



Seroprevalence of *Toxoplasma gondii* and associated risk factors for infection in the Netherlands: third cross-sectional national study

Original Paper

Cite this article: van den Berg OE, Stanoeva KR, Zonneveld R, Hoek-van Deursen D, van der Klis FR, van de Kasstele J, Franz E, Opsteegh M, Friesema IHM and Kortbeek LM (2023). Seroprevalence of *Toxoplasma gondii* and associated risk factors for infection in the Netherlands: third cross-sectional national study. *Epidemiology and Infection*, **151**, e136, 1–8
<https://doi.org/10.1017/S095026882300122X>

Received: 19 January 2023

Revised: 10 July 2023

Accepted: 24 July 2023




Keywords:

risk factors; serology; seroprevalence; The Netherlands; *Toxoplasma gondii*

Corresponding author:

Oda E. van den Berg;

Email: oda.van.den.berg@rivm.nl

Oda E. van den Berg¹ , Kamelia R. Stanoeva^{1,2} , Rens Zonneveld^{1,3}, Denise Hoek-van Deursen¹, Fiona R. van der Klis¹, Jan van de Kasstele¹, Eelco Franz¹, Marieke Opsteegh¹, Ingrid H. M. Friesema¹  and Laetitia M. Kortbeek¹

¹Centre for Infectious Disease Control, National Institute for Public Health and the Environment (RIVM), Bilthoven, The Netherlands; ²European Public Health Microbiology Training Programme (EUPHEM), European Centre for Disease Prevention and Control (ECDC), Stockholm, Sweden and ³Department of Medical Microbiology and Infection Prevention, Academic Medical Centre, Amsterdam University Medical Centre, Amsterdam, The Netherlands

Abstract

A third nationally representative serosurvey was performed to study the changes in *Toxoplasma gondii* (*T. gondii*) seroprevalence in the Netherlands over a 20-year time span and to identify and confirm risk factors for acquired toxoplasmosis. This cross-sectional study (conducted in 2016/2017) was designed similarly to the previous two studies (1995/1996 and 2006/2007) and included a questionnaire and serum sampling among Dutch residents. Factors associated with seropositivity for *T. gondii* were determined using multivariable analysis of the questionnaire-derived data. The earlier observed decrease in *T. gondii* seroprevalence between 1995/1996 and 2006/2007 (from 40.5% to 26.0%) did not continue into 2016/2017 (29.9%). Similarly to the previous studies, the seroprevalence increased with age and varied among regions. In all studies, higher *T. gondii* seropositivity was associated with increasing age, lower educational level, not living in the Southeast, and eating raw or semi-cooked pork. The incidence of congenital toxoplasmosis was estimated at 1.3/1000 (95% CI 0.9–1.8) live-born children in 2017. As the seroprevalence of *T. gondii* in the Netherlands did not decrease over the last decade, an increase in public health awareness is needed and prevention measures may need to be taken to achieve a further reduction in *T. gondii* infections in the Netherlands.

Introduction

The protozoan *Toxoplasma gondii* (*T. gondii*) is a zoonotic and ubiquitous parasite infecting more than one-third of the global human population [1]. Warm-blooded animals are hosts, but only Felidae, such as domestic cats, are definitive hosts and shed oocysts into the environment. Human infection occurs through the consumption of tissue cysts in undercooked meat from livestock, through the ingestion of sporulated oocysts from contaminated water or food, and, much less frequently, through the transplantation of organs with tissue cysts [2]. When immunocompetent people become infected, the disease is most typically asymptomatic or mild with flu-like symptoms [2]. There is a small risk of developing ocular toxoplasmosis, which can lead to blindness [3].

T. gondii infection may have severe consequences for two main risk groups: pregnant women and immunocompromised individuals [3]. Transplacental transmission of *T. gondii* may result in spontaneous abortion, prematurity, stillbirth, or neurological or ocular damage to the foetus [4]. Eye lesions from congenital infection are often not identified at birth but occur in 20–80% of congenitally infected persons by adulthood [3]. In immunocompromised people, toxoplasmosis almost always happens as a result of the reactivation of latent infection and can result in neurological symptoms, including headache, disorientation, drowsiness, hemiparesis, reflex changes, and convulsions [5]. Acute acquired *T. gondii* infection in immunocompromised patients may also involve encephalitis, pneumonia, retinochoroiditis, and other disseminated systemic diseases [6].

Risk factors for detectable *T. gondii* antibodies were described in two earlier Dutch serosurveys and included increasing age, urban living, low educational level, cat ownership, raw meat consumption, and gardening [7, 8]. Exactly how these risk factors relate to different routes of transmission remains under debate. The association between *T. gondii* seroprevalence and cat ownership indicates the relevance of the oocyst-based transmission route in human infection [9]. However, the association of the consumption of raw meat with human infection indicates a relevant tissue cyst-based transmission route [10].

© The Author(s), 2023. Published by Cambridge University Press. This is an Open Access article, distributed under the terms of the Creative Commons Attribution licence (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted re-use, distribution and reproduction, provided the original article is properly cited.

The estimated incidence of congenital toxoplasmosis (CT) and overall toxoplasmosis-related disability-adjusted life years (DALYs) in the Netherlands are relatively high, which warrants continuous evaluation of seroprevalence and risk factors [11]. Many European countries have reported various degrees of decrease in seroprevalence between 1980 and recent years, either in the general population or in women of reproductive age [12, 13]. Here, we present data from the third national serosurvey collected in 2016–2017. These data will help monitor age-related seroprevalence dynamics in a 20-year time span, confirm and identify risk factors, and help determine relevant public health interventions.

