Population-based surveillance study of *Campylobacter* **infections in Finland**

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(Accepted 12 February 2010; first published online 22 March 2010)

SUMMARY

The annual incidence in 14361 campylobacteriosis cases reported in Finland in 2002–2005 varied between 61 and 76/100 000 population. The mean incidence was highest (148/100 000) in the 25–29 years age group and lowest (range 21–24/100 000) in children aged 5–14 years and patients aged ≥ 75 years. The number of domestic cases was low in winter and peaked in summer. A total of 622 strains isolated from domestic infections and 785 foreign travel-related strains were serotyped. Serotypes Pen 3 and Pen 37 had the strongest association with travel-related infections (96%, P < 0.001), and Pen 6,7, Pen 12 and Pen 27 were significantly associated with domestic infections (>70% domestic within each serotype, P < 0.001). Pen 2 and Pen 1,44 were less common in older than in younger patients. Of domestic strains, a higher proportion of Pen 2 strains was isolated in winter (18%) compared to the other serotypes (0–10%).

Key words: Campylobacter jejuni, epidemiology, serotype.

INTRODUCTION

Campylobacter spp. are the most common bacterial causes of gastroenteritis in developed countries. In Finland, around 3500–4000 *Campylobacter* cases have been reported in recent years (http://www3.ktl.fi). Most of the cases are sporadic, which complicates the identification of infection sources. The incidence of *Campylobacter* infections is highest

in the summer months showing a peak in July [1, 2]. Finnish clinical microbiology laboratories are obliged to report their *Campylobacter* findings to the National Infectious Disease Register (NIDR) maintained at the Department of Infectious Disease Surveillance and Control, National Institute for Health and Welfare (THL) (formerly National Public Health Institute, KTL). Patients were asked about their travel history since 2004. In that year, information was obtained from 61% of cases. Of those, 68% had been abroad just prior to becoming ill [2]. However, the actual proportions of domestic and imported infections have not been reliably determined because of the high percentage of data without known travel history.

In this study, *Campylobacter* strains isolated from patients were collected for epidemiological typing. Patients were asked about their travel histories to obtain reliable information regarding the foreign or

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Parts of the data on serotypes and seasonal distribution of infections were presented at the 12th International Workshop on *Campylobacter*, *Helicobacter* and Related Organisms. Aarhus, Denmark, 6–10 September 2003, and at the 6th Valamo Conference on Environmental Health and Risk Assessment: Food Safety. Valamo Monastery, Heinävesi, Finland, 10–12 May 2004.

domestic origin of the *Campylobacter* infections. The overall incidence, and demographical and seasonal distribution of the infections in Finland were also investigated. In addition, serotypes of domestic *C. jejuni* strains were analysed to find features characteristic of the Finnish *Campylobacter* strains. This knowledge will help in directing epidemiological investigations and focusing on control measures for *Campylobacter* infections.

METHODS

Reporting findings

Between 1 January 2002 and 31 December 2005, 14361 *Campylobacter* infections were reported to NIDR. The NIDR provided data on the overall incidence, seasonal changes in the number of *Campylobacter* infections and demographical distribution of the cases diagnosed in Finland.

Strains studied

Campylobacter strains (n=2364, 16%) of cases reported to NIDR in 2002–2005) were collected during a 3-year period (from 1 July 2002 to 31 June 2005) and represented all Campylobacter strains found in nine Finnish clinical microbiology laboratories of nine hospital districts. The laboratories enquired about the patient's travel history, and this together with the date when the specimen was taken were recorded on a special form accompanying the patient's strain submitted to Enteric Bacteria Laboratory (EBL) of KTL (presently the Bacteriology Unit of THL) for further testing. The strain was regarded as being associated with foreign travel if the patient had travelled abroad and the onset of symptoms was within 10 days or the specimen was taken within 17 days after the patient's return. Of the 2364 strains collected, 1407 C. jejuni strains (60%) were further analysed by heatstable serotyping. These included all C. jejuni isolates from domestic (n=622) and foreign travel-related (n=785) infections isolated during a 2-year period (July 2002–June 2003 and July 2004–June 2005). Only one strain per case was studied.

Identification and serotyping

Preliminary species identification was carried out at the clinical laboratories by hippurate hydrolysis test. Hippurate-positive strains were identified as *C. jejuni*. The species identification of hippuratenegative strains was carried out in EBL by PCR as described previously [3]. The strains were subcultured twice on blood agar and grown for 48 h at 42 °C in a microaerobic atmosphere. Serotyping was performed according to the Penner serotyping scheme [4] using a commercially available serotyping kit (Denka-Seiken Co. Ltd, Japan), which contains 25 absorbed antisera against heat-stable serotypes (1,44), 2, 3, (4,13,16,43,50), 5, (6,7), 8, 10, 11, 12, 15, 18, 19, 21, (23,36,52), 27, 31, 32, 37, 38, 41, 45, 52, 55, and 57. Non-typable strains were designated NT.

Statistical methods

Yates-corrected χ^2 and Fisher's exact one-tailed tests (Epi-InfoTM 3.3, Centers for Disease Control and Prevention, USA) were used to compare the proportions of serotypes in domestic infections with those in travel-associated infections. A *P* value of <0.05 indicated statistical significance.

