enriched diet	(206)
DHA-enriched fish oil	NZB/W F1 mice	Decreased plasma autoantibodies, proteinuria and glomerulonephritis Down-regulated CD4 ⁺ T-cell-related genes in kidney and/or spleen Reduced expression of CD80, CTLA-4, IL-10, IL-18, CCL-5, CXCR3, IL-6, TNF-α and	60 g/kg enriched diet	(60)
		osteopontin mHNA in kidney and/or spleen	9% opriched diat	(81)
		Suppressed LPS-induced IL-18 in Suppressed LPS-induced IL-18 in		
Concen-trated fish oil (Lovaza®)	NZB/W F1 mice	Decreased anti-dsDNA antibolities, IL-1 β , IL-6 and TNF- α in splenocytes and kidney, serum TAG and liver adinosity	4% enriched diet	(207)
		Increased renal antioxidant enzymes GPx and catalase activity		
<i>n</i> -6 PUFA		Down-regulated full-kD activation		
Evening primrose oil	MRL/lpr mice	Decreased serum anti-dsDNA antibodies, proteinuria and clinical features	5% enriched diet	(208)
Maize öil, soyabean oil, ruminant meat	MRL/lpr mice	Reduced oxidative stress	30 mg/d by oral	(88)
		Enhanced detoxifying enzyme activity (GST, NQO1 and γGCS). Enhanced nuclear translocation of Nrf-2 in liver	administration	(07)
		25 μм: enhanced GCLC expression and intracellular GSH concentration in splenocytes 100 μм: down-regulated IFN-γ synthesis and cell proliferation and decreased apoptosis induction and intracellular thiols (GSH, GSSG) in splenocytes	25 and 100 μм	(87)
		Decreased anti-dsDNA and anti-tTG IgG, IL-4, IL-10 and IFN-y mRNA and down-regulated	30 mg/d by oral	
	NZB/W F1 mice	NF-κB activity and apoptosis induction in spleen cells Decreased SLE symptoms	administration 0.5 % enriched diet	(89)
MUFA		וויטוסמססע סעו אועמו		
Extra virgin olive oil	Pristane BALB/c mice	Decreased serum MMP-3 and PGE ₂ in kidney and proinflammatory cytokine production in spleen cells	100 g/kg diet	(103)

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Table 1. Continued

Source	Experimental system	Mechanism(s) of action	Effective dose	Reference
Proteins Royal jelly	NZB/W F1	Reduced IL-10, anti-ssDNA and anti-erythrocyte surface antigen level Decreased number of splenic autoreactive B cells Increased survival	2 mg by oral administration	(106)
Arinno acids: taurine Eggs, meat, seafood	NZB/W F1	Reduced cardiac abnormalities Increased fibrotic signalling molecules and IGF1R survival signalling components Reduced caspase-3 activity, TUNEL-positive cells and Fas- and mitochondrial-dependent anonosis	1% enriched diet 1% enriched diet	(107) (108)
i		Increased Att, ERK1/2 and p65 NF-kB phosphorylation Decreased p38 phosphorylation		
Fibre Beans, cereals, whole grains, vegetables	Patients with SLE	Decreased risk of active disease by decreasing levels of some cytokines and homocysteine	>4.7 g soluble fibre or >14 g insoluble fibre/d	(111)
Akt, protein kinase B; CCL-5, chemokine signal-regulated protein kinase; GCL 5-transferase; HO-1, haem oxygen: LPS, lipopolysaccharide; Mac-1, macr factor-2; NZB/M, New Zealand black' SOD, superoxide dismutase; SSDNA,	(C-C motif) ligand 5; COX, cyclo-oo C, ryellutamylcysteine ligase cataly sise-1; LOAM-1, intercellular ache phage-1 antigen, intercellular ache phage-1 antigen, intercellular single-stranded DNA; STAT, signal	vgenase; CTLA-4, cytotoxic T-lymphocyte-associated protein-4; CXCR3, C-X-C motif chemokine receptor-3; dsl tic subunit; VGCS, y glutamyl cysteine synthetase; GPx, glutathione peroxidase; GSH, reduced glutathione; C sion molecule-1; IGF1R, insulin-like growth factor receptor-1; IFN, interferorn; JAK, Janus kinase; LFA-1 -activated protein kinase; MMP-3, metalloproteinase-3; MRL, Murphy Roths ange; MOD(1, ND(P)H; quinone ox nonuclear cell; PGP-1, permeabilty glycoproteinase; PI3K, phosphoinositide 3-kinase; ROS, reactive oxygen transducer and activator of transcription; TGF, transforming growth factor; ITG, IVE	DNA, double-stranded DNA; ERK 3SSG, oxidised glutathione; GST 1, lymphocyte function-associate idoreductase-1; Nrf2, nuclear fact species; SLEDAI, SLE disease f L, terminal deoxynucleotidyl trans	, extracellular ; glutathione- ad antigen-1; or E2-related activity index; sferase dUTP

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Macronutrients in systemic lupus erythematosus

Lipids

Among traditional atherosclerotic risk factors in SLE, dyslipidaemia is believed to decisively affect the long-term prognosis of patients with SLE, not only with regard to cardiovascular events but also by influencing other manifestations, such as lupus nephritis and brain damage among others⁽⁷⁰⁾. Therefore, dyslipidaemia in SLE should be prompt and adequately treated in order to reduce overall morbidity and mortality and improve patients' quality of life. Moreover, dyslipaemia is considered as a modifiable risk factor that can be managed with an accurate treatment, which includes the promotion of a healthy and varied diet^(70–72).

Specifically, there is both preclinical and clinical evidence that patients with SLE may benefit from the consumption of certain lipids. In particular, the n-3 PUFA possess the most potent immunomodulatory activity; among them, EPA and DHA are the most biologically active. These n-3 PUFA suppress proinflammatory cytokine production, lymphocyte proliferation, cytotoxic T-cell activity, macrophage-mediated cytotoxicity and neutrophil/monocyte chemotaxis⁽⁷³⁾. Some of the effects of n-3 PUFA are produced by the modulation of the amount and types of eicosanoids, but other effects have eicosanoid-independent mechanisms, including modulation of intracellular signalling pathways, transcription factor activity and gene expression⁽⁷⁴⁾. As humans and other mammals require PUFA but cannot synthesise them, the consumption of PUFA through the daily diet is essential. The available data show that an increased daily intake of dietary n-3 PUFA decreased the severity of autoimmune disorders. A low intake of n-3 PUFA and high intake of carbohydrate in SLE appear to be associated with worse disease activity, adverse serum lipids and plaque presence⁽⁷⁵⁾. Similarly, consumption of fish oil, which is rich in n-3 PUFA, has been shown to be useful in preventing and/or ameliorating SLE symptomatology. In fact, different clinical studies have assessed the association between dietary supplements and fish oil in patients with SLE, revealing a number of positive effects, including reducing cardiovascular, inflammatory and neuromotor symptoms and improving depressive symptoms and fatigue^(76–79). In addition, n-3 PUFA from fish oil have demonstrated anti-inflammatory effects when used as supplements in murine SLE models, decreasing autoantibody production and inflammatory gene expression in the kidney and spleen, suppressing glomerulonephritis and extending lifespan^(80,81). However, a major question in the use of PUFA is whether the doses required to suppress autoimmunity and inflammation could have a negative impact on innate and adaptive immune responses against pathogens or neoplastic events⁽⁸²⁾.

Conjugated linoleic acid (CLA) is the term used to describe positional and geometric isomers of the *n*-6 PUFA linoleic acid (18 : 2*n*-6). The main sources of CLA in the human diet are ruminant meat and dairy products, which are extensively used in Western diets, but it can be also found in small amounts in oils derived from plants^(83,84). CLA has shown potential antiinflammatory and cancer-preventive properties, most probably through its ability to modify eicosanoid signalling by altering cell membrane composition and modulate genes through PPAR^(85,86). CLA has shown beneficial effects in SLE mouse



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Table 2. Beneficial effects of micronutrients in systemic lupus erythematosus (SLE)

Source	Experimental system	Mechanism(s) of action	Effective dose	Reference
Vitamins				
Carrot, sweet potato, fish, spinach, liver, tropical fruits	Patients with SLE	Decreased Th17 cells Increased Treg percentages in CD4 ⁺ T cells cultures from hypovitaminosis A natients	0·3 µg/ml retinoic acid	(118)
		Decreased proteinuria	10 mg/d ATRA	(116)
	NZB/W F ₁ mice	Reduced IL-2, IFN-γ and IL-12 and IL-4 in serum Improved survival and proteinuria Suppressed iNOS and MCP-1 in kidney Beduced IgG2-specific anti-DNA antibody	0-1 mg ATRA i.p.	(117)
		Improved survival, splenomegaly, proteinuria and renal pathological findings and serum anti-DNA levels	0.5 mg ATRA i.p.	(116)
	MRL/lpr mice	Reduced IFIN-Y, IL-2 and IL-10 in spienic CD4 ⁻¹ cells Reduced dermal thickness Suppressed appearance of skin lesions by inducing apoptosis and regulation of cvtokine expression	5–10 mg/kg etretinate by oral administration	(120)
Vitamin B ₂		Reduced lymphoproliferation and glomerulonephritis	10 mg/kg per d	(119)
Fish, beef, liver, potatoes, fruits, cereals	Patients with SLE	Suppressed active inflammation and prevented the occurrence of SLE flares	>1.7 mg/d	(111)
Vitamin C		D	//	(143)
Fruits, broccoli, tomatoes	Patients with SLE	Decreased lipid peroxidation	500 mg/d	(60)
Vitamia D		Decreased risk of active disease	109-99 mg/d	()
Animal-based foods, fish oil, mushrooms	Patients with SLE	Decreased IL-17A-producing T cells increasing FOXP3 expression	2000, 4000 or 50000 IU/week for 6 months	(131)
		Reduced IL-6 secretion Augmented angiogenic capacity and paracrine regulation of endothelial NOS	400000 and 20000 IU/week	(135)
		Decreased disease activity	50000 IU/week	(136)
		Regulatory effects on apoptosis and cell cycle progression in PBMC Up-regulated expression of <i>Bcl-2</i>	50 пм	(134)
		Down-regulated expression of <i>Bax</i> and FasL Decreased endothelial cell early apoptosis through inhibition of externalised	10 пм	(209)
		Inhibited DC maturation, decreased CD40, CD86 and HLA-DR expressions and cytokine secretion of IL-12p70	10 пм	(128)
		Decreased percentage of Th17 cells and IL-17A levels in CD4 ⁺ T cells Restored immune homeostasis through inhibitory effects on DC maturation	10 пм	(210)
	BALB/c mice with activated chromatin	and activation. Reduced IL-6 and IL-10 mRNA expression Increased TGF-6 and FOXP3 mRNA expression	50 ng/d by oral administration	(133)
		Enhanced Treg percentage in spleen		
	MRL/lpr mice	Reduction of proteinuria and serum anti-ssDNA	0·1 μg–0·15 μg/d i.p	(132)
Vitamin E Sunflower seeds, almonds, groundnuts, cereals,	Patients with SLE	Suppressed anti-dsDNA autoantibody production via antioxidant activity	150–300 mg/d	(141)
spinacii, aspaiayus, avucauu		Decreased linid peroxidation	800 IU/d	(143)
	NZB/W F1 mice	Increased IL-2 in splenocytes	250 mg/kg enriched diet	(140)

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Source	Experimental system	Mechanism(s) of action	Effective dose	Reference
	MRL/lpr mice	Decreased oxidative stress and anti-dsDNA antibodies Regulated cytokines and lymphocyte subsets Improved survival and proteinuria Normalised mitogenic responses of B and T cells Decreased anti-dsDNA antibodies and amyloid P component in serum	275 mg/kg enriched diet 0.4 mg/d five times per week	(139) (211)
Mineral elements Se Plants, grains	NZB/W F1 mice	Improved survival Increased natural killer cell activity	4 ppm in water	(144)
ATRA, all- <i>trans</i> -retinoic acid; Bax, BCL2 associated X protei related; IFN, interferon; INOS, inducible NO symhase; i.p. mononuclear cell; ppm, parts per million; sSDNA, single-s	in; Bcl-2, B-cell lymphoma 2; DC ., intraperitoneally; MCP-1, monc stranded DNA; TGF, transformin	c) dendritic cell; dsDNA, double-stranded DNA; FasL, Fas ligand; FOXP3, forkhead box poyle chemoattractant protein-1; MRL, Murphy Roths large; NOS, NO synthase; NZBM, g growth factor; Th, T-helper; Treg, regulatory T cell.	-3; HLA-DR, human leucocyte antiger New Zealand black/white; PBMC, per	– antigen D ipheral blood

models, reducing splenomegaly, autoantibody and cytokine production, NF-κB activity (independent of PPAR-γ activation) and oxidative stress, thus improving SLE symptoms and survival rates^(87–89). Moreover, some animal studies have shown that CLA consumption has anti-sclerotic and anti-oxidative effects and improves the blood lipid profile, which could be also beneficial for SLE^(90,91). On the other hand, a previous study suggested that dietary CLA may accelerate the appearance of autoimmune symptoms in NZB/W F1 mice, despite protecting against disease-related body-weight loss and prolonged survival⁽⁹²⁾. By the way, only a few studies examined the effects of CLA in human subjects in vivo and their results do not show the favourable results observed in animal studies⁽⁹³⁻⁹⁵⁾. In this sense, the majority of the clinical studies did not provide conclusive evidence for the effectiveness of CLA on human health, except for anti-obesity properties^(96,97). Although the interest in CLA still remains, there are multiple questions related to the safety and efficacy on its consumption that must be scientifically solved⁽⁹⁸⁾.