Material and methods

Study population and questionnaire

The serosurvey, known as PIENTER-3 or P3, was originally conducted by the National Institute for Public Health and the Environment (RIVM) to evaluate the (long-term) epidemiological effects of the nationwide immunisation programme. The acronym PIENTER stands for ‘Peiling immunisatie effect Nederland ter evaluatie van het Rijksvaccinatieprogramma’. This survey follows a similar design to the two previous national serosurveys, PIENTER-1 (P1) in 1995/1996 and PIENTER-2 (P2) in 2006/2007, which are described elsewhere [14–17]. Kortbeek *et al.* [7] published *T. gondii* seroprevalence results for P1 and Hofhuis *et al.* [8] for P2. In short, a two-stage cluster sampling technique was used to draw a sample of Dutch residents aged 0–89 years. Cluster sampling was done to ensure that the included municipalities represented all regions in the Netherlands and all age groups. Forty municipalities were sampled within five regions proportional to size, including an oversampling of people living in low vaccination coverage (LVC) areas and non-Western migrants living in the Netherlands. To estimate seroprevalences, people living in LVC areas were excluded from the national sample (NS). This was done because the NS (including the oversampling of non-Western migrants) is rather comparable to the Dutch population, especially regarding urbanisation degree and religion [17]. As life expectancy is increasing, the maximum age in this study was extended from 79 (in the previous surveys) to 89 years. Sampling was performed independently for P1 to P3. Therefore, the overlap between selected municipalities was only coincidental. The data collection took place from 1 February 2016 to 16 October 2017 and included a questionnaire and serum sampling.

The study proposal was approved by the Medical Ethics Committee Noord-Holland (METC number: M015–022), written informed consent was obtained from all adult participants, and parents or legal guardians of minors were included in the study.

Antibody assay

The same methods for antigen production using *T. gondii* strain and the same batch of freeze-dried control sera were used as in the previous two national serosurveys [7, 8]. Therefore, data from all PIENTER studies can be compared.

After collection, the sera were stored at -20°C until testing. Sera were diluted 1:20 and tested in duplicate for IgG antibodies to *T. gondii* in a sandwich ELISA [18]. The antigen was derived from a crude extract of a *Toxoplasma* RH strain, and the conjugate was a peroxidase-labelled anti-human IgG conjugate (Dako, Denmark). A ratio ≥ 1.0 between the optical density (OD) of the samples and the mean OD of the cut-off controls was considered positive.

Samples with discrepant results for duplicates (i.e. one above and one below the cut-off value) were excluded.

Statistical analysis

To determine the seroprevalence of *T. gondii* IgG antibodies representative of the general population of the Netherlands, seroprevalence was weighted within each municipality for age and gender and also for ethnicity and urbanisation degree, up to their proportion in the total population of the Netherlands (on 1 January 2017) [19]. Seroprevalence analyses were further adjusted for the two-stage cluster sampling by considering the strata (i.e. five regions) and clusters (i.e. 40 municipalities).

To estimate the incidence of live-born children with CT, the number of live-born children in the Netherlands in 2017 per age group of the mother was multiplied by the age-specific force of infection among the susceptible population of women of reproductive age. The mother-to-child transmission per trimester was based on previous studies [20]. The force of infection (FOI) was estimated as the derivative of the age-specific seroprevalence relative to one minus the age-specific seroprevalence [21]. The age-specific seroprevalence was estimated by survey-weighted logistic regression using natural cubic splines with three degrees of freedom. Because of the non-linear relation between prevalence and FOI, the uncertainty in the FOI estimates was calculated using 1,000 Monte Carlo samples from the estimated prevalence. Logistic regression analysis was used to determine whether any of the selected variables were independent predictors of seropositivity for *T. gondii*, after adjustment for age group and gender. In contrast to the weighted seroprevalence estimation, all 5,878 participants were included in the logistic regression to increase power, as living in a low immunisation coverage municipality was not significantly associated with *T. gondii* seropositivity (P -value = 0.154) or with any of the relevant demographics. Variables that reached a significance level of P -value < 0.10 in the univariable analyses for the 0–19 and > 20 years of age groups were selected for inclusion in the multivariable logistic regression model. Odds ratios (ORs) were corrected for gender and age. Models for the 0–19 and > 20 years of age groups were developed using multivariable logistic regression with backwards elimination. A two-sided p -value of ≤ 0.05 was considered statistically significant. Multicollinearity was tested using a correlation matrix and the variance inflation factor. All statistical analyses were performed with R [22].

Results

Prevalence of *Toxoplasma gondii* IgG antibodies

The overall weighted estimate of *T. gondii* seroprevalence for people aged 0–89 years was 30.9% (95% CI 29.4–32.4). The average seroprevalence for children and adolescents aged 0–19 years was 10.9% (95% CI 9.12–12.7) and that for adults aged 20–89 years was 36.7% (95% CI 34.7–38.6). The average seroprevalence for women of reproductive age (15–49 years) was 17.5% (95% CI 15.2–20.1). No differences were found between men and women (31% versus 28%, P -value = 0.18). As can be seen in Figure 1, seroprevalence increased with age, with the steepest slope in people aged 60 to 70 years, resulting in seroprevalences of more than 80% among the highest age groups. The highest seroprevalences were seen in the Northwest and the lowest in the Southeast. The group of 0–19 years old ($n = 989$) showed a higher seroprevalence in the Northwest and the Northeast compared with the Southeast (8%, OR: 2.05 resp. 2.11, P -value = 0.03 resp. 0.02). Figure 2 shows the age-specific FOI in

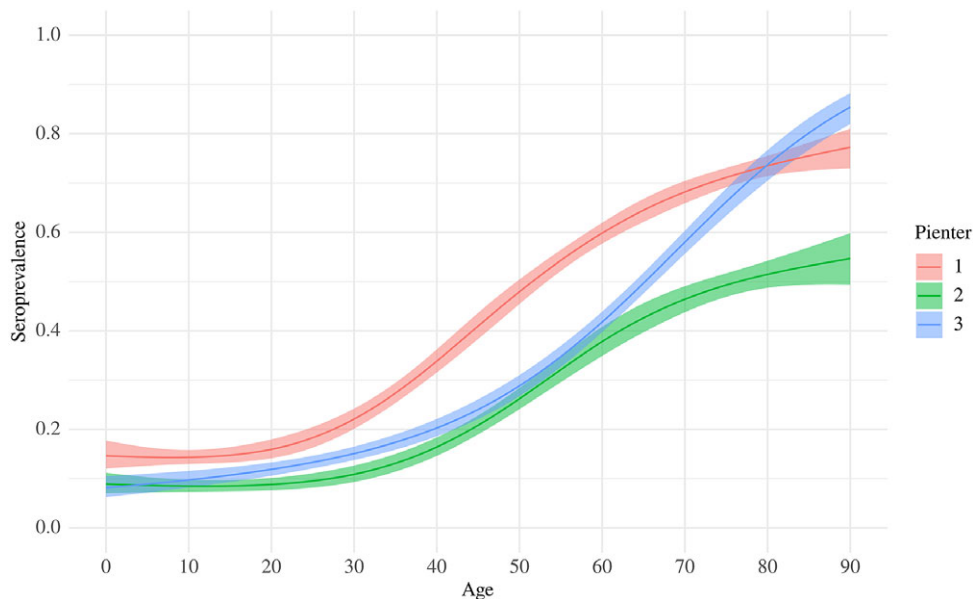


Figure 1. Estimated age-specific weighted prevalence of *Toxoplasma gondii* IgG antibodies in the three Dutch PIENTER serosurveys.

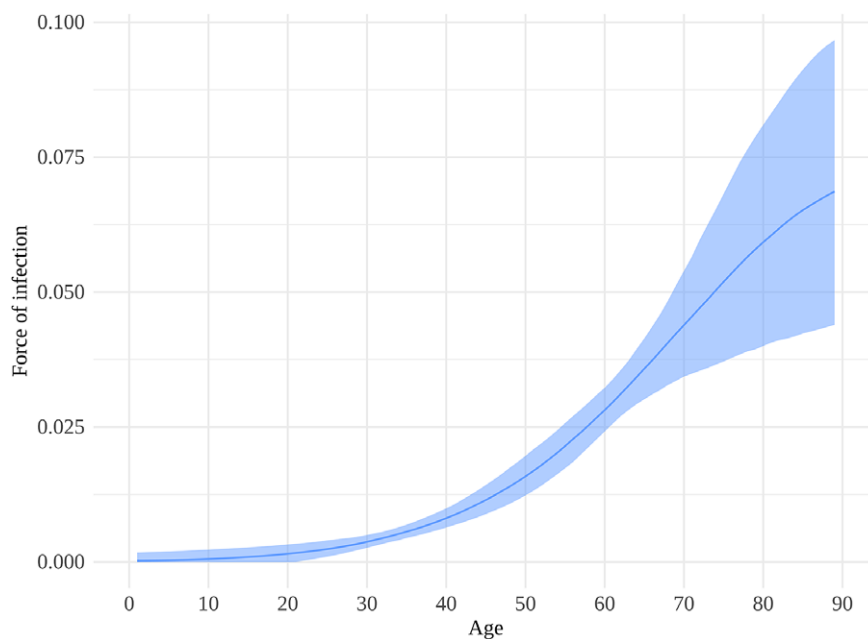


Figure 2. Estimated age-specific force of infection for *Toxoplasma gondii* in women (P3).

women in P3, which was used to estimate the incidence of congenital toxoplasmosis. Based on the FOI in women of reproductive age and the likelihood of transmission per trimester from mother to child based on previous studies [20], the incidence of congenital toxoplasmosis was estimated at 1.3/1000 (95% CI 0.9–1.8) live-born children in 2017.

Predictors of seropositivity for *T. gondii* IgG antibodies

Univariable analyses were performed on all selected variables with a priori adjustment for age and gender (Supplementary Table S1).

Variables that were associated with seropositivity in multivariable analyses are shown in Table 1. For those <20 years of age, age and living in the Northeast or Northwest of the Netherlands were independently associated with *T. gondii* seropositivity. For those aged 20–89 years, independently associated variables included age, not living in the Southeast of the Netherlands, being born outside of the Netherlands, low educational level, eating meat, having kept cattle for the past 5 years, eating raw pork, and having ever been abroad to Asia, Africa, or America. The risk factors associated with seropositivity in participants aged 0–19 and 20–89 years are shown in Table 1.

Table 1. Uncorrected prevalence of specific antibodies to *Toxoplasma gondii* (%) and multivariable logistic regression analyses of risk factors associated with seropositivity in participants aged 0–19 and 20–89 years

| | Age group of 0–19 years (n = 1,250) | | | | Age group of 20–89 years (n = 4,625) | | | |
|--|-------------------------------------|------------------|--|---------|--------------------------------------|------------------|--|---------|
| | N | Prevalence IgG % | Multivariable OR ^a (95% CI) | P-value | N | Prevalence IgG % | Multivariable OR ^a (95% CI) | P-value |
| Geographical region^b | | | | | | | | |
| Southeast | 207 | 8% | 1.0 | | 968 | 26% | 1.0 | |
| Central | 342 | 11% | 1.4 (0.8–2.6) | 0.252 | 1,050 | 34% | 1.6 (1.3–2.0) | <0.001 |
| Northwest | 212 | 15% | 1.9 (1.0–3.6) | 0.046 | 717 | 43% | 2.0 (1.6–2.5) | <0.001 |
| Northeast | 277 | 16% | 2.0 (1.1–3.8) | 0.028 | 891 | 36% | 1.6 (1.3–2.0) | <0.001 |
| Southwest | 212 | 6% | 0.8 (0.4–1.6) | 0.502 | 999 | 32% | 1.7 (1.4–2.1) | <0.001 |
| Country of birth | | | | | | | | |
| The Netherlands | 1,178 | 11% | ns | | 4,027 | 32% | 1.0 | |
| Other | 72 | 15% | | | 595 | 44% | 1.5 (1.2–1.8) | <0.001 |
| Educational level^c | | | | | | | | |
| High | 350 | 9% | ns | | 1,527 | 29% | 1.0 | |
| Medium | 436 | 12% | | | 1,549 | 26% | 1.0 (0.8–1.2) | 0.711 |
| Low | 399 | 11% | | | 1,281 | 49% | 1.3 (1.1–1.6) | 0.001 |
| Eating meat | | | | | | | | |
| No | 22 | 23% | ns | | 102 | 18% | 1.0 | |
| Yes | 1,140 | 11% | | | 4,260 | 33% | 2.1 (1.2–3.8) | 0.008 |
| missing | 88 | 13% | | | 263 | 42% | 3.1 (1.7–5.9) | <0.001 |
| Kept cattle during the past 5 years | | | | | | | | |
| No | 1,228 | 11% | ns | | 4,578 | 33% | 1.0 | |
| Yes | 19 | 26% | | | 43 | 44% | 2.8 (1.4–5.5) | 0.003 |
| Eating raw pork | | | | | | | | |
| No | 867 | 11% | ns | | 2,886 | 32% | 1.0 | |
| <=3 days a month | 137 | 9% | | | 812 | 32% | 1.3 (1.0–1.5) | 0.014 |
| >1 day a week | 19 | 21% | | | 100 | 34% | 1.5 (0.9–2.4) | 0.084 |
| missing | 227 | 13% | | | 827 | 41% | 1.5 (1.3–1.8) | <0.001 |
| Having ever been abroad to Asia, Africa, or America | | | | | | | | |
| No | 867 | 10% | ns | | 2,159 | 31% | 1.0 | |
| Yes | 380 | 12% | | | 2,462 | 36% | 1.2 (1.0–1.3) | 0.0227 |