Controversially, an essential fatty acid-deficient diet has shown beneficial effects in NZB/W F₁ mice by reducing levels of arachidonic acid (20 : 4n-6), a precursor of proinflammatory eicosanoids, PG and leukotriene metabolites, decreasing autoantibody production and improving nephritis⁽⁹⁹⁾. Furthermore, an increased intake of *n*-6 PUFA may increase the production of proinflammatory cytokines and the incidence of autoimmune diseases by increasing free radicals and decreasing antioxidant enzyme mRNA levels⁽¹⁰⁰⁾.

Recent experimental and clinical studies have confirmed that regular extra virgin olive oil (EVOO) consumption can exert positive effects in rheumatic diseases^(101,102). The beneficial properties of EVOO are linked to its high MUFA content and also to its multiple minor components, particularly polyphenol compounds such as flavonoids, lignans and secoiridoids and their hydrolysis products (hydroxytyrosol and tyrosol, among others). Recently, we evaluated the effects of an EVOO diet in a pristane-induced SLE model in mice, and we demonstrated that the EVOO diet significantly reduced renal damage and decreased metalloproteinase (MMP)-3 serum and PGE2 kidney levels as well as proinflammatory cytokine production in splenocytes. In addition, our data indicated that Nrf-2 and haem oxygenase protein expression was up-regulated by the EVOO diet and that the activation of the Janus kinase and signal transducer and activator of transcription (JAK/STAT), mitogenactivated protein kinases (MAPK) and NF-kB pathways was markedly ameliorated, supporting the interest in EVOO as a beneficial functional food exerting a preventive/palliative role in the management of SLE⁽¹⁰³⁾.

Proteins

The restriction of dietary protein has been shown to have beneficial effects in controlling renal disease progression in animal models and human studies. Specifically, a protein-restricted diet (0-6 g/kg per d) improved nutritional status and glomerular filtration rate in patients with SLE with chronic kidney disease⁽¹⁰⁴⁾. In addition, excessive protein intake has been shown to produce bone mineral loss in patients with juvenile SLE⁽¹⁰⁵⁾. By contrast,





Table 3. Beneficial effects of phenolic compounds in systemic lupus erythematosus (SLE)

Phenol	Source	Experimental system	Mechanism(s) of action	Effective dose	Reference
Polyphenols Flavonoids Flavones					
Apigenin	Parsley, thyme, peppermint, chamomile olives	SNF1 mice	Suppressed IFN-γ response and anti-dsDNA, anti-ssDNA, anti-nucleosome and anti-histone autoantibody production in splenocytes Decreased IL-6 production by DC Suppressed IEN y and IL 17 production in antigan procenting cells	0-3–100 µм	(150)
U H			Induced apoptosis of T cells, B cells, DC and macrophages Reduced IFN-γ and IL-17 in splenocytes Decreased anti-dsDNA, anti-ssDNA, anti-nucleosome and anti-histone autoantibodies in serum and spleen cells Improved lupus nephritis Reduced COX-2 expression in antigen-presenting cells, CD4 ⁺ T cells, B cells, DC	20 mg/kg per d i.p.	
Flavonols			and macrophages		
Astilbin	Grapes, Smilax glabra	MRL/lpr mice	Reduced serum antinuclear antibodies and cytokines (IFN-γ, IL-17A, IL-1β, TNF-α and IL-6) Decreased functional activated T and B cells from splenocytes	10, 20 or 40 mg/kg enriched diet	(153)
			Improved renal damage		
Flavanols	Croop too		Destroaded DOS lovels in communities and kidney	100 mg//g opriched dist	(159)
EGCG	Green tea	NZB/W F1 mice	Prevented proteinuria and maintained renal function Increased Nrf-2 and NF-κB pathways and GPx in kidney Inhibited renal NLRP3 inflammasome expression Decreased mature caspase-1, IL-1β and IL-18 in kidney and IL-1β and IL-18 levels in serum	120 mg/kg enriched diet	()
ОН		MPL /lpr mico	Enhanced splenic regulatory Treg activity	50.00	(160)
lasflovonos			Blocked induced expression of iNOS and production of NO and IL-6 Attenuated inflammation via PI3K/Akt/mTOR pathway by decreasing Akt phosphorylation	50 µm	
			in mesangial cells Decreased lymph node hyperplasia, fatty accumulation, proteinuria and blood urea N Increased survival Decreased serum levels of anti-DNA antibodies and kidney immune complexes	2 % green tea-enriched diet	(158)
		Human primary epidermal keratin- ocytes	Improved glomerulonephritis and nephric vasculitis Down-regulated the expression of antinuclear autoantibodies SS-B/la and SS-A/Ro, coilin, DNA topoisomerase I and α-fodrin	100 µм	(161)
Daidzein	Soya beans	MRL/lpr mice	Improved body weight and survival Decreased anti-dsDNA IgG and anticardiolipin IgG serum levels Decreased <i>in vitro</i> IFN-γ production on mitogen-stimulated T cells from spleen	20 mg soya isoflavones/kg BW/d by oral administration	(163)
Coursetrol	Lucerne, sova beans	MBL /Inr mice	Delayed the onset of proteinuria	25 ma lucerne sprout ethyl	(170)
но сторон	Brussels sprouts, spinach, legumes		Improved renal function and survival Suppressed TNF- α and IL-1 β from LPS-stimulated peritoneal exudate cells and IFN- γ and	acetate extract/kg BW by oral administration	
		BALB/c mice	Reduced serum levels of TNF- α , IL-6, and IL-1 β and improved survival after 9h of LPS challenge	25 mg lucerne sprout ethyl acetate extract/kg BW by oral administration	(169)
		NZB/W F₁ mice	Inhibited IL-6 and TNF-α production in LPS-primary macrophages Decreased splenomegaly and proteinuria	50 µg/ml 0.01 % enriched diet	(168)

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Nutrition and systemic lupus erythematosus

Table 3. Continued

Phenol	Source	Experimental system	Mechanism(s) of action	Effective dose	Reference
Lignans	-1 - 1				(177)
SDG	Flax, pumpkin, sunflower, sesame seeds	Patients with SLE	Decreased serum creatinine and microalbumin Improved renal function, decreasing serum creatinine, plasma lipids, blood viscosity and increasing complement C3 I	30 g ground flaxseed/d 30 g ground flaxseed/d	(176)
HO- OH OH		MRL/lpr	Decreased proteinuria and improved renal function	600, 1200 and 4800 μg by oral administration	(175)
HO, OH OH			Improved survival Delayed and decreased proteinuria and splenic lymphocyte proliferation Attenuated glomerular filtration and lymphoproliferation	15 % flaxseed-enriched diet	(174)
Diarylheptanoids Curcumin	and arylalkanones Curcuma longa L.	Patients with SLE	Modulated Th17/Treg balance of CD4 ⁺ T cells by reducing Th17 response and IL-17A	0.1 and 1 µg/ml	(178)
но Сталина стали		MRL/lpr	Decreased proteinuria levels and systolic blood pressure Delayed onset of anti-RNP, anti-Sm, ANA and anti-dsDNA autoantibodies and proteinuria Decreased solicy gland infiltration and lymphedopopathy	22.1 mg three times/d 5 mg/ml in water	(181) (179)
		NZB/W F1 mice	Decreased salivary gland initiation and tymphadehopathy Decreased proteinuria, serum levels of IgG1, IgG2a and anti-dsDNA and IgG glomeruli immune complex deposition Reduced TNF-α and MCP-1 in kidney and FOXP3 in spleen	1 % enriched diet	(180)
Resveratrol	Grapes	Pristane BALB/c mice	Decreased proteinuria, IgG and IgM Ig deposition in kidney and glomerulonephritis and serum IgG1 and IgG2a Suppressed CD69 and CD71 expression on CD4 ⁺ T cells, CD4 ⁺ T cell proliferation, induced CD4 ⁺ T cell apoptosis, decreased CD4 IFN-γ+ Th1 cells and Th1:Th2 ratio in splenic mononuclear cells Inhibited IgG1, IgG2a, IgG2b, IgG3, IgM and IgA antibody production and proliferation of stimulated B lymphocytes from splenic mononuclear cells	50–75 mg/kg enriched diet 0, 10, 20, 40 or 80 µм	(184)
	Cruciferous vegetables	NZB/W F ₁	Increased lifespan	0.2 g/kg enriched diet	(185)
			Decreased proteinuria, glomerulonephritis and interstitial nephritis Decreased proteinuria and renal abnormalities and prolonged survival Decreased anti-dsDNA, anti-chromatin and anti-RNA helicase A antibody levels Increased 2-OH:16a-OH ratio metabolites Blocked B, and Tcoll maturation	2000 ppm enriched diet	(186)
Malatania		Patients with SLE	Changed urinary ratio of 16α -OH:2-OH by an increase of 2-hydroxylation	375 mg/d	(187)
	Fruits, vegetables, olive oil and nuts	Patients with SLE Pristane BALB/c mice	Increased number of Tregs expressing FOXP3 and BAFF mRNA expression Antagonised increasing levels of IgM, anti-ssDNA and histone autoantibodies Decreased renal lesions Decreased induced IL-6 and IL-13 production Increased II -2 levels in stimulated splenocytes	1 × 10 ⁻⁴ м 0·01, 0·1, 1·0 mg/kg intragastric	(194) (193)
		MRL/lpr mice	Decreased serum IgG, IgM, anti-dsDNA and anti-CII autoantibodies, cytokines (IL-2, IL-6, IFN- γ , TNF- α , and IL-1 β) and nitrite/nitrate production in females Increased IL-10 in females	30 mg melatonin/kg BW in water	(191)

2-OH, 2-hydroxyestrone; 16α-OH, 16α-hydroxyestrone; Akt, protein kinase B; AMPK; adenosine 5'-monophosphate-activated protein kinase; ANA, antinuclear antibodies; BAFF, B-cell activating factor; BW, body weight; COX, cyclooxygenase; DC, dendritic cell; dsDNA, double-stranded DNA; EGCG, epigallocatechin gallate; FOXP3, forkhead box P3; GPx, glutathione peroxidase; IFN, interferon; iNOS, inducible NO synthase; i.p., intraperitoneally; LPS, lipopolysaccharide; MCP-1, monocyte chemoattractant protein-1; MRL, Murphy Roths large; mTOR, mammalian target of rapamycin; NLRP3, NLR family pyrin domain containing 3; Nrf-2, nuclear factor E2-related factor-2; NZB/W, New Zealand black/white; PI3K, phosphoinositide 3-kinase; ppm, parts per million; RNP, ribonucleoprotein; ROS, reactive oxygen species; SDG, secoisolariciresinol diglucoside; SNF, sucrose non-fermenting; ssDNA, single-stranded DNA; TGF, transforming growth factor; Th, T-helper; Treg, regulatory T cell.

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oral supplementation with royal jelly, a honeybee secretion rich in proteins, free amino acids, SCFA and vitamins, has also been considered beneficial because its reduces cholesterol and has immunomodulating and anti-inflammatory activities. In fact, the intake of 2 mg of protein from royal jelly induced a reduction of IL-10, anti-single-stranded DNA and anti-erythrocyte surface antigen autoantibody serum levels, decreased the number of splenic autoreactive B cells and increased survival in NZB/W F₁ mice, suggesting a beneficial effect in preventing the early onset of SLE and in controlling the active progression of SLE manifestations⁽¹⁰⁶⁾.

Taurine, the major intracellular free β-amino acid in most mammalian tissues, is obtained largely from the diet through eggs, meat and seafood, and it plays crucial roles in protecting biological systems owing to its antioxidant, anti-inflammatory and antiapoptotic properties. The potential benefits of taurine intake in SLE have been demonstrated in an in vivo model of NZB/W F1 mice fed with a cholesterol-rich diet supplemented with taurine (1%); taurine supplementation ameliorated cardiac abnormalities with reductions in aggravated histopathological changes, Fas- and mitochondrial-dependent apoptosis and fibrotic signalling molecules and an increase in insulin-like growth factor receptor 1 survival signalling components⁽¹⁰⁷⁾. In another study with NZB/W F₁ mice that were fed a cholesterol-rich diet supplemented with taurine (1%), taurine significantly reduced caspase-3 activity, terminal deoxynucleotidyl transferase dUTP nick end labelling (TUNEL)-positive cells and Fas- and mitochondrial-dependent apoptosis, increased phosphorylation of protein kinase B (Akt), extracellular signal-regulated protein kinases 1 and 2 and p65 NF-kB proteins and decreased phosphorylation of p38 protein in the liver. These results strongly suggest the therapeutic potential of taurine intake in SLE management⁽¹⁰⁸⁾. By contrast, an increased intake of the amino acid 1-arginine increased the severity of renal fibrosis and the likelihood of death in Murphy Roths large (MRL)/lpr mice by enhancing cytotoxic NO generation via inducible NO synthase (iNOS)⁽¹⁰⁹⁾. Moreover, a diet with restricted phenylalanine and tyrosine had beneficial effects on NZB/W F1 mice by decreasing autoantibody production⁽¹¹⁰⁾.

Fibre

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An adequate intake of dietary fibre is recommended in SLE because of the beneficial effects of fibre in decreasing cardiovascular risk, promoting gut mobility and reducing serum levels of inflammation markers such as C-reactive protein, cytokines and homocysteine. A study with 279 women with SLE showed an inverse association between dietary fibre intake and SLE; increased dietary fibre decreased the risk of active disease in patients with SLE by decreasing the levels of some cytokines and homocysteine⁽¹¹¹⁾. However, excessive fibre intake may reduce the absorption of vitamins, minerals and proteins⁽⁵⁹⁾.

Micronutrients

Vitamins

Several epidemiological studies have explored the potential role of antioxidant nutrient intake and supplementation in patients with SLE. In SLE, oxidative stress acts as an autoimmunity trigger that contributes to immune system dysregulation, abnormal apoptotic events and autoantibody production. Levels of oxidative stress have been shown to be correlated with SLE disease activity and organ damage in patients and in SLE-prone mouse models^(112,113). In addition to spreading inflammation through the bloodstream, mediators of oxidative stress contribute to organ damage, including the cardiovascular system, kidneys and skin. Thus, suppressing pathways of oxidative stress may reduce the toxicity of immunosuppressive therapies, decrease disease activity and improve the quality of life of patients with SLE⁽¹¹⁴⁾.

Vitamin A is essential for multiple functions, including the maintenance of immune system integrity, and its physiological form, all-trans-retinoic acid (ATRA), regulates gene transcription by binding to nuclear retinoic acid receptors. ATRA has demonstrated paradoxical effects on the development of autoimmune lupus in the MRL/lpr mouse, decreasing inflammation in some organs while generating more severe disease in others⁽¹¹⁵⁾. However, ATRA has also shown beneficial effects, alone or in combination with low-dose immunosuppressive drugs, in lupus nephritis and cytokine modulation in both mouse models and patients with SLE, suggesting that ATRA treatment significantly alleviates hyper-reactive autoimmune and renal disorder, most likely via cytokine regulation^(116–119). Furthermore, oral treatment with etretinate, a synthetic vitamin A derivative, every 2 d for 2 months in lupus mice reduced dermal thickness through the inhibitory effects on collagen synthesis and inhibited skin lesions by inducing apoptosis in dermal infiltrating cells and, perhaps, regulating cytokine production⁽¹²⁰⁾.

Higher serum levels of homocysteine are associated with atherosclerotic vascular events in patients with SLE. In this sense, an increase of one log unit of homocysteine concentrations led to a 2·4-fold increase in the risk of stroke, and a 3·5-fold increase in the risk of arterial thrombosis in $SLE^{(121)}$. In this regard, dietary intake of vitamin B₆, which has been shown to decrease homocysteine levels and acts as a coenzyme in the metabolism of antibodies and cytokines, decreased the risk of active SLE in female patients, suggesting that its daily intake may lead to the suppression of active inflammation and prevent the occurrence of SLE flares. However, in the present study there was no significant association between B₆ intake and a decreased risk of atherosclerotic vascular events⁽¹¹¹⁾.

Vitamin C may mediate the oxidative stress response in SLE and, consequently, exert beneficial effects on the repair of abnormal immune components and inflammation. Vitamin C intake (500 mg/d for 6 weeks) may positively modulate oxidative DNA damage in healthy subjects, but these positive effects were not observed in patients with SLE. These significant differences in the response to vitamin C supplementation between patients with SLE and healthy subjects might be associated with DNA repair abnormalities observed in SLE⁽¹²²⁾. Another study reported that vitamin C intake (109-99 mg/d) is inversely associated with the risk of active SLE, decreasing oxidative stress and suppressing autoantibody production, suggesting that its intake may prevent the occurrence of SLE flares⁽⁶⁰⁾.

Vitamin D is found in small quantities in eggs, fatty fish, and supplemented dairy products; however, the main source of vitamin D is sunlight exposure⁽¹²³⁾.Vitamin D has a critical role in Ca homeostasis, mineralisation of bone tissue, muscle

function and coordination. Indeed, low levels of vitamin D result in a decrease of the Ca reserves in bone in an attempt to correct for the reduced Ca that will be absorbed from the gut⁽¹²⁴⁾. Nowadays it is clear that vitamin D deficiency contributes to the morbidity and mortality of multiple chronic diseases, including SLE⁽¹²⁵⁾. Particularly, vitamin D deficiency is even more prevalent in patients with SLE than in the general population, in part because patients with SLE are recommended to avoid the sun, in order to prevent disease flares⁽¹²⁶⁾. In this sense, some studies have shown that vitamin D deficiency increases predisposition to suffer SLE partly owing to the photosensitive nature of the disease, exacerbates symptoms, decreased bone health and it is correlated with the disease activity⁽¹²⁷⁻¹²⁹⁾. Moreover, in a study with adolescent women with SLE supplemented with Ca and vitamin D (400-800 IU daily), a lack of vitamin D was associated with a decrease of bone mineral density, highlighting the importance of well-defined vitamin D supplementation protocols in $SLE^{(130)}$. Vitamin D supplementation (2000, 4000 or 50000 IU weekly for 6 months) in patients with SLE showed positive immunological effects by decreasing IL-17A-producing T cells and increasing FOXP3 expression, confirming the relationship between vitamin D status and immunological balance⁽¹³¹⁾. Furthermore, vitamin D consumption attenuated the disease progression and increased the number of regulatory T cells in mouse models^(132,133). In patients with SLE, in vitro treatment with vitamin D (50 nm) has regulatory effects on apoptosis and cell cycle progression and modifies the expression levels of apoptotic genes⁽¹³⁴⁾; in addition, vitamin D intake (400 000 IU followed by 20 000 IU weekly for 12 weeks) can positively modulate endothelial function in patients with stable SLE, reducing cardiovascular risk⁽¹³⁵⁾. Moreover, vitamin D supplementation (50 000 IU per week for 24 weeks) has been shown to be effective in decreasing disease activity and improving fatigue in juvenile-onset SLE⁽¹³⁶⁾. Therefore, vitamin D supplementation is currently recommended as a treatment for some patients with SLE, although determining the effectiveness and safety of vitamin D use as a treatment in SLE, as well as establishing the exact doses, still requires additional extensive interventional studies⁽¹³⁷⁾. Nevertheless, general international recommendations have established that vitamin D supplementation with 800 to 1000 IU/d or 50000 IU monthly is safe for most individuals and can ensure levels of vitamin D within the optimal range. This intake is within the currently recommended safe upper tolerable limit for vitamin D of 2000 IU/d for those aged 1 year and older⁽¹³⁸⁾.

Vitamin E is a potent free radical and peroxyl radical scavenger and an essential nutrient for maintaining the normal function of the immune system. A diet supplemented with vitamin E decreased oxidative stress and anti-dsDNA antibody production, regulated cytokines and lymphocyte subsets and alleviated the severity of SLE under oxidative stress in NZB/W F₁ mice⁽¹³⁹⁾. However, while low vitamin E supplementation increased the survival of MRL/lpr mice, high supplementation had the opposite effect and inhibited the Th1 pathway, which may not be beneficial for Th2-prone autoimmune diseases, such as SLE. This duality might be due to selective modulation through PGE₂ production or through PPAR γ , inhibitor of NF- κ B (I κ B- α) and apoptotic pathways under different doses of vitamin $E^{(139,140)}$. In patients with SLE, the oral administration of vitamin E (150–300 mg/d) together with prednisolone decreased autoantibody production but not the urinary levels of 8-hydroxydeoxyguanosine, an indicator of oxidative DNA damage, suggesting that vitamin E can suppress autoantibody production via a mechanism independent of antioxidant activity⁽¹⁴¹⁾.

Despite the aforementioned evidence, a survey study showed that the dietary intake of antioxidant nutrients, including vitamins A, C and E, α -carotene, β -carotene, cryptoxanthin, lycopene and lutein, from foods and supplements does not change the risk of developing SLE in women, suggesting that antioxidants do not offer protection against SLE⁽¹⁴²⁾. In addition, a study investigating the effects of vitamin E and C supplementation (500 mg vitamin C and 800 IU vitamin E daily) on endothelial function and markers of oxidative stress and antioxidant defence in patients with SLE showed that their combined administration decreased lipid peroxidation but did not affect endothelial function after 3 months of therapy⁽¹⁴³⁾. Insufficient dosing and/or the inability of these vitamins to regulate intracellular signalling pathways within the immune system may explain this lack of therapeutic efficacy.

Mineral elements

Se, a trace mineral widely distributed in plants and grains, is a required nutrient for animals and humans that has been shown to have immunological and anti-inflammatory properties. Se supplementation (sodium selenite 4 parts per million (ppm) in the drinking water) demonstrated beneficial effects in female NZB/W F_1 mice, improving survival and increasing natural killer (NK) cell activity. However, Se supplementation had no effect on autoantibody production⁽¹⁴⁴⁾.

Nowadays, it is recommended that patients with SLE follow a Na-restricted diet because evidence suggests that excess sodium chloride content in the diet might be a potential risk factor for autoimmune diseases. A study in MRL/lpr mice demonstrated that excessive dietary sodium chloride intake aggravated lupus nephritis through the effect of the inducible serine/threonine protein kinase 1 pathway on the Th1/Th2 and Th17/Treg balance *in vivo*⁽¹⁴⁵⁾.

Dietary restriction of Zn also has shown beneficial effects in SLE. Moderate (5-0 ppm daily) to severe (2-5 ppm daily) dietary restriction of Zn and decreased energy intake in NZB/W F_1 mice decreased the onset of haemolytic anaemia, autoantibody production and renal disease⁽¹¹⁰⁾. In MRL/lpr mice, a Zn-restricted diet has been shown to reduce lymphoproliferation and anti-dsDNA titres and improve glomerulonephritis⁽¹⁴⁶⁾.

Dietary phenols in systemic lupus erythematosus

Polyphenols

Polyphenols are common constituents of plant-based foods and the main dietary source of antioxidants through vegetables, fruits, cereals, legumes and drinks such as tea, coffee and wine. The term polyphenol comprises a wide variety of molecules with phenolic structural features that are classified into several groups according to their number of phenol rings and the structural elements that bind these rings together. The main Nutrition Research Reviews

categories of polyphenols are phenolic acids, flavonoids, diarylheptanoids and arylalkanones lignans and stilbenes. There is marked variation in bioavailability among polyphenols, mainly owing to differences in chemical structure. Furthermore, the most abundant polyphenols in our diet are not necessarily those that present the best bioavailability profile⁽¹⁴⁷⁾.

In recent years, the interest in dietary polyphenols has increased considerably among nutritionists and food scientists owing to their beneficial roles in human health; polyphenols act as anti-inflammatory, anti-cancerous and immunomodulatory agents in several diseases. The potential beneficial effects of polyphenols in SLE are derived from their abilities to protect against oxidative damage, modulate different inflammationrelated enzymes and interact with signal-transduction pathways, cell cycle regulators and cell receptors implicated in the immunoinflammatory process^(65,148). Evidence from both *in vitro* and *in vivo* studies demonstrates the beneficial effect of phenolic compounds in the management of SLE (Table 3).

Flavonoids: flavones. Apigenin, a flavonoid widely distributed in dietary plants such as parsley, thyme and chamomile, among others, has vasorelaxing, antiplatelet and antioxidant properties, which may reduce the risk of coronary disease and improve endothelial function in SLE^(149,150). Apigenin has shown beneficial effects in an SLE mouse model, suppressing autoantibody production and the IFN-y and IL-17 response of stimulated splenocytes, IL-6 production by lupus dendritic cells and inducible isoform of cyclo-oxygenase-2 (COX-2) expression in CD4⁺ T cells, B cells, dendritic cells and macrophages. Both IFN-y-producing Th1 cells and IL-17-producing Th17 cells are critical for help in the production of pathogenic autoantibodies and the development of lupus nephritis^(151,152). In addition, apigenin treatment *in vitro* induced significant apoptosis of T cells, B cells, dendritic cells and macrophages after 24 h of incubation⁽¹⁵⁰⁾. Nevertheless, the presence of apigenin in the diet is insufficient to reach bioavailable therapeutic levels owing to first-pass metabolism in the gut and liver, but its bioavailability could potentially be improved by the pharmaceutical industry.