Abbreviations: 95% CI, confidence interval; OR, odds ratio; ns, not significant.

^aAdjusted for age and gender.

^bThe following categories were used for the educational level of those aged ≥15 years and of parents for those aged <15 years: 'low' (primary school, lower vocational or lower general secondary education), 'medium' (intermediate vocational or intermediate general secondary and higher general secondary education), and 'high' (higher vocational secondary education and university education).

^cThe geographical regions were based on the Dutch provinces: 'Central' (Utrecht and Gelderland), 'Southeast' (Noord-Brabant and Limburg), 'Northwest' (Noord-Holland and Flevoland), 'Southwest' (Zeeland and Zuid-Holland), and 'Northeast' (Groningen, Drenthe, Overijssel, and Friesland).

Comparison with previous national surveys

Table 2 displays the prevalence of *T. gondii* antibodies in 2006/2007 and 2016/2017, categorised by gender, degree of urbanisation, being born in the Netherlands, and ethnicity. For people aged 0–79 years in the Netherlands, the overall weighted estimate of *T. gondii* seroprevalence decreased from 40.5% (95% CI 37.5–43.4) in 1995/1996 to 26.0% (95% CI 24.0–28.0) in 2006/2007 and increased to 29.9% (95% CI 28.3–31.4) in 2016/2017. Including participants aged 80–89 years in 2016/2017 did not significantly change the overall weighted seroprevalence (30.9%; 95% CI 29.4–32.4). The average seroprevalence for women of reproductive age

(15–49 years) decreased from 35.2% (95% CI 32.9–38.6) in 1995/1996 to 18.5% (95% CI 16.2–20.7) in 2006/2007, but remained stable in 2016/2017 (17.5%; 95% CI 15.2–20.1). The incidence of congenital toxoplasmosis shows a similar pattern and was estimated at 3.4/1000 (95% CI 2.8–3.9) live-born children in 1996, 1.5/1000 (95% CI 1.1–2.0) in 2007, and 1.3/1000 (95% CI 0.9–1.8) in 2017. Among people aged 65 years and older, there was no significant difference in seroprevalence between 1996/1997 and 2016/2017. The seroprevalence in the age group of 75–79 years was significantly higher in 2016/2017 compared with 2006/2007 (Figure 1).

Table 2. Prevalence^a of *Toxoplasma gondii* antibodies in 2006/2007 and 2016/2017, stratified for gender, degree of urbanisation, being born in the Netherlands, and ethnicity

| | 2006/2007 (n = 5,541) | | | 2016/2017 (n = 4,759) | | |
|--|-----------------------|----------------|-------------|-----------------------|----------------|-------------|
| | n | Prevalence IgG | | n | Prevalence IgG | |
| | | % | (95% CI) | | % | (95% CI) |
| Overall | 5,541 | 26.0% | (24.0–28.0) | 4,759 | 30.9% | (29.4–32.4) |
| Male | 2,522 | 25.9% | (23.2–28.5) | 2,147 | 31.4% | (28.8–34.2) |
| Female | 3,019 | 26.1% | (24.0–28.2) | 2,621 | 30.3% | (28.3–32.5) |
| Female of reproductive age (15–49) | 1,195 | 18.5% | (16.2–20.7) | 1,339 | 17.3% | (14.8–20.0) |
| Urbanisation ^b | | | | | | |
| Rural area | 1,861 | 24.9% | (21.6–28.3) | 2,178 | 28.7% | (25.5–32.2) |
| Urban area | 3,682 | 26.5% | (23.9–29.2) | 2,581 | 32.6% | (29.8–35.5) |
| Country of birth | | | | | | |
| The Netherlands | 4,681 | 26.3% | (24.4–28.1) | 4,116 | 29.8% | (27.9–31.7) |
| Other | 801 | 24.5% | (19.8–29.1) | 643 | 38.5% | (32.6–44.7) |
| Ethnicity | | | | | | |
| Autochthonous | 4,154 | 27.0% | (25.0–29.1) | 3,758 | 30.6% | (28.7–32.6) |
| Immigrant from Western country | 386 | 25.9% | (22.0–29.9) | 322 | 32.4% | (27.1–38.2) |
| Immigrant from Morocco/Turkey | 328 | 19.4% | (14.4–24.3) | 112 | 33.3% | (22.6–45.4) |
| Immigrant from Surinam/Aruba/Antilles | 336 | 19.3% | (14.5–23.7) | 235 | 30.0% | (24.0–36.6) |
| Immigrant from other non-Western countries | 339 | 18.8% | (15.0–22.5) | 332 | 30.7% | (25.5–36.3) |

CI, confidence interval.