Flavonoids: flavonols. Astilbin, a natural flavonol found in some food and medicinal plants, exhibits multiple pharmacological functions, including anti-inflammatory and immunomodulatory properties through the induction of apoptosis in activated T cells and the suppression of activated T-cell adhesion and migration and modulation of dendritic cells. Oral administration of astilbin, isolated from the rhizome of *Smilax glabra* Roxb (family Liliaceae), delayed disease development in lupus-prone mice when preventive oral administration was started before the onset of disease and also when the treatment was started after disease onset. In addition, astilbin treatment reduced levels of circulating antinuclear antibodies and several serum cytokines and decreased functional activated T and B cells, suggesting that astilbin treatment decreases the capacity of B cells to stimulate T lymphocytes⁽¹⁵³⁾.

Flavonoids: flavanols. Several studies suggest that the regular consumption of tea, the most widely consumed beverage in the world after water, is associated with an array of health benefits.

In particular, epidemiological studies indicate that the incidence of death due to heart, cerebrovascular and respiratory diseases is considerably lower in China and Japan, the two leading green tea-consuming countries⁽¹⁵⁴⁻¹⁵⁶⁾. All tea is derived from Camellia sinensis (L.) kuntze, an evergreen shrub of the Theaceae family, whose health benefits are mainly associated with its high concentration of flavanols known as catechins⁽¹⁵⁴⁾. Epigallocatechin gallate (EGCG), the major bioactive polyphenol present in green tea, has been reported to have antioxidant and anti-inflammatory effects by inhibiting NF-κB and suppressing T-cell activation⁽¹⁵⁷⁾. A number of studies have proved the beneficial effects of green tea or EGCG supplementation in SLE models. An in vivo study demonstrated that a diet supplemented with green tea powder increased mouse survival and improved SLE progression by decreasing serum levels of anti-DNA antibodies and renal damage⁽¹⁵⁸⁾. Furthermore, daily treatment with EGCG by oral administration in lupus-prone mice had prophylactic effects by promoting the Nrf-2 antioxidant signalling pathway, inhibiting NLR family pyrin domain-containing 3 (NLRP3) inflammasome activation in the kidney and enhancing Treg activity⁽¹⁵⁹⁾. The potential therapeutic role of EGCG has also been studied in mesangial cells, which are macrophage-like cells resident in the kidney with immune and vascular functions, from lupus mice pretreated with EGCG and stimulated with lipopolysaccharide (LPS) or IFN-y. EGCG activated the metabolic regulator adenosine 5'-monophosphate-activated protein kinase (AMPK), which has been shown to inhibit the production of several proinflammatory mediators, and blocked the induced expression of iNOS and NO and IL-6 production via a mechanism partially independent of the AMPK activation. Furthermore, EGCG attenuated inflammation via the immune-stimulated phosphoinositide 3-kinase/Akt/mammalian target of rapamycin (PI3K-Akt-mTOR) pathway by decreasing phosphorylation of the kinase Akt⁽¹⁶⁰⁾. EGCG has also been reported to decrease the expression of different autoantigens at the mRNA and/or protein levels in an in vivo model of normal human primary epidermal keratinocytes, a cell type particularly affected in patients with SLE, reducing the expression of several antinuclear autoantibodies⁽¹⁶¹⁾. All these data suggest the potential therapeutic role of EGCG as an important component in novel approaches to SLE management by regulating inflammation and preventing lupus nephritis.

Flavonoids: isoflavones. Nowadays, it is clear that high oestrogen levels exacerbate SLE symptoms and kidney disease and increase numbers of autoreactive B cells and autoantibodies⁽¹⁶²⁾. Phyto-oestrogens are plant-derived compounds that are found in a wide variety of foods and that are structurally and/or functionally similar to mammalian oestrogens and their active metabolites. Phyto-oestrogens can interact with oestrogen receptors (ER) with oestrogenic or anti-oestrogen activities. Genistein and daidzein are the major bioactive isoflavones, and they are mainly available in soya, the cornerstone of the traditional Asian diet. Soya has shown anti-inflammatory effects and is able to reduce proteinuria and renal pathological lesions associated with progressive renal failure, which could be beneficial for SLE. *In vivo* studies showed that

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dietary supplementation with soya isoflavones extracted from natural soya germ, containing 50% daidzin, 30% glycitin and 20% genistein, alleviated disease severity in lupus mice and decreased autoantibody levels and in vitro IFN-y production in splenocytes. Furthermore, methanol extracts of soya isoflavones demonstrated a higher affinity for ER β than ER α , suggesting that ER activation is selectively modulated⁽¹⁶³⁾. These results are consistent with those of previous studies, which proved that SLE disease severity is aggravated by ER α agonists but ameliorated by ER β agonists in a murine lupus model⁽¹⁶⁴⁾. By contrast, a previous study showed that a diet supplemented with 20% soyabean protein and 5% soyabean oil exacerbated the SLE clinical course in autoimmune MRL/lpr mice, decreasing survival and increasing renal damage, thymus weight and T-cell proliferation in the spleen and B cells in lymph nodes. Thus, further research on the mechanism underlying the effect of soya-rich diets is necessary to evaluate the potential for isoflavone supplementation in patients with SLE⁽¹⁶⁵⁾. However, in recent years isoflavone research in SLE has not evolved so far. Even though animal data showed that isoflavones have a wide range of molecular, cellular and behavioural effects at doses and plasma concentrations attainable in humans, the consumption of sova or sova phyto-oestrogen has produced mixed and often conflicting results and potential adverse effects on humans, which may limit their use in further studies^(166,167).

Cournestrol is the major phyto-oestrogen in lucerne sprouts but can be also found in beans and other vegetables. Dietary supplementation with coumestrol in SLE mice ameliorated some aspects of the disease⁽¹⁶⁸⁾. Furthermore, a diet supplemented with lucerne sprout ethyl acetate extract significantly improved proteinuria, prolonged lifespan and suppressed the production of proinflammatory cytokines in stimulated peritoneal cells and splenocytes from SLE mice. However, there was no change in SLE autoantibody production, suggesting that its beneficial effects might be more attributable to the down-regulation of inflammatory cytokines rather than autoantibody regulation. Nevertheless, a group of mice fed a diet supplemented with lucerne sprouts had similar proteinuria progression and lifespan as the lupus group, suggesting that the ingestion of whole lucerne sprouts had no beneficial effects on SLE. Accordingly, lucerne sprout ethyl acetate extract reduced serum levels of proinflammatory cytokines and improved survival after 9h of LPS challenge, suggesting that lucerne sprout ethyl acetate extract alleviated acute inflammatory hazards. Moreover, pretreatment with lucerne sprout ethyl acetate extract significantly inhibited proinflammatory cytokine production in LPS-stimulated primary macrophage^(169,170). By contrast, previous reports have shown that lucerne seed or sprouts can induce SLE-like disease in monkeys and exacerbate disease severity in patients with SLE who have ingested lucerne tablets^(171,172).

Lignans. Flaxseed (*Linum usitatissimum* L.), the richest source of lignan precursors, has been a part of the human diet for thousands of years. The principal dietary lignan precursor in flaxseed is the secoisolariciresinol diglucoside (SDG), which has demonstrated possible nutraceutical actions to prevent and alleviate lifestyle-related diseases⁽¹⁷³⁾. A flaxseed-supplemented diet in a lupus mouse model improved renal damage and decreased splenic lymphocyte proliferation⁽¹⁷⁴⁾. Similarly, oral administration with SDG from flaxseed showed dose-dependent protective

effects in the kidney, with similar effects to those previously noted for dietary supplementation with flaxseed⁽¹⁷⁵⁾. In a randomised cross-over trial, flaxseed supplementation over 17 weeks was well tolerated by patients with lupus nephritis and exerted significant positive effects on renal function⁽¹⁷⁶⁾. In another randomised cross-over trial, daily supplementation with flaxseed in patients with SLE appeared to be renoprotective at the end of the 2-year study⁽¹⁷⁷⁾.

Diarylheptanoids and arylalkanones. Curcumin (1,7-bis-[4-hydroxy-3-methoxyphenyl]-1,6-heptadiene-3,5-dione), a component of the rhizomes of the golden spice turmeric (Curcuma longa L.), has demonstrated antioxidant, anti-tumour and anti-inflammatory activities. Recent studies have suggested that curcumin may have therapeutic effects in SLE treatment. In in vitro studies, low doses of curcumin modulated the Th17/Treg balance of CD4⁺ T cells isolated from patients with SLE without affecting CD4⁺ T cells of healthy subjects. As SLE pathogenesis involves the enhancement of Th17 cells with a parallel reduction in the Treg population in patients with SLE, the regulation of this balance has become a subject of interest in SLE management, which is intended to improve the clinical outcome and prevent organ damage⁽¹⁷⁸⁾. Curcumin has also shown beneficial effects in SLE mouse models, producing a delayed onset of autoantibody production and proteinuria and significantly decreasing salivary gland infiltration and lymphadenopathy compared with the control group⁽¹⁷⁹⁾. Moreover, a curcumin-supplemented diet improved renal damage and autoantibody and proinflammatory cytokine production in female lupus mice. Interestingly, these therapeutic effects disappeared after Treg depletion by anti-CD25 antibody injection, suggesting that the protective effects of curcumin involve, at least partially, its interaction with Tregs⁽¹⁸⁰⁾. In addition, a randomised and placebo-controlled study demonstrated that oral supplementation with curcumin (three capsules per d over 3 months, each capsule containing 500 mg of turmeric (22.1 mg of curcumin)) has beneficial effects without side effects in lupus patients with refractory nephritis, suggesting that short-term curcumin supplementation may be used safely as an adjuvant therapy in patients with SLE⁽¹⁸¹⁾. However, curcumin (30 mg/kg body weight administered from 12 to 20 weeks by daily intraperitoneal injection) showed negative effects in the central nervous system of MRL/lpr mice, increasing IgG and anxiety behaviour and decreasing central complement protein (C3) deposits in the brain and circulation. Curcumin also worsened brain atrophy, reduced the volume of the hippocampus and increased the mRNA and protein expression of aquaporin 4, the main water channel of the brain, aggravating brain oedema. Furthermore, increased splenocyte proliferation was suggestive of peripheral immune response activation. Therefore, until a safe dose range is established by additional studies, caution is warranted in the use of curcumin even as adjuvant therapy for central nervous system lupus⁽¹⁸²⁾.

Stilbenes. Resveratrol (3,5,4'-trihydroxystilbene) is a natural compound found in various plants and fruits, especially in grapes, that possess anti-inflammatory, immune-regulatory, antioxidant and blood fat-regulatory activities. In particular, resveratrol can modulate inflammatory genes and signalling transcription factors,

such as STAT3, NF- κ B and COX-2, with critical roles in SLE pathogenesis⁽¹⁸³⁾. A recent study showed that resveratrol had protective effects in a pristane-induced lupus BALB/c mouse model after 7 months of treatment and *in vitro* immunomodulatory effects on splenic mononuclear cells, suggesting that it may represent a novel approach for the management of SLE⁽¹⁸⁴⁾.

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Indole-3-carbinol. Indole-3-carbinol (I3C), the breakdown product of glucobrassicin, is obtained from the dietary consumption of cruciferous vegetables (family Brassicaceae), such as broccoli, cabbage, Brussels sprouts and cauliflower. An in vivo study showed that I3C dietary supplementation (0.2 g/kg) increased the lifespan of lupus-prone NZB/W F1 mice and decreased proteinuria, glomerulonephritis and interstitial nephritis. Furthermore, I3C supplementation was effective when initiated before or after disease onset and may be beneficial in SLE prevention as well as in the early stages of SLE. In addition, the ratio the urine oestrogen metabolites 2-hydroxyestrone of (2-OH) and 16 α -hydroxyestrone (16 α -OH) was greater after the I3C diet, showing anti-oestrogenic activities⁽¹⁸⁵⁾. Similarly, another study demonstrated that a diet enriched with 2000 ppm I3C preserved renal function by decreasing proteinuria and renal pathological abnormalities and prolonged survival when fed to NZB/W F₁ mice for 40 weeks. I3C supplementation decreased anti-dsDNA, anti-chromatin and anti-RNA helicase A antibody levels; increased the ratio of 2-OH to 16α -OH metabolites by maintaining 2-OH oestrone levels; and blocked B- and T-cell maturation, inhibiting disease progression. In addition, in vitro concanavalin A stimulation of T cells from the spleen of I3C-diet mice produced Th1 cytokines (IL-2 and IFN-y) in contrast to the higher production of Th2 cytokines (IL-4 and IL-10) in the SLE control animals⁽¹⁸⁶⁾. Oestrogen metabolism in women with SLE is weighted towards 16a-OH, which might increase SLE disease activity. In an open-label 1-week metabolic study in women with SLE, the consumption of 375 mg/d of I3C changed the ratio of urinary 16a-OH to 2-OH predominantly owing to an increase in 2-hydroxylation, demonstrating that I3C could play a role in the therapy of SLE by attenuating oestrogen-dependent disease activity⁽¹⁸⁷⁾. These studies provide a preliminary understanding of mechanisms whereby I3C leads to the amelioration of lupus-like disease and suggest that I3C may be an effective adjunctive therapy for the human disease.