^aWeighted prevalence for age, gender, ethnicity, and urbanisation degree.

^bThe following categories were used for the level of urbanisation: 'urban' (>1,500 addresses/km²) and 'rural' (<1,500 addresses/km²).

For those <20 years of age, 'living in the Northeast' was protective in P1, but was associated with an increased risk in P3. Significant predictors in this age group also included 'country of birth', 'urbanisation degree', and 'gardening' in P1 and 'having kept sheep or cattle in the past 5 years' in P2. None of these variables were significant predictors in P3.

For those 20–79 years of age, 'low educational level' and 'living outside of the Southeast of the Netherlands' were significant predictors in all three surveys. 'Keeping a cat in the past 5 years' was a significant predictor in P1 and P2, but not in P3. 'Eating meat' was only a significant predictor in P3 and 'consumption of raw or undercooked pork' was a significant predictor in P2 and P3, but not in P1. A comparison of the associated risk factors for those aged 20–79 years is shown in Table 3. Extending the maximum age to 89 years in P3 resulted in the same risk factors.

Discussion

Interestingly, the strong decrease in *T. gondii* seroprevalence between 1996/1997 (40.5%) and 2006/2007 (26.0%) did not continue into 2016/2017 (29.9% for 0–79 years of age). The seroprevalence in women of reproductive age in 2016/2017 (17.5%) was lower than in 1995/1996 (35.2%; 95% CI 32.9–38.6), but not different from 2006/2007 (18.5%; 95% CI 16.2–20.7). This was surprising, since other high-income countries have been reporting decreasing *T. gondii* seroprevalences. A national perinatal survey in France, conducted in 1995, 2003, 2010, and 2016, showed a continuously decreasing seroprevalence of *T. gondii* infection among pregnant women from 54% in 1995 to 31% in 2016 [13]. In addition, a

decreasing seroprevalence has been reported in a longitudinal study among immunocompromised patients in Paris [23]. However, due to differences in the study populations and varying consumption habits and soil exposure between countries in general, the relative importance of sources of infection could differ greatly, leading to different trends. Moreover, the seroprevalence in pregnant women in France was much higher to begin with and it is possible that the seroprevalence will plateau at a lower level. In Portugal, a study of the general population showed an overall decrease from 47% in 1979/1980 to 36% in 2002/2003 to 22% in 2013 [24]. However, the difference in time points makes it difficult to compare our data. Based on the age-related increase in seroprevalence in women of reproductive age, the estimated incidence of congenital toxoplasmosis in our study (1.3/1,000 (0.9–1.8) in live-born children in 2017) was relatively high compared with other European countries [13, 25]. However, as women are advised to avoid potential sources of *T. gondii* infection during pregnancy, our method could overestimate the actual incidence of congenital toxoplasmosis.

As noted in the previous national serosurvey studies, seropositivity increased with increasing age. However, until 60 years of age the curve of P3 is more or less comparable to P2, but from 60 years of age and older, there is a substantial increase in seroprevalence and the curve starts moving towards the P1 curve. This may indicate a strong FOI in this age group. However, since the infection is presumed to persist lifelong, this could alternatively be explained by a birth cohort effect. In 1995/1996, the steepest slope was observed in the age group of 25–44 years [7]. This does, 20 years later, partly concern the same birth cohort.

In further agreement with the previous serosurvey studies, seroprevalence was higher in those who had lower education and

Table 3. Comparison of the associated risk factors for those aged 20-79 years between the different PIENTER studies

| | 1995/1996 (n = 7,521) | 2006/2007 (n = 7,030) | 2016/2017 (n = 5,875) |
|--|--------------------------|--------------------------|--------------------------|
| | Odds ratio | Odds ratio | Odds ratio |
| Marital status | | | |
| Married/living together/unmarried | 1.0 | n.s. | N/A |
| Divorced/widowed | 1.4 (1.1–1.7) | | |
| Educational level | | | |
| High | 1.0 | 1.0 | 1.0 |
| Medium | | | 1.0 (0.8–1.1) |
| Low | 1.4 (1.2–1.7) | 1.4 (1.2–1.7) | 1.3 (1.1–1.5) |
| Region | | | |
| Southeast | 1.0 | 1.0 | 1.0 |
| Central | 1.8 (1.5–2.2) | 2.0 (1.5–2.5) | 1.7 (1.4–2.1) |
| Northwest | 3.1 (2.5–3.8) | 2.3 (1.8–3.0) | 1.9 (1.5–2.4) |
| Northeast | 1.8 (1.5–2.1) | 1.5 (1.2–1.9) | 1.7 (1.3–2.1) |
| Southwest | 1.9 (1.6–2.3) | 1.9 (1.5–2.3) | 1.6 (1.3–2.0) |
| Urbanisation^a | | | |
| Very high | sign | sign | n.s. |
| Keeping a cat in the past 5 years | | | |
| No | 1.0 | 1.0 | n.s. |
| Yes | 1.2 (1.1–1.4) | 1.4 (1.2–1.6) | |
| Consumption of raw or undercooked pork | | | |
| No | n.s. | 1.0 | 1.0 |
| Yes | | 1.4 (1.1–1.6) | 1.3 (1.1–1.6) |
| Missing | | | 1.5 (1.2–1.8) |
| Country of birth | | | |
| The Netherlands | n.s. | n.s. | 1.0 |
| Other | | | 1.3 (1.1–1.7) |
| Eating meat | | | |
| No | n.s. | n.s. | 1.0 |
| Yes | | | 2.7 (1.5–5.0) |
| Missing | | | 2.8 (1.6–5.4) |
| Having kept cattle for the past 5 years | | | |
| No | n.s. | n.s. | 1.0 |
| Yes | | | 3.1 (1.5–6.1) |
| Having ever been abroad to Asia, Africa, or America | | | |
| No | n.s. | n.s. | 1.0 |
| Yes | | | 1.2 (1.0–1.3) |

Abbreviations: N/A, not included in the questionnaire; n.s., not significant; sign, significant.
^aUrbanisation degree was categorised in different ways for the different studies and ORs could therefore not be compared.

those who lived in more urbanised areas. In addition, people living in the Southeast of the Netherlands have consistently had a lower seroprevalence compared with the rest of the country since 1995, which may, for example, be due to dietary habits or a lower level of

oocyst contamination in the environment. The inconsistencies in the geographical regions in the younger age group are likely due to the small number of cases.

Transmission of *T. gondii* infection is subject to complex environmental, socioeconomic, and dietary factors and can change over time. In general, the observed decreases in seroprevalences in other countries have partly been attributed to reduced exposure to contaminated meat due to confined husbandry, changes in food storage and preparation, and changing dietary habits [24, 26]. Others have speculated on the influence of changes in attitudes towards cat ownership on environmental contamination [26, 27].

Within the Netherlands, having kept a cat in the past 5 years was a risk factor in P1 and P2, but not in P3. In addition, the consumption of raw or poorly cooked pork was a risk factor in P2 and P3, but not in P1, and eating meat in general was a risk factor only in P3. This could possibly be due to changes in soil or cat faeces-related hygiene and food safety behaviours, indicating a shift from oocyst-based transmission routes to more tissue cyst-based transmissions as the main contributors to human infection. However, this could also be due to a misclassification bias, since infection in seropositive individuals may have happened long before their exposure to risk factors.

Since cats are the only known definitive hosts for *T. gondii*, they are essential to the persistence of *T. gondii* in hosts. In the Netherlands, about 24% of the households owned a cat in 2017 [28] and the overall population of owned cats in the Netherlands was estimated at 3.1 million in 2015 and 2.6 million in 2017 [29]. However, due to cats' fastidious grooming habits, the short duration of oocyst shedding, and the fact that when freshly passed in stools, the oocysts are not infective, and direct contact with cats is not thought to be a risk for human infection [30]. Furthermore, factors such as changes in the management of cat faeces, changes in cat diet, delay in neutering, or increases in stray cats could limit the impact of the decreasing number of owned cats on human seroprevalence [31, 32]. This could explain the lack of decrease in *T. gondii* seroprevalence, despite a decreasing cat population. However, geographically looking at cats and *T. gondii* in the Netherlands, the percentage of households that own a cat is highest in the North and the highest absolute number of cats live in the West, and these regions also showed the highest seroprevalences. Therefore, there could possibly be a geographical link between the density of cats and infection with *T. gondii* in humans, even though direct cat contact may not be the most important risk factor for infection, for example through environmental exposure or consumption of locally produced food products.

Eating raw or undercooked meat has frequently been linked to the presence of antibodies to *T. gondii* [33, 34]. While this overall finding is relatively consistent between studies, the types of meat posing the greatest risk are variable. A systematic review of case-controlled studies published in 2018 found consumption of raw or undercooked beef and sheep meat, but not pork meat, to be important risk factors for *T. gondii* infection [35]. Other cross-sectional studies do find pork meat to be significantly associated with *T. gondii* seropositivity [36, 37].

In general, *T. gondii* infections seem to be most prevalent in sheep meat and least prevalent in beef and seem to differ for pork and chicken, with significantly higher prevalence rates in pork from organic farms compared with pork from conventional pig farming [35, 38]. Despite cattle having a high natural resistance to *T. gondii*, beef can contribute to human infection due to the higher likelihood of being consumed raw or undercooked compared with pork [34]. Therefore, it is difficult to speculate on the proportional

contributions of different types of meat to human infection as dietary habits and environmental factors are constantly changing and differ between countries. In this study, 'eating meat' was a risk factor for *T. gondii* seropositivity, underlining the potential importance of eating meat as a major source of *T. gondii* infection even more. Confined husbandry and home freezing have likely contributed to a reduction in seroprevalence, but changes in dietary habits and the import of non-frozen meat could have counteracted this effect [39, 40]. Potential strategies to try to further reduce exposure to *T. gondii* include educating consumers, combined with improved labelling of the source of meat and the type of processing (i.e. farmed indoors or frozen), and measures to reduce infection in domestic animals (i.e. improved farm hygiene). There will always be trends that influence *T. gondii* infection pressure in different directions, and therefore, it is worthwhile to keep repeating serosurveys like the current one.

One of the strengths of this study is the application of an identical robust design with a nationwide representative sample for the third time with 10-year intervals, enabling the detection of changes in seroprevalence over the years and ensuring maximum comparability with previous studies [41]. In addition, we corrected selective responses using post-stratification weights.

A limitation of the design is that the measurement of seroprevalence is an indicator of *T. gondii*, but does not provide information on the time of infection. There could be a significant delay between infection and serological testing. Consequently, the results are conditional on the absence of any change in the respondents' exposures over time. Lastly, the found association with the variables 'eating meat' and 'eating raw or undercooked pork' has to be interpreted with caution, as missing values within these variables were positively associated with seropositivity. The missingness of these variables was not related to any of the other variables and could be due to residual or unmeasured confounding.

Conclusion

The decrease in *T. gondii* seroprevalence in the Netherlands between 1996/1997 and 2006/2007 stagnated in 2016/2017. Furthermore, 17.5% of women of reproductive age (15–49) were seropositive, leaving the majority of pregnant women susceptible to infection. The risk factor analysis underlines the importance of meat as a major source of *T. gondii* infection. However, it remains difficult to quantify exactly how the different risk factors relate to human infection, as environmental and socioeconomic factors can influence seroprevalences.

Better public awareness might reduce the risk of *T. gondii* and other infections, for instance sufficiently heating and freezing meat. Public health prevention measures may help further mitigate the risk of *T. gondii* infection for the Dutch population.

Supplementary material. The supplementary material for this article can be found at <https://doi.org/10.1017/S095026882300122X>.