Melatonin. Melatonin (*N*-acetyl-5-methoxytryptamine) is an indoleamine synthesised from the essential dietary amino acid tryptophan, which can be found in a several components of the diet, for example, fruits, vegetables, olive oil and nuts. Consuming such foods gives rise to a significant increase in serum melatonin levels^(188,189). Melatonin concentrations are decreased in lupus patients and inversely correlated with disease activity. Melatonin administration has shown immunomodulatory effects, stimulating the immune system under basal immunosuppressive conditions and inhibiting exacerbated immune responses⁽¹⁹⁰⁾. Administration of melatonin to MRL/lpr mice at a dose of 30 mg/kg body weight in drinking water over 1 month improved the histological kidney pattern in females and worsened it in males. Moreover, female mice treated with melatonin showed

a decrease in serum IgG, IgM, anti-dsDNA and anti-CII autoantibodies, proinflammatory cytokines (IL-2, IL-6, IFN-y, TNF- α and IL-1 β) and nitrite/nitrate production and an increase in anti-inflammatory cytokines (IL-10). However, in male mice the treatment with melatonin elicited the opposite effect, worsening all the immunological parameters with elevated autoantibodies titres and a prevalence of proinflammatory v. anti-inflammatory cytokines. Similar results were obtained when lymphocytes from the spleen and lymph nodes were cultured: the indoleamine decreased the proinflammatory cytokines and increased the anti-inflammatory cytokines produced by lymphocytes in females but had the opposite effect in males, suggesting that melatonin action in MRL/lpr mice is sex dependent, probably through the modulation and inhibition of sex hormones⁽¹⁹¹⁾. In MRL/lpr mice treated with the same amount of melatonin and hormone therapy, adding testosterone to female mice and oestradiol to males, melatonin alone prevented lupus development in females, but the addition of testosterone to melatonin-treated female mice produced a similar negative pattern to that observed previously in male lupus mice. These effects were similar in oestradiol-treated males, confirming that melatonin action in MRL/lpr mice could be sex dependent through the modulation of sex hormones⁽¹⁹²⁾. Furthermore, melatonin treatment (0.01, 0.1, 1.0 mg/kg) in pristane-induced SLE BALB/c mice antagonised the increasing levels of IgM, anti-single-stranded DNA and histone autoantibodies and decreased renal lesions caused by pristane. In an in vitro experiment with concanavalin A- or LPS-stimulated splenocytes isolated from melatonin-treated mice, melatonin decreased induced IL-6 and IL-13 production and increased IL-2 levels, showing that the indoleamine could modulate the disturbance of the cytokine network in SLE mice by regulating the Th1/Th2 cytokine imbalance⁽¹⁹³⁾. In vitro administration of melatonin $(1 \times 10^{-4} \text{ M})$ in peripheral mononuclear cells from treated patients with SLE increased the number of Tregs expressing FOXP3 and BAFF mRNA expression in patients with SLE, whereas it caused up-regulation of BAFF mRNA levels in healthy subjects, supporting the dual role of melatonin in the cells of patients v. controls and its potential use as an immunomodulatory therapy or co-therapy for SLE⁽¹⁹⁴⁾.

Conclusion

The increasing body of evidence linking the intake of different nutrients to the potential beneficial effects derived from their antioxidant, anti-inflammatory and immunomodulatory properties suggests a possible supportive role of diet therapy in the primary and secondary prevention and management of SLE. Over previous years, human and mouse models have supplied several dietary candidates and have shed light on the possible mechanisms underlying the response to different nutrients, with promising results.

Although the studies are not conclusive and further research is required, it seems to be clear that a balanced diet can be helpful in the prevention and management of SLE, contributing to the management of the disease activity as well as the reduction of co-morbidities, thus improving health and quality of life of patients with SLE. In particular, widespread evidence highlighted the importance of a diet rich in vitamins (mainly A, B₆, C, D and E) and MUFA/PUFA (particularly n-3 PUFA and MUFA) with an adequate fibre intake, protein and Na restriction and moderate energy consumption in reducing co-morbidities and preventing SLE flares, thus minimising unnecessary burden in patients with SLE. It is also remarkable the promising role of dietary polyphenols included in the diet through vegetables, fruits, cereals, legumes and drinks such as tea and wine in the management of SLE. Likewise, it is important to encourage patients to stop smoking, avoid being overweight and optimise their blood pressure, lipid profile, and control of disease activity to decrease cardiovascular morbidity.

Currently, researchers continue to search and identify potential dietary candidates that could play beneficial roles in SLE. Nevertheless, despite the positive preliminary results obtained over the last years, most of the reported beneficial effects need further verification before they can be translated into clinical practice. The efficacy of these dietary candidates and other nutrients, as well as the effect of dietary interventions aimed at promoting adequate nutritional status in SLE patients, requires further evaluation through prospective and randomised trials in human subjects.

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References

- Noble PW, Bernatsky S, Clarke AE, et al. (2016) DNAdamaging autoantibodies and cancer: the lupus butterfly theory. Nat Rev Rheumatol 12, 429–434.
- Tan EM, Cohen AS, Fries JF, *et al.* (1982) The 1982 revised criteria for the classification of systemic lupus erythematosus. *Arthritis Rheum* 25, 1271–1277.
- Crispín JC, Liossis SN, Kis-Toth K, *et al.* (2010) Pathogenesis of human systemic lupus erythematosus: recent advances. *Trends Mol Med* 16, 47–57.
- Lisnevskaia L, Murphy G & Isenberg D (2014) Systemic lupus erythematosus. *Lancet* 384, 1878–1888.
- Petri M, Orbai AM, Alarcón GS, *et al.* (2012) Derivation and validation of the Systemic Lupus International Collaborating Clinics classification criteria for systemic lupus erythematosus. *Arthritis Rheum* 64, 2677–2686.
- Vina ER, Utset TO, Hannon MJ, et al. (2014) Racial differences in treatment preferences among lupus patients: a two-site study. Clin Exp Rheumatol 32, 680–688.
- 7. Mina R & Brunner HI (2013) Update on differences between childhood-onset and adult-onset systemic lupus erythematosus. *Arthritis Res Ther* **15**, 218.

- Borba EF, Carvalho JF & Bonfá E (2006) Mechanisms of dyslipoproteinemias in systemic lupus erythematosus. *Clin Dev Immunol* 13, 203–208.
- 9. Moser KL, Kelly JA, Lessard CJ, *et al.* (2009) Recent insights into the genetic basis of systemic lupus erythematosus. *Genes Immun* **10**, 373–379.
- Xiong J, He Z, Zeng X, *et al.* (2014) Association of vitamin D receptor gene polymorphisms with systemic lupus erythematosus: a meta-analysis. *Clin Exp Rheumatol* 32, 174–181.
- 11. Ruiz-Larrañaga O, Migliorini P, Uribarri M, *et al.* (2016) Genetic association study of systemic lupus erythematosus and disease subphenotypes in European populations. *Clin Rheumatol* **35**, 1161–1168.
- Mirabelli G, Cannarile F, Bruni C, *et al.* (2015) One year in review 2015: systemic lupus erythematosus. *Clin Exp Rheumatol* 33, 414–425.
- 13. Cai L, Zhang JW, Xue XX, *et al.* (2014) Meta-analysis of associations of IL1 receptor antagonist and estrogen receptor gene polymorphisms with systemic lupus erythematosus susceptibility. *PLOS ONE* **9**, e109712.
- Mak A & Tay SH (2014) Environmental factors, toxicants and systemic lupus erythematosus. *Int J Mol Sci* 15, 16043–16056.
- 15. Selmi C (2010) Autoimmunity in 2009. *Autoimmun Rev* **9**, 795–800.
- Pan Y, Ke H, Yan Z, *et al.* (2016) The Western-type diet induces anti-HMGB1 autoimmunity in *Apoe^{-/−}* mice. *Atherosclerosis* **251**, 31–38.
- Mu Q, Zhang H & Luo XM (2015) SLE: another autoimmune disorder influenced by microbes and diet? *Front Immunol* 6, 608.
- Rodríguez Huerta MD, Trujillo-Martín MM, Rúa-Figueroa Í, et al. (2016) Healthy lifestyle habits for patients with systemic lupus erythematosus: a systemic review. Semin Arthritis Rheum 45, 463–470.
- Podolska MJ, Biermann MH, Maueröder C, *et al.* (2015) Inflammatory etiopathogenesis of systemic lupus erythematosus: an update. *J Inflamm Res* 8, 161–171.
- Dolff S, Bijl M, Huitema MG, *et al.* (2011) Disturbed Th1, Th2, Th17 and T(reg) balance in patients with systemic lupus erythematosus. *Clin Immunol* **141**, 197–204.
- 21. Chun HY, Chung JW, Kim HA, *et al.* (2007) Cytokine IL-6 and IL-10 as biomarkers in systemic lupus erythematosus. *J Clin Immunol* **27**, 461–466.
- 22. Ganguly D, Haak S, Sisirak V, *et al.* (2013) The role of dendritic cells in autoimmunity. *Nat Rev Immunol* **13**, 566–577.
- Wu M, Yang J, Li X, *et al.* (2016) The role of γδ T cells in systemic lupus erythematosus. *J Immunol Res* 2016, 2932531.
- Nalbandian A, Crispín JC & Tsokos GC (2009) Interleukin-17 and systemic lupus erythematosus: current concepts. *Clin Exp Immunol* 157, 209–215.
- Gaffen SL (2008) An overview of IL-17 function and signaling. *Cytokine* 43, 402–407.
- Nie J, Li YY, Zheng SG, *et al.* (2015) FOXP3⁺ Treg cells and gender bias in autoimmune diseases. *Front Immunol* 6, 493.
- Bloch O, Amit-Vazina M, Yona E, *et al.* (2014) Increased ERK and JNK activation and decreased ERK/JNK ratio are associated with long-term organ damage in patients with systemic lupus erythematosus. *Rheumatology (Oxford)* 53, 1034–1042.
- Kuryłowicz A & Nauman J (2008) The role of nuclear factor-κB in the development of autoimmune diseases: a link between genes and environment. *Acta Biochim Pol* 55, 629–647.

- Kawasaki M, Fujishiro M, Yamaguchi A, et al. (2011) Possible role of the JAK/STAT pathways in the regulation of T cell-interferon related genes in systemic lupus erythematosus. *Lupus* 20, 1231–1239.
- 30. Zhao J, Wang H, Huang Y, *et al.* (2015) Lupus nephritis: glycogen synthase kinase 3β promotion of renal damage through activation of the NLRP3 inflammasome in lupus-prone mice. *Arthritis Rheumatol* **67**, 1036–1044.
- Telles RW, Ferreira GA, da Silva NP, *et al.* (2010) Increased plasma myeloperoxidase levels in systemic lupus erythematosus. *Rheumatol Int* **30**, 779–784.
- Squatrito D, Emmi G, Silvestri E, et al. (2014) Pathogenesis and potential therapeutic targets in systemic lupus erythematosus: from bench to bedside. Auto Immun Highlights 5, 33–45.
- 33. Kuhn A, Bonsmann G, Anders HJ, *et al.* (2015) The diagnosis and treatment of systemic lupus erythematosus. *Dtsch Arztebl Int* **112**, 423–432.
- Jordan N & D'Cruz D (2015) Key issues in the management of patients with systemic lupus erythematosus: latest developments and clinical implications. *Ther Adv Musculoskelet Dis* 7, 234–246.
- 35. Bertsias GK, Tektonidou M, Amoura Z, *et al.* (2012) Joint European League Against Rheumatism and European Renal Association-European Dialysis and Transplant Association (EULAR/ERA-EDTA) recommendations for the management of adult and paediatric lupus nephritis. *Ann Rheum Dis* **71**, 1771–1782.
- 36. Buttgereit F, Saag KG, Cutolo M, *et al.* (2005) The molecular basis for the effectiveness, toxicity, and resistance to glucocorticoids: focus on the treatment of rheumatoid arthritis. *Scand J Rheumatol* **34**, 14–21.
- Ruiz-Irastorza G, Danza A & Khamashta M (2012) Glucocorticoid use and abuse in SLE. *Rheumatology* (Oxford) **51**, 1145–1153.
- Gerstein HC, Thorpe KE, Taylor DW, *et al.* (2002) The effectiveness of hydroxychloroquine in patients with type 2 diabetes mellitus who are refractory to sulfonylureas – a randomized trial. *Diabetes Res Clin Pract* 55, 209–219.
- Wallace DJ, Metzger AL, Stecher VJ, *et al.* (1990) Cholesterollowering effect of hydroxychloroquine in patients with rheumatic disease: reversal of deleterious effects of steroids on lipids. *Am J Med* **89**, 322–326.
- Borba EF & Bonfá E (2001) Longterm beneficial effect of chloroquine diphosphate on lipoprotein profile in lupus patients with and without steroid therapy. *J Rheumatol* 28, 780–785.
- 41. Bertsias G, Ioannidis JP, Boletis J, *et al.* (2008) EULAR recommendations for the management of systemic lupus erythematosus. Report of a Task Force of the EULAR Standing Committee for International Clinical Studies Including Therapeutics. *Ann Rheum Dis* **67**, 195–205.
- Hahn BH, McMahon MA, Wilkinson A, et al. (2012) American College of Rheumatology guidelines for screening, treatment, and management of lupus nephritis. Arthritis Care Res (Hoboken) 64, 797–808.
- Jordan N, Lutalo PM & D'Cruz DP (2013) Novel therapeutic agents in clinical development for systemic lupus erythematosus. *BMC Med* 11, 120.
- Hui-Yuen JS, Reddy A, Taylor J, *et al.* (2015) Safety and efficacy of belimumab to treat systemic lupus erythematosus in academic clinical practices. *J Rheumatol* 42, 2288–2295.
- 45. Vilas-Boas A, Morais SA & Isenberg DA (2015) Belimumab in systemic lupus erythematosus. *RMD Open* **1**, e000011.
- Scheinberg MA, Hislop CM & Martin RS (2016) Blisibimod for treatment of systemic lupus erythematosus: with trials you become wiser. *Expert Opin Biol Ther* 16, 723–733.

- 47. Stohl W, Merrill JT, Looney RJ, *et al.* (2015) Treatment of systemic lupus erythematosus patients with the BAFF antagonist "peptibody" blisibimod (AMG 623/A-623): results from randomized, double-blind phase 1a and phase 1b trials. *Arthritis Res Ther* **17**, 215.
- Isenberg DA, Petri M, Kalunian K, *et al.* (2016) Efficacy and safety of subcutaneous tabalumab in patients with systemic lupus erythematosus: results from ILLUMINATE-1, a 52week, phase III, multicentre, randomised, double-blind, placebo-controlled study. *Ann Rheum Dis* **75**, 323–331.
- 49. Witcher J, Fleischmann R, Chindalore VL, *et al.* (2016) Pharmacokinetics and safety of single doses of tabalumab in subjects with rheumatoid arthritis or systemic lupus erythematosus. *Br J Clin Pharmacol* **81**, 908–917.
- Daikh DI & Wofsy D (2001) Cutting edge: reversal of murine lupus nephritis with CTLA4Ig and cyclophosphamide. *J Immunol* 166, 2913–2916.
- Furie R, Nicholls K, Cheng TT, *et al.* (2014) Efficacy and safety of abatacept in lupus nephritis: a twelve-month, randomized, double-blind study. *Arthritis Rheumatol* 66, 379–389.
- 52. Mathian A, Hie M, Cohen-Aubart F, *et al.* (2015) Targeting interferons in systemic lupus erythematosus: current and future prospects. *Drugs* **75**, 835–846.
- Postal M, Sinicato NA, Appenzeller S, *et al.* (2016) Drugs in early clinical development for systemic lupus erythematosus. *Expert Opin Investig Drugs* 25, 573–583.
- Greco CM, Nakajima C & Manzi S (2013) Updated review of complementary and alternative medicine treatments for systemic lupus erythematosus. *Curr Rheumatol Rep* 15, 378.
- Shah M, Adams-Huet B, Kavanaugh A, *et al.* (2004) Nutrient intake and diet quality in patients with systemic lupus erythematosus on a culturally sensitive cholesterol lowering dietary program. *J Rheumatol* **31**, 71–75.
- Bruce IN (2005) 'Not only...but also': factors that contribute to accelerated atherosclerosis and premature coronary heart disease in systemic lupus erythematosus. *Rheumatology (Oxford)* 44, 1492–1502.
- 57. Chung CP, Avalos I, Oeser A, *et al.* (2007) High prevalence of the metabolic syndrome in patients with systemic lupus erythematosus: association with disease characteristics and cardiovascular risk factors. *Ann Rheum Dis* 66, 208–214.
- Oeser A, Chung CP, Asanuma Y, *et al.* (2005) Obesity is an independent contributor to functional capacity and inflammation in systemic lupus erythematosus. *Arthritis Rheum* 52, 3651–3659.
- Klack K, Bonfa E & Borba Neto EF (2012) Diet and nutritional aspects in systemic lupus erythematosus. *Rev Bras Reumatol* 52, 384–408.
- Minami Y, Sasaki T, Arai Y, *et al.* (2003) Diet and systemic lupus erythematosus: a 4 year prospective study of Japanese patients. *J Rheumatol* **30**, 747–754.
- Borges MC, dos Santos FeM, Telles RW, *et al.* (2012) Nutritional status and food intake in patients with systemic lupus erythematosus. *Nutrition* 28, 1098–1103.
- Fernandes G, Yunis EJ & Good RA (1976) Influence of diet on survival of mice. *Proc Natl Acad Sci U S A* 73, 1279–1283.
- 63. Jolly CA, Muthukumar A, Avula CP, *et al.* (2001) Life span is prolonged in food-restricted autoimmune-prone (NZB × NZW)F(1) mice fed a diet enriched with (*n*-3) fatty acids. *J Nutr* **131**, 2753–2760.
- 64. Muthukumar AR, Jolly CA, Zaman K, *et al.* (2000) Calorie restriction decreases proinflammatory cytokines and polymeric Ig receptor expression in the submandibular glands of autoimmune prone (NZB×NZW)F₁ mice. *J Clin Immunol* **20**, 354–361.

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- Cárdeno A, Sánchez-Hidalgo M & Alarcón-de-la-Lastra C (2013) An up-date of olive oil phenols in inflammation and cancer: molecular mechanisms and clinical implications. *Curr Med Chem* 20, 4758–4776.
- 66. Toledo E, Hu FB, Estruch R, *et al.* (2013) Effect of the Mediterranean diet on blood pressure in the PREDIMED trial: results from a randomized controlled trial. *BMC Med* **11**, 207.
- Casas R, Sacanella E & Estruch R (2014) The immune protective effect of the Mediterranean diet against chronic low-grade inflammatory diseases. *Endocr Metab Immune Disord Drug Targets* 14, 245–254.
- Miggiano GA & Gagliardi L (2005) Diet, nutrition and rheumatoid arthritis [article in Italian]. *Clin Ter* 156, 115–123.
- McKellar G, Morrison E, McEntegart A, *et al.* (2007) A pilot study of a Mediterranean-type diet intervention in female patients with rheumatoid arthritis living in areas of social deprivation in Glasgow. *Ann Rheum Dis* 66, 1239–1243.
- Tselios K, Koumaras C, Gladman DD, et al. (2016) Dyslipidemia in systemic lupus erythematosus: just another comorbidity? Semin Arthritis Rheum 45, 604–610.
- Ardoin SP, Sandborg C & Schanberg LE (2007) Management of dyslipidemia in children and adolescents with systemic lupus erythematosus. *Lupus* 16, 618–626.
- 72. Borba EF & Bonfá E (1997) Dyslipoproteinemias in systemic lupus erythematosus: influence of disease, activity, and anticardiolipin antibodies. *Lupus* **6**, 533–539.
- Calder PC (2010) The 2008 ESPEN Sir David Cuthbertson Lecture: fatty acids and inflammation – from the membrane to the nucleus and from the laboratory bench to the clinic. *Clin Nutr* 29, 5–12.
- 74. Simopoulos AP (2002) Omega-3 fatty acids in inflammation and autoimmune diseases. *J Am Coll Nutr* **21**, 495–505.
- 75. Elkan AC, Anania C, Gustafsson T, *et al.* (2012) Diet and fatty acid pattern among patients with SLE: associations with disease activity, blood lipids and atherosclerosis. *Lupus* **21**, 1405–1411.
- 76. Arriens C, Hynan LS, Lerman RH, *et al.* (2015) Placebocontrolled randomized clinical trial of fish oil's impact on fatigue, quality of life, and disease activity in systemic lupus erythematosus. *Nutr J* **14**, 82.
- Lozovoy MA, Simão AN, Morimoto HK, *et al.* (2015) Fish oil *n*-3 fatty acids increase adiponectin and decrease leptin levels in patients with systemic lupus erythematosus. *Mar Drugs* 13, 1071–1083.
- Wright SA, O'Prey FM, McHenry MT, et al. (2008) A randomised interventional trial of omega-3-polyunsaturated fatty acids on endothelial function and disease activity in systemic lupus erythematosus. Ann Rheum Dis 67, 841–848.
- Duffy EM, Meenagh GK, McMillan SA, *et al.* (2004) The clinical effect of dietary supplementation with omega-3 fish oils and/or copper in systemic lupus erythematosus. *J Rheumatol* **31**, 1551–1556.
- 80. Pestka JJ, Vines LL, Bates MA, *et al.* (2014) Comparative effects of *n*-3, *n*-6 and *n*-9 unsaturated fatty acid-rich diet consumption on lupus nephritis, autoantibody production and CD4⁺ T cell-related gene responses in the autoimmune NZBWF₁ mouse. *PLOS ONE* **9**, e100255.
- Halade GV, Rahman MM, Bhattacharya A, *et al.* (2010) Docosahexaenoic acid-enriched fish oil attenuates kidney disease and prolongs median and maximal life span of autoimmune lupus-prone mice. *J Immunol* 184, 5280–5286.
- Fenton JI, Hord NG, Ghosh S, *et al.* (2013) Immunomodulation by dietary long chain omega-3 fatty acids and the potential for adverse health outcomes. *Prostaglandins Leukot Essent Fatty Acids* 89, 379–390.

- Chin S, Liu W, Storkson J, *et al.* (1992) Dietary sources of conjugated dienoic isomers of linoleic acids, a newly recognized class of anticarcinogens. *J Fodd Compons Anal* 5, 185–197.
- Hennessy AA, Ross PR, Fitzgerald GF, *et al.* (2016) Sources and bioactive properties of conjugated dietary fatty acids. *Lipids* **51**, 377–397.
- Pariza MW, Park Y & Cook ME (2000) Mechanisms of action of conjugated linoleic acid: evidence and speculation. *Proc Soc Exp Biol Med* **223**, 8–13.
- Bassaganya-Riera J, Hontecillas R & Beitz DC (2002) Colonic anti-inflammatory mechanisms of conjugated linoleic acid. *Clin Nutr* 21, 451–459.
- Bergamo P, Luongo D, Maurano F, *et al.* (2006) Conjugated linoleic acid enhances glutathione synthesis and attenuates pathological signs in MRL/MpJ-Fas^{lpr} mice. *J Lipid Res* 47, 2382–2391.
- Bergamo P, Maurano F & Rossi M (2007) Phase 2 enzyme induction by conjugated linoleic acid improves lupusassociated oxidative stress. *Free Radic Biol Med* 43, 71–79.
- Yang M & Cook ME (2003) Dietary CLA decreased weight loss and extended survival following the onset of kidney failure in NZB/W F₁ mice. *Lipids* 38, 21–24.
- 90. Kritchevsky D, Tepper SA, Wright S, *et al.* (2004) Conjugated linoleic acid isomer effects in atherosclerosis: growth and regression of lesions. *Lipids* **39**, 611–616.
- McLeod RS, LeBlanc AM, Langille MA, et al. (2004) Conjugated linoleic acids, atherosclerosis, and hepatic very-low-density lipoprotein metabolism. Am J Clin Nutr 79, 11698–11748.
- 92. Yang M, Pariza MW & Cook ME (2000) Dietary conjugated linoleic acid protects against end stage disease of systemic lupus erythematosus in the NZB/W F₁ mouse. *Immunopharmacol Immunotoxicol* **22**, 433–449.
- 93. Lambert EV, Goedecke JH, Bluett K, et al. (2007) Conjugated linoleic acid versus high-oleic acid sunflower oil: effects on energy metabolism, glucose tolerance, blood lipids, appetite and body composition in regularly exercising individuals. Br J Nutr 97, 1001–1011.
- Tricon S, Burdge GC, Williams CM, *et al.* (2005) The effects of conjugated linoleic acid on human health-related outcomes. *Proc Nutr Soc* 64, 171–182.
- Nugent AP, Roche HM, Noone EJ, et al. (2005) The effects of conjugated linoleic acid supplementation on immune function in healthy volunteers. Eur J Clin Nutr 59, 742–750.
- 96. Gaullier JM, Halse J, Høivik HO, *et al.* (2007) Six months supplementation with conjugated linoleic acid induces regional-specific fat mass decreases in overweight and obese. *Br J Nutr* **97**, 550–560.
- 97. Carvalho RF, Uehara SK & Rosa G (2012) Microencapsulated conjugated linoleic acid associated with hypocaloric diet reduces body fat in sedentary women with metabolic syndrome. *Vasc Health Risk Manag* **8**, 661–667.
- 98. Benjamin S, Prakasan P, Sreedharan S, *et al.* (2015) Pros and cons of CLA consumption: an insight from clinical evidences. *Nutr Metab (Lond)* **12**, 4.
- 99. Harbige LS (2003) Fatty acids, the immune response, and autoimmunity: a question of *n*-6 essentiality and the balance between *n*-6 and *n*-3. *Lipids* **38**, 323–341.
- Lin BF, Jeng SJ, Chiang BL, *et al.* (1997) Dietary fat affects lipids and anti-cardiolipin antibody levels in autoimmuneprone NZB/W F₁ mice. *Br J Nutr* 77, 657–669.
- 101. Rosillo M, Alcaraz MJ, Sánchez-Hidalgo M, et al. (2014) Anti-inflammatory and joint protective effects of extra-virgin olive-oil polyphenol extract in experimental arthritis. J Nutr Biochem 25, 1275–1281.