Data availability statement. The authors confirm that the data supporting the findings of this study are available within the article and its supplementary materials.

Acknowledgements. The authors thank Eric R.A. Vos for the helpful discussions about oversampling and weighing survey data.

Author contribution. Data curation: K.R.S., I.H.F., L.K., D.H.D., J.v.d.K., O.E.v.d.B.; Investigation: K.R.S., I.H.F., L.K., D.H.D., F.R.v.d.K., R.Z.; Resources:

K.R.S., L.K., D.H.D., F.R.v.d.K., R.Z.; Writing – original draft: K.R.S., R.Z., O.E.v.d.B.; Writing – review & editing: K.R.S., I.H.F., L.K., D.H.D., E.F., F.R.v.d.K., J.v.d.K., M.O., R.Z., O.E.v.d.B.; Conceptualization: I.H.F., L.K., E.F., M.O., O.E.v.d.B.; Formal analysis: I.H.F., J.v.d.K., O.E.v.d.B.; Methodology: I.H.F., J.v.d.K., O.E.v.d.B.; Project administration: I.H.F., E.F., M.O., O.E.v.d.B.; Supervision: I.H.F., L.K., E.F.; Validation: I.H.F., O.E.v.d.B.; Funding acquisition: L.K., E.F.; Software: J.v.d.K.; Visualization: J.v.d.K., O.E.v.d.B.

Financial support. This research received no specific grant from any funding agency, commercial, or not-for-profit sectors.

Competing interest. The authors declare none.

References

- [1] Flegr J, Prandota J, Sovičková M and Israili ZH (2014) Toxoplasmosis - A global threat. Correlation of latent toxoplasmosis with specific disease burden in a set of 88 countries. *PLoS ONE* **9**, e90203.
- [2] Montoya JG and Liesenfeld O (2004) Toxoplasmosis. *The Lancet* **363**, 1965–1976.
- [3] Prevention CfDca. Parasites - Toxoplasmosis (Toxoplasma infection); 2018 [cited 2021 Dec 7]. Available at <https://www.cdc.gov/parasites/toxoplasmosis/>.
- [4] McAuley JB (2014) Congenital toxoplasmosis. *Journal of the Pediatric Infectious Diseases Society* **3**, S30–S35.
- [5] Robert-Gangneux F and Dardé M-L (2012) Epidemiology of and diagnostic strategies for toxoplasmosis. *Clinical Microbiology Reviews* **25**, 264–296.
- [6] Machala L, Kodym P, Malý M, Golenek M, Beran O and Jilich D (2015) Toxoplasmosis in immunocompromised patients. *Epidemiologie, mikrobiologie, imunologie: casopis Společnosti pro epidemiologii a mikrobiologii Ceske lekarske společnosti JE Purkyně*. **64**, 59–65.
- [7] Kortbeek LM, de Melker HE, Veldhuijzen IK and Conyn-van Spaendonck MAE (2004) Population-based toxoplasma seroprevalence study in the Netherlands. *Epidemiology and Infection* **132**, 839–845.
- [8] Hofhuis A, van Pelt W, van Duynhoven YT, Nijhuis CD, Mollema L, van der Klis FR, Havelaar AH and Kortbeek LM (2010) Decreased prevalence and age-specific risk factors for Toxoplasma gondii IgG antibodies in The Netherlands between 1995/1996 and 2006/2007. *Epidemiology and Infection* **139**, 530–538.
- [9] Opsteegh M, Haveman R, Swart AN, Mensink-Beerepoot ME, Hofhuis A, Langelaar MFM and van der Giessen JWB (2012) Seroprevalence and risk factors for toxoplasma gondii infection in domestic cats in the Netherlands. *Preventive Veterinary Medicine* **104**, 317–326.
- [10] Opsteegh M, Prickaerts S, Frankena K and Evers EG (2011) A quantitative microbial risk assessment for meatborne Toxoplasma gondii infection in the Netherlands. *International Journal of Food Microbiology* **150**, 103–114.
- [11] van Lier A, McDonald SA, Bouwknegt M, EPI group, Kretzschmar ME, Havelaar AH, Mangan M-JJ, Wallinga J and de Melker HE (2016) Disease burden of 32 infectious diseases in the Netherlands, 2007–2011. *PLoS One* **11**, e0153106.
- [12] Pappas G, Rousso N and Falagas ME (2009) Toxoplasmosis snapshots: Global status of toxoplasma gondii seroprevalence and implications for pregnancy and congenital toxoplasmosis. *International Journal for Parasitology* **39**, 1385–1394.
- [13] Robinson E, de Valk H, Villena I, le Strat Y and Tourdjman M (2021) National perinatal survey demonstrates a decreasing seroprevalence of Toxoplasma gondii infection among pregnant women in France, 1995 to 2016: Impact for screening policy. *Eurosurveillance* **26**, 1900710.
- [14] De Melker H and Conyn-van Spaendonck M (1998) Immunoserveillance and the evaluation of national immunization programmes: A population-based approach. *Epidemiology & Infection* **121**, 637–643.
- [15] De Melker H, Nagelkerke N and Conyn-van Spaendonck M (2000) Non-participation in a population-based seroprevalence study of vaccine-preventable diseases. *Epidemiology & Infection* **124**, 255–262.