- Rosillo MA, Sánchez-Hidalgo M, Sánchez-Fidalgo S, et al. (2016) Dietary extra-virgin olive oil prevents inflammatory response and cartilage matrix degradation in murine collageninduced arthritis. Eur J Nutr 55, 315–325.
- 103. Aparicio-Soto M, Sánchez-Hidalgo M, Cárdeno A, et al. (2016) Dietary extra virgin olive oil attenuates kidney injury in pristane-induced SLE model via activation of HO-1/Nrf-2 antioxidant pathway and suppression of JAK/ STAT, NF-κB and MAPK activation. J Nutr Biochem 27, 278–288.
- Milovanov IS, Lysenko LV, Milovanova LIu, *et al.* (2009) The role of balanced low-protein diet in inhibition of predialysis chronic kidney disease progression in patients with systemic diseases [article in Russian]. *Ter Arkh* 81, 52–57.
- Caetano MC, Ortiz TT, Terreri MT, *et al.* (2009) Inadequate dietary intake of children and adolescents with juvenile idiopathic arthritis and systemic lupus erythematosus. *J Pediatr (Rio J)* 85, 509–515.
- Mannoor MK, Shimabukuro I, Tsukamotoa M, *et al.* (2009) Honeybee royal jelly inhibits autoimmunity in SLE-prone NZB × NZW F₁ mice. *Lupus* 18, 44–52.
- 107. Huang CY, Hsu TC, Kuo WW, *et al.* (2009) Beneficial effects of taurine on cardiac abnormality in NZB/W F₁ mice fed with a high-cholesterol diet. *J Agric Food Chem* **57**, 8635–8642.
- Hsu TC, Chiang SY, Wu JH, et al. (2008) Treatment with taurine attenuates hepatic apoptosis in NZB/W F₁ mice fed with a high-cholesterol diet. J Agric Food Chem 56, 9685–9691.
- Peters H, Border WA, Rückert M, *et al.* (2003) L-Arginine supplementation accelerates renal fibrosis and shortens life span in experimental lupus nephritis. *Kidney Int* 63, 1382–1392.
- 110. Corman LC (1985) The role of diet in animal models of systemic lupus erythematosus: possible implications for human lupus. *Semin Artbritis Rheum* **15**, 61–69.
- 111. Minami Y, Hirabayashi Y, Nagata C, *et al.* (2011) Intakes of vitamin B₆ and dietary fiber and clinical course of systemic lupus erythematosus: a prospective study of Japanese female patients. *J Epidemiol* **21**, 246–254.
- 112. Gergely P, Niland B, Gonchoroff N, *et al.* (2002) Persistent mitochondrial hyperpolarization, increased reactive oxygen intermediate production, and cytoplasmic alkalinization characterize altered IL-10 signaling in patients with systemic lupus erythematosus. *J Immunol* **169**, 1092–1101.
- 113. Bethunaickan R, Sahu R, Liu Z, *et al.* (2012) Anti-tumor necrosis factor α treatment of interferon- α -induced murine lupus nephritis reduces the renal macrophage response but does not alter glomerular immune complex formation. *Arthritis Rheum* **64**, 3399–3408.
- 114. Perl A (2013) Oxidative stress in the pathology and treatment of systemic lupus erythematosus. *Nat Rev Rheumatol* **9**, 674–686.
- 115. Liao X, Ren J, Wei CH, *et al.* (2015) Paradoxical effects of all-*trans*-retinoic acid on lupus-like disease in the MRL/lpr mouse model. *PLOS ONE* **10**, e0118176.
- Kinoshita K, Kishimoto K, Shimazu H, et al. (2010) Successful treatment with retinoids in patients with lupus nephritis. Am J Kidney Dis 55, 344–347.
- 117. Nozaki Y, Yamagata T, Yoo BS, *et al.* (2005) The beneficial effects of treatment with all-*trans*-retinoic acid plus corticosteroid on autoimmune nephritis in NZB/WF mice. *Clin Exp Immunol* **139**, 74–83.
- 118. Handono K, Firdausi SN, Pratama MZ, *et al.* (2016) Vitamin A improve Th17 and Treg regulation in systemic lupus erythematosus. *Clin Rheumatol* **35**, 631–638.

- 119. Pérez de Lema G, Lucio-Cazaña FJ, Molina A, *et al.* (2004) Retinoic acid treatment protects MRL/lpr lupus mice from the development of glomerular disease. *Kidney Int* **66**, 1018–1028.
- Ikeda T, Nishide T, Ohtani T, *et al.* (2005) The effects of vitamin A derivative etretinate on the skin of MRL mice. *Lupus* 14, 510–516.
- 121. Petri M, Roubenoff R, Dallal GE, et al. (1996) Plasma homocysteine as a risk factor for atherothrombotic events in systemic lupus erythematosus. *Lancet* 348, 1120–1124.
- 122. Evans MD, Cooke MS, Akil M, *et al.* (2000) Aberrant processing of oxidative DNA damage in systemic lupus erythematosus. *Biochem Biophys Res Commun* **273**, 894–898.
- 123. Dusso AS & Brown AJ (1998) Mechanism of vitamin D action and its regulation. *Am J Kidney Dis* **32**, S13–S24.
- Lane NE (2010) Vitamin D and systemic lupus erythematosus: bones, muscles, and joints. *Curr Rheumatol Rep* 12, 259–263.
- Kamen DL & Aranow C (2008) The link between vitamin D deficiency and systemic lupus erythematosus. *Curr Rheumatol Rep* 10, 273–280.
- Borba VZ, Vieira JG, Kasamatsu T, *et al.* (2009) Vitamin D deficiency in patients with active systemic lupus erythematosus. *Osteoporos Int* 20, 427–433.
- 127. Marques CD, Dantas AT, Fragoso TS, *et al.* (2010) The importance of vitamin D levels in autoimmune diseases. *Rev Bras Reumatol* **50**, 67–80.
- 128. Wahono CS, Rusmini H, Soelistyoningsih D, *et al.* (2014) Effects of 1,25(OH)₂D₃ in immune response regulation of systemic lupus erithematosus (SLE) patient with hypovitamin D. *Int J Clin Exp Med* 7, 22–31.
- Casella CB, Seguro LP, Takayama L, *et al.* (2012) Juvenile onset systemic lupus erythematosus: a possible role for vitamin D in disease status and bone health. *Lupus* 21, 1335–1342.
- Caetano M, Terreri MT, Ortiz T, *et al.* (2015) Bone mineral density reduction in adolescents with systemic erythematosus lupus: association with lack of vitamin D supplementation. *Clin Rheumatol* 34, 2065–2070.
- 131. Marinho A, Carvalho C, Boleixa D, *et al.* (2016) Vitamin D supplementation, effects on FoxP3 expression in T cells and FoxP3⁺/IL-17A ratio and clinical course in systemic lupus erythematosus patients: a study in a Portuguese cohort. *Immunol Res* (epublication ahead of print version 16 July 2016).
- Lemire JM, Ince A & Takashima M (1992) 1,25-Dihydroxyvitamin D3 attenuates the expression of experimental murine lupus of MRL/l mice. *Autoimmunity* 12, 143–148.
- 133. Lavi Arab F, Rastin M, Faraji F, *et al.* (2015) Assessment of 1,25-dihydroxyvitamin D3 effects on Treg cells in a mouse model of systemic lupus erythematosus. *Immunopharma-col Immunotoxicol* **37**, 12–18.
- 134. Tabasi N, Rastin M, Mahmoudi M, *et al.* (2015) Influence of vitamin D on cell cycle, apoptosis, and some apoptosis related molecules in systemic lupus erythematosus. *Iran J Basic Med Sci* **18**, 1107–1111.
- 135. Reynolds JA, Haque S, Williamson K, *et al.* (2016) Vitamin D improves endothelial dysfunction and restores myeloid angiogenic cell function via reduced CXCL-10 expression in systemic lupus erythematosus. *Sci Rep* **6**, 22341.
- 136. Lima GL, Paupitz J, Aikawa NE, *et al.* (2016) Vitamin D supplementation in adolescents and young adults with juvenile systemic lupus erythematosus for improvement in disease activity and fatigue scores: a randomized, double-blind, placebo-controlled trial. *Artbritis Care Res* (*Hoboken*) **68**, 91–98.

- M. Aparicio-Soto et al.
- Azrielant S & Shoenfeld Y (2016) Eppur Si Muove: vitamin D is essential in preventing and modulating SLE. *Lupus* 25, 563–572.
- 138. Kennel KA, Drake MT & Hurley DL (2010) Vitamin D deficiency in adults: when to test and how to treat. *Mayo Clin Proc* **85**, 752–757; quiz 757–758.
- 139. Hsieh CC & Lin BF (2005) The effects of vitamin E supplementation on autoimmune-prone New Zealand black \times New Zealand white F₁ mice fed an oxidised oil diet. *Br J Nutr* **93**, 655–662.
- Hsieh CC & Lin BF (2005) Opposite effects of low and high dose supplementation of vitamin E on survival of MRL/ *lpr* mice. *Nutrition* 21, 940–948.
- 141. Maeshima E, Liang XM, Goda M, et al. (2007) The efficacy of vitamin E against oxidative damage and autoantibody production in systemic lupus erythematosus: a preliminary study. Clin Rheumatol 26, 401–404.
- 142. Costenbader KH, Kang JH & Karlson EW (2010) Antioxidant intake and risks of rheumatoid arthritis and systemic lupus erythematosus in women. *Am J Epidemiol* **172**, 205–216.
- 143. Tam LS, Li EK, Leung VY, *et al.* (2005) Effects of vitamins C and E on oxidative stress markers and endothelial function in patients with systemic lupus erythematosus: a double blind, placebo controlled pilot study. *J Rheumatol* **32**, 275–282.
- 144. O'Dell JR, McGivern JP, Kay HD, *et al.* (1988) Improved survival in murine lupus as the result of selenium supplementation. *Clin Exp Immunol* **73**, 322–327.
- 145. Yang X, Yao G, Chen W, *et al.* (2015) Exacerbation of lupus nephritis by high sodium chloride related to activation of SGK1 pathway. *Int Immunopharmacol* **29**, 568–573.
- 146. Brown AC (2000) Lupus erythematosus and nutrition: a review of the literature. *J Ren Nutr* **10**, 170–183.
- D'Archivio M, Filesi C, Di Benedetto R, et al. (2007) Polyphenols, dietary sources and bioavailability. Ann Ist Super Sanita 43, 348–361.
- D'Archivio M, Santangelo C, Scazzocchio B, *et al.* (2008) Modulatory effects of polyphenols on apoptosis induction: relevance for cancer prevention. *Int J Mol Sci* 9, 213–228.
- Woodman OL & Chan ECH (2004) Vascular and anti-oxidant actions of flavonols and flavones. *Clin Exp Pharmacol Physiol* **31**, 786–790.
- 150. Kang HK, Ecklund D, Liu M, et al. (2009) Apigenin, a non-mutagenic dietary flavonoid, suppresses lupus by inhibiting autoantigen presentation for expansion of autoreactive Th1 and Th17 cells. Arthritis Res Ther 11, R59.
- 151. Hsu HC, Yang P, Wang J, *et al.* (2008) Interleukin 17producing T helper cells and interleukin 17 orchestrate autoreactive germinal center development in autoimmune BXD2 mice. *Nat Immunol* **9**, 166–175.
- Haas C, Ryffel B & Le Hir M (1998) IFN-γ receptor deletion prevents autoantibody production and glomerulonephritis in lupus-prone (NZB×NZW)F₁ mice. *J Immunol* 160, 3713–3718.
- 153. Guo L, Liu W, Lu T, *et al.* (2015) Decrease of functional activated T and B cells and treatment of glomerulonephitis in lupus-prone mice using a natural flavonoid astilbin. *PLOS ONE* **10**, e0124002.
- 154. Khan N & Mukhtar H (2013) Tea and health: studies in humans. *Curr Pharm Des* **19**, 6141–6147.
- 155. Saito E, Inoue M, Sawada N, *et al.* (2015) Association of green tea consumption with mortality due to all causes and major causes of death in a Japanese population: the Japan Public Health Center-based Prospective Study (JPHC Study). *Ann Epidemiol* **25**, 512–518.e513.
- Hsu S & Dickinson D (2006) A new approach to managing oral manifestations of Sjogren's syndrome and skin manifestations of lupus. *J Biochem Mol Biol* **39**, 229–239.