- [16] van der Klis FR, Mollema L, Berbers GA, de Melker HE and Coutinho RA (2009) Second national serum bank for population-based seroprevalence studies in the Netherlands. *The Netherlands Journal of Medicine* **67**, 301–308.
- [17] Verberk JDM, Vos RA, Mollema L, van Vliet J, van Weert JWM, de Melker HE and van der Klis FRM (2019) Third national biobank for population-based seroprevalence studies in the Netherlands, including the Caribbean Netherlands. *BMC Infectious Diseases* **19**, 1–12.
- [18] Ruitenbergh E and Van Knapen F (1977) The enzyme-linked immunosorbent assay and its application to parasitic infections. *Journal of Infectious Diseases* **136**, S267–S273.
- [19] Central Bureau for Statistics (2017) *Regionale kerncijfers Nederland* [Data file]; 2017 [cited on 2021 Dec 7]. Available at <http://www.cbs.nl/>.
- [20] Dunn D, Wallon M, Peyron F, Petersen E, Peckham C and Gilbert R (1999) Mother-to-child transmission of toxoplasmosis: Risk estimates for clinical counselling. *The Lancet* **353**, 1829–1833.
- [21] Hens N, Shkedy Z, Aerts M, Faes C, van Damme P and Beutels P (2012) *Modeling Infectious Disease Parameters Based on Serological and Social Contact Data: A Modern Statistical Perspective*. New York, NY: Springer Science & Business Media.
- [22] R. Core Team (2022) *R: A Language and Environment for Statistical Computing*. Vienna, Austria: R Foundation for Statistical Computing.
- [23] Guigue N, Léon L, Hamane S, Gits-Muselli M, le Strat Y, Alanio A and Bretagne S (2018) Continuous decline of toxoplasma gondii seroprevalence in hospital: A 1997–2014 longitudinal study in Paris, France. *Frontiers in Microbiology* **9**, 2369.
- [24] Gargaté MJ, Ferreira I, Vilares A, Martins S, Cardoso C, Silva S, Nunes B and Gomes JP (2016) Toxoplasma gondii seroprevalence in the Portuguese population: Comparison of three cross-sectional studies spanning three decades. *BMJ Open* **6**, e011648.
- [25] Robert-Gagneux F (2014) It is not only the cat that did it: How to prevent and treat congenital toxoplasmosis. *Journal of Infection* **68**, S125–S133.
- [26] Opsteegh M, Kortbeek TM, Havelaar AH and van der Giessen JWB (2015) Intervention strategies to reduce human Toxoplasma gondii disease burden. *Clinical Infectious Diseases* **60**, 101–107.
- [27] Wei H-X, He C, Yang P-L, Lindsay DS and Peng H-J (2016) Relationship between cat contact and infection by Toxoplasma gondii in humans: A meta-analysis. *Comparative Parasitology* **83**, 11–19.
- [28] The European Pet Food Industry Federation (FEDIAF) (2020) Facts & Figures, European Overview; 2020 [cited on 2022 March 7]. Available at <https://www.fediaf.org/who-we-are/european-statistics.html>
- [29] Nederlandse Voedingsindustrie Gezelschapsdieren (2018) Ondernemen in het digitale tijdperk, Pet Monitor; 2018 [cited on 2022 March 7]. Available at <https://www.nvg-diervoeding.nl/assets/files/pet-monitor-2018-1.pdf>
- [30] Elmore SA, Jones JL, Conrad PA, Patton S, Lindsay DS and Dubey JP (2010) Toxoplasma gondii: Epidemiology, feline clinical aspects, and prevention. *Trends in Parasitology* **26**, 190–196.
- [31] Ahmed F, Cappai MG, Morrone S, Cavallo L, Berlinguer F, Dessi G, Tamponi C, Scala A and Varcasia A (2021) Raw meat based diet (RMBD) for household pets as potential door opener to parasitic load of domestic and urban environment. Revival of understated zoonotic hazards? A review. *One Health* **13**, 100327.
- [32] Dabritz H and Conrad PA (2010) Cats and toxoplasma: Implications for public health. *Zoonoses and Public Health* **57**, 34–52.
- [33] Ducrocq J, Simon A, Lemire M, de Serres G and Lévesque B (2021) Exposure to toxoplasma gondii through consumption of raw or undercooked meat: A systematic review and meta-analysis. *Vector-Borne and Zoonotic Diseases* **21**, 40–49.
- [34] Almeria S and Dubey J (2021) Foodborne transmission of toxoplasma gondii infection in the last decade. An overview. *Research in Veterinary Science* **135**, 371–385.
- [35] Belluco S, Simonato G, Mancin M, Pietrobelli M and Ricci A (2018) Toxoplasma gondii infection and food consumption: A systematic review and meta-analysis of case-controlled studies. *Critical Reviews in Food Science and Nutrition* **58**, 3085–3096.
- [36] Markon AO, Ryan KA, Wadhawan A, Pavlovich M, Groer MW, Punzalan C, Gensheimer K, Jones JL, Daue ML, Dagdag A, Donnelly P, Peng X, Pollin TI, Mitchell BD and Postolache TT (2021) Risk factors for Toxoplasma gondii seropositivity in the Old Order Amish. *Epidemiology & Infection* **149**, e89.
- [37] Jones JL, Dargelas V, Roberts J, Press C, Remington JS and Montoya JG (2009) Risk factors for toxoplasma gondii infection in the United States. *Clinical Infectious Diseases* **49**, 878–884.
- [38] Deng H, Swart A, Bonačić Marinović AA, van der Giessen JWB and Opsteegh M (2020) The effect of salting on Toxoplasma gondii viability evaluated and implemented in a quantitative risk assessment of meat-borne human infection. *International Journal of Food Microbiology* **314**, 108380.
- [39] Gollub EL, Leroy V, Gilbert R, Chêne G, Wallon M and European Toxoprevention Study Group (EUROTOXO) (2008) Effectiveness of health education on Toxoplasma-related knowledge, behaviour, and risk of seroconversion in pregnancy. *European Journal of Obstetrics & Gynecology and Reproductive Biology* **136**, 137–145.
- [40] Ferreira IMR, Vidal JE, de Mattos CdCB, de Mattos LC, Qu D, Su C and Pereira-Chioccola VL (2011) Toxoplasma gondii isolates: Multilocus RFLP–PCR genotyping from human patients in Sao Paulo state, Brazil identified distinct genotypes. *Experimental Parasitology* **129**, 190–195.
- [41] Postema AE, Ferreira JA, van der Klis FR, de Melker H and Mollema L (2022) Investigating sources of non-response bias in a population-based seroprevalence study of vaccine-preventable diseases in The Netherlands.