- 157. Singh BN, Shankar S & Srivastava RK (2011) Green tea catechin, epigallocatechin-3-gallate (EGCG): mechanisms, perspectives and clinical applications. *Biochem Pharmacol* **82**, 1807–1821.
- 158. Sayama K, Oguni I, Tsubura A, *et al.* (2003) Inhibitory effects of autoimmune disease by green tea in MRL-Faslprcg/Faslprcg mice. *In Vivo* **17**, 545–552.
- 159. Tsai PY, Ka SM, Chang JM, *et al.* (2011) Epigallocatechin-3-gallate prevents lupus nephritis development in mice via enhancing the Nrf2 antioxidant pathway and inhibiting NLRP3 inflammasome activation. *Free Radic Biol Med* **51**, 744–754.
- Peairs A, Dai R, Gan L, *et al.* (2010) Epigallocatechin-3-gallate (EGCG) attenuates inflammation in MRL/lpr mouse mesangial cells. *Cell Mol Immunol* 7, 123–132.
- 161. Hsu S, Dickinson DP, Qin H, *et al.* (2005) Inhibition of autoantigen expression by (–)-epigallocatechin-3-gallate (the major constituent of green tea) in normal human cells. *J Pharmacol Exp Ther* **315**, 805–811.
- Fairweather D, Frisancho-Kiss S & Rose NR (2008) Sex differences in autoimmune disease from a pathological perspective. *Am J Pathol* **173**, 600–609.
- Hong Y, Wang T, Huang C, *et al.* (2008) Soy isoflavones supplementation alleviates disease severity in autoimmuneprone MRL-lpr/lpr mice. *Lupus* 17, 814–821.
- 164. Li J & McMurray RW (2007) Effects of estrogen receptor subtype-selective agonists on autoimmune disease in lupus-prone NZB/NZW F₁ mouse model. *Clin Immunol* 123, 219–226.
- Zhao JH, Sun SJ, Horiguchi H, et al. (2005) A soy diet accelerates renal damage in autoimmune MRL/Mp-lpr/ lpr mice. Int Immunopharmacol 5, 1601–1610.
- Patisaul HB & Jefferson W (2010) The pros and cons of phytoestrogens. *Front Neuroendocrinol* **31**, 400–419.
- 167. Bedell S, Nachtigall M & Naftolin F (2014) The pros and cons of plant estrogens for menopause. *J Steroid Biochem Mol Biol* **139**, 225–236.
- Schoenroth LJ, Hart DA, Pollard KM, *et al.* (2004) The effect of the phytoestrogen coumestrol on the NZB/W F₁ murine model of systemic lupus. *J Autoimmun* 23, 323–332.
- 169. Hong YH, Chao WW, Chen ML, et al. (2009) Ethyl acetate extracts of alfalfa (*Medicago sativa* L.) sprouts inhibit lipopolysaccharide-induced inflammation in vitro and in vivo. J Biomed Sci 16, 64.
- 170. Hong YH, Huang CJ, Wang SC, *et al.* (2009) The ethyl acetate extract of alfalfa sprout ameliorates disease severity of autoimmune-prone MRL-lpr/lpr mice. *Lupus* **18**, 206–215.
- 171. Malinow MR, Bardana EJ, Pirofsky B, et al. (1982) Systemic lupus erythematosus-like syndrome in monkeys fed alfalfa sprouts: role of a nonprotein amino acid. Science 216, 415–417.
- 172. Roberts JL & Hayashi JA (1983) Exacerbation of SLE associated with alfalfa ingestion. *N Engl J Med* **308**, 1361.
- 173. Imran M, Ahmad N, Anjum FM, *et al.* (2015) Potential protective properties of flax lignan secoisolariciresinol diglucoside. *Nutr J* **14**, 71.
- Hall AV, Parbtani A, Clark WF, *et al.* (1993) Abrogation of MRL/lpr lupus nephritis by dietary flaxseed. *Am J Kidney Dis* 22, 326–332.
- 175. Clark WF, Muir AD, Westcott ND, *et al.* (2000) A novel treatment for lupus nephritis: lignan precursor derived from flax. *Lupus* **9**, 429–436.
- Clark WF, Parbtani A, Huff MW, et al. (1995) Flaxseed: a potential treatment for lupus nephritis. *Kidney Int* 48, 475–480.

- 177. Clark WF, Kortas C, Heidenheim AP, *et al.* (2001) Flaxseed in lupus nephritis: a two-year nonplacebo-controlled crossover study. *J Am Coll Nutr* **20**, 143–148.
- 178. Handono K, Pratama MZ, Endharti AT, *et al.* (2015) Treatment of low doses curcumin could modulate Th17/Treg balance specifically on CD4⁺ T cell cultures of systemic lupus erythematosus patients. *Cent Eur J Immunol* **40**, 461–469.
- 179. Kurien BT, Harris VM, Quadri SM, et al. (2015) Significantly reduced lymphadenopathy, salivary gland infiltrates and proteinuria in MRL-lpr/lpr mice treated with ultrasoluble curcumin/turmeric: increased survival with curcumin treatment. Lupus Sci Med 2, e000114.
- Lee H, Kim H, Lee G, *et al.* (2013) Curcumin attenuates lupus nephritis upon interaction with regulatory T cells in New Zealand black/white mice. *Br J Nutr* **110**, 69–76.
- 181. Khajehdehi P, Zanjaninejad B, Aflaki E, *et al.* (2012) Oral supplementation of turmeric decreases proteinuria, hematuria, and systolic blood pressure in patients suffering from relapsing or refractory lupus nephritis: a randomized and placebo-controlled study. *J Ren Nutr* **22**, 50–57.
- Foxley S, Zamora M, Hack B, *et al.* (2013) Curcumin aggravates CNS pathology in experimental systemic lupus erythematosus. *Brain Res* 1504, 85–96.
- Nakata R, Takahashi S & Inoue H (2012) Recent advances in the study on resveratrol. *Biol Pharm Bull* 35, 273–279.
- Wang ZL, Luo XF, Li MT, *et al.* (2014) Resveratrol possesses protective effects in a pristane-induced lupus mouse model. *PLOS ONE* 9, e114792.
- Auborn KJ, Qi M, Yan XJ, et al. (2003) Lifespan is prolonged in autoimmune-prone (NZB/NZW) F₁ mice fed a diet supplemented with indole-3-carbinol. J Nutr 133, 3610–3613.
- Yan XJ, Qi M, Telusma G, et al. (2009) Indole-3-carbinol improves survival in lupus-prone mice by inducing tandem B- and T-cell differentiation blockades. *Clin Immunol* 131, 481–494.
- McAlindon TE, Gulin J, Chen T, *et al.* (2001) Indole-3-carbinol in women with SLE: effect on estrogen metabolism and disease activity. *Lupus* 10, 779–783.
- Peuhkuri K, Sihvola N & Korpela R (2012) Dietary factors and fluctuating levels of melatonin. *Food Nutr Res* 56, 10.3402/fnr.v56i0.17252.
- 189. Sae-Teaw M, Johns J, Johns NP, *et al.* (2013) Serum melatonin levels and antioxidant capacities after consumption of pineapple, orange, or banana by healthy male volunteers. *J Pineal Res* **55**, 58–64.
- Robeva R, Tanev D, Kirilov G, *et al.* (2013) Decreased daily melatonin levels in women with systemic lupus erythematosus – a short report. *Balkan Med J* 30, 273–276.
- 191. Jimenez-Caliani AJ, Jimenez-Jorge S, Molinero P, et al. (2006) Sex-dependent effect of melatonin on systemic erythematosus lupus developed in Mrl/Mpj-Fas^{lpr} mice: it ameliorates the disease course in females, whereas it exacerbates it in males. *Endocrinology* 147, 1717–1724.
- 192. Jimenez-Caliani AJ, Jimenez-Jorge S, Molinero P, et al. (2008) Treatment with testosterone or estradiol in melatonin treated females and males MRL/MpJ-Fas^{lpr} mice induces negative effects in developing systemic lupus erythematosus. J Pineal Res 45, 204–211.
- Zhou LL, Wei W, Si JF, et al. (2010) Regulatory effect of melatonin on cytokine disturbances in the pristaneinduced lupus mice. *Mediators Inflamm* **2010**, 951210.
- 194. Medrano-Campillo P, Sarmiento-Soto H, Álvarez-Sánchez N, et al. (2015) Evaluation of the immunomodulatory effect of melatonin on the T-cell response in peripheral blood from systemic lupus erythematosus patients. J Pineal Res 58, 219–226.

- Clark WF & Parbtani A (1994) Omega-3 fatty acid supplementation in clinical and experimental lupus nephritis. *Am J Kidney Dis* 23, 644–647.
- 196. Walton AJ, Snaith ML, Locniskar M, *et al.* (1991) Dietary fish oil and the severity of symptoms in patients with systemic lupus erythematosus. *Ann Rheum Dis* **50**, 463–466.
- 197. Kim YJ, Yokozawa T & Chung HY (2005) Effects of energy restriction and fish oil supplementation on renal guanidino levels and antioxidant defences in aged lupus-prone B/W mice. *Br J Nutr* **93**, 835–844.
- Muthukumar A, Zaman K, Lawrence R, et al. (2003) Food restriction and fish oil suppress atherogenic risk factors in lupus-prone (NZB×NZW) F₁ mice. J Clin Immunol 23, 23–33.
- 199. Bhattacharya A, Lawrence RA, Krishnan A, et al. (2003) Effect of dietary n-3 and n-6 oils with and without food restriction on activity of antioxidant enzymes and lipid peroxidation in livers of cyclophosphamide treated autoimmune-prone NZB/ W female mice. J Am Coll Nutr 22, 388–399.
- 200. Chang SC, Chiang BL, Wu WM, *et al.* (1999) Different dietary fats influence serum and tissue lipids and anticardiolipin antibody levels in autoimmune-prone NZB/W F₁ mice. *Br J Nutr* **81**, 331–340.
- 201. Chandrasekar B, Troyer DA, Venkatraman JT, *et al.* (1995) Dietary omega-3 lipids delay the onset and progression of autoimmune lupus nephritis by inhibiting transforming growth factor β mRNA and protein expression. *J Autoimmun* **8**, 381–393.
- 202. Chandrasekar B & Fernandes G (1994) Decreased proinflammatory cytokines and increased antioxidant enzyme gene expression by omega-3 lipids in murine lupus nephritis. *Biochem Biophys Res Commun* **200**, 893–898.
- Robinson DR, Prickett JD, Polisson R, *et al.* (1985) The protective effect of dietary fish oil on murine lupus. *Prostaglandins* **30**, 51–75.
- 204. Venkatraman JT & Chu WC (1999) Effects of dietary ω3 and ω6 lipids and vitamin E on proliferative response, lymphoid cell subsets, production of cytokines by spleen cells, and splenic protein levels for cytokines and oncogenes in MRL/ MpJ-*lpr/lpr* mice. *J Nutr Biochem* **10**, 582–597.
- Spurney RF, Ruiz P, Albrightson CR, *et al.* (1994) Fish oil feeding modulates leukotriene production in murine lupus nephritis. *Prostaglandins* 48, 331–348.
- 206. Robinson DR, Prickett JD, Makoul GT, *et al.* (1986) Dietary fish oil reduces progression of established renal disease in (NZB×NZW)F₁ mice and delays renal disease in BXSB and MRL/1 strains. *Arthritis Rheum* **29**, 539–546.
- 207. Halade GV, Williams PJ, Veigas JM, et al. (2013) Concentrated fish oil (Lovaza®) extends lifespan and attenuates kidney disease in lupus-prone short-lived (NZB×NZW)F₁ mice. Exp Biol Med (Maywood) 238, 610–622.
- Godfrey DG, Stimson WH, Watson J, *et al.* (1986) Effects of dietary supplementation on autoimmunity in the MRL/lpr mouse: a preliminary investigation. *Ann Rheum Dis* 45, 1019–1024.
- 209. Handono K, Sidarta YO, Pradana BA, et al. (2014) Vitamin D prevents endothelial damage induced by increased neutrophil extracellular traps formation in patients with systemic lupus erythematosus. Acta Med Indones 46, 189–198.
- Ben-Zvi I, Aranow C, Mackay M, *et al.* (2010) The impact of vitamin D on dendritic cell function in patients with systemic lupus erythematosus. *PLoS ONE* 5, e9193.
- Weimann BJ & Hermann D (1999) Inhibition of autoimmune deterioration in MRL/lpr mice by vitamin E. *Int J Vitam Nutr Res* 69, 255–261.