RESULTS

Seasonal variation and incidence of all reported infections

In total, 14361 Campylobacter cases were reported during 2002-2005. The seasonal variation in the number of cases was similar each year: low in winter, and peaking in July and August (Fig. 1). The majority of the 2364 cases for which the travel history was known, were of domestic origin during the seasonal peak, only 28-43% were associated with foreign travel, compared to 65-93% in winter (December-February, Fig. 1). The overall annual incidence varied from 61/100000 in 2003 to 76/100000 in 2005. The mean incidence over the 4-year period was highest, 148/100000, in the 25-29 years age group (Fig. 2). The lowest incidences were detected in children aged 5-9 (21/100 000), and 10-14 (23/100 000) years, and in patients aged ≥ 75 years (24/100000). In children aged <5 years, the incidence was higher (33/100000) than in older children. The most marked change in the incidence was the increase from $49/100\,000$ in the 15-19 years age group to 112/100000 in the 20-24 years age group. From 0 to 14 years, the number of domestically acquired infections decreased, whereas the number of foreign travel-related infections increased (Fig. 2). The number of domestic infections



Fig. 1. Monthly incidence of *Campylobacter* cases reported to the National Infectious Disease Register (NIDR) between July 2002 and June 2005 (—), and the proportion of foreign travel-related cases of the strains collected in this study (\blacksquare , n=2364) during the same time period.



Fig. 2. Mean incidence of *Campylobacter* infection in different age groups of patients in Finland between 2002 and 2005 as reported to the National Infectious Disease Register (n = 14361), and travel history of the patients in nine hospital districts between July 2002 and June 2005 (n = 2364). \square , Domestic; \square , travel related; \square , travel history unknown.

remained constant in patients aged 20-59 years whereas there was more variation in the number of travel-associated infections. The proportion of foreign travel-related infections varied from 54% to 68% in patients aged 10-59 years, and from 13% to 45% in other patient groups (Fig. 2).

Data on species and serotype distributions

Of the 2364 strains collected, *C. jejuni* accounted for 92% (2184 strains), and *C. coli* for 7% (168). In addition, there were *C. upsaliensis* (n=8), *C. fetus* (n=2) and *C. lari* (n=2) strains. The data on the patients' travel history was available for 92% (2186) of the strains. Of the *C. jejuni* strains, 41% were of domestic

origin, 51% were associated with foreign travel and data was unavailable for 8%. Of the 168 *C. coli* strains, 81% were associated with foreign travel, 17% were of domestic origin and data was unavailable for 2%.

The 1407 *C. jejuni* strains that were serotyped, represented all of the 21 specific serotypes and four serogroups included in the serotyping kit. In addition, 15 strains reacted with several antisera (mixed serotypes) and 593 strains (42%) were NT.

The most common serotypes were Pen 2 (14%), Pen 4-complex (8%), Pen 12 (6%) and Pen 1,44 (6%) (Table 1). Serotypes Pen 12, Pen 6,7 and Pen 27 were more common (12%, 6%, 3%, respectively) in domestic strains than in travel-associated strains

All strains ($n = 1407$) Serotype	Domestic infections $(n=622)$	Travel-associated infections $(n=785)$	<i>P</i> value	Proportion of travel-associated infections within each serotype
Pen 2 $(n = 192, 14\%)$	80 (13%)	112 (14%)	n.s.	58 %
Pen 4, 13, 16, 43, 50 (n=118, 8%)	56 (9%)	62 (8%)	n.s.	53 %
Pen 12 $(n = 88, 6\%)$	72 (12%)	16 (2%)	< 0.001	18 %
Pen 1,44 $(n = 79, 6\%)$	25 (4%)	54 (7%)	< 0.02	68 %
Pen 6,7 $(n = 50, 4\%)$	37 (6%)	13 (2%)	< 0.001	26 %
Pen 3 $(n = 46, 3\%)$	2 (<1%)	44 (6%)	< 0.001	96 %
Pen 37 $(n=28, 2\%)$	1 (<1%)	27 (3%)	< 0.001	96 %
Pen 27 $(n=25, 2\%)$	21 (3%)	4 (1%)	< 0.001	16%
Other serotypes $(n=188, <2\% \text{ each})$	84 (14%)	104 (13%)	n.d.	55%
Not typable $(n = 593, 42\%)$	244 (39%)	349 (44%)	n.s.	59 %

Table 1. Serotypes of the strains (n = 1407) isolated from domestic (n = 622) and foreign travel-associated (n = 785) C. jejuni infections, and comparison of proportional data by χ^2 test

n.s., Not significant; n.d., not determined.

(2%, 2%, 1%, P < 0.001 for each). In contrast, majority of the isolates within serotypes Pen 1,44 (4% vs. 7%, P < 0.05), Pen 3 (<1% vs. 6%, P < 0.001) and Pen 37 (<1% vs. 3%; P < 0.001) were from patients with a history of foreign travel (Table 1).

Of the domestic C. jejuni strains, 71 % were found in summer (June–August, n = 442), 19% in autumn (September–November, n=121), 6% in winter (December–February, n=38), and 3% in spring (March–May, n = 21). The distributions of the specific serotypes in the domestic strains followed the same seasonal trend showing a peak in summer and low incidences in other seasons, except for Pen 2. A higher proportion of Pen 2 strains was isolated in winter (18%) compared to the other serotypes (0-10%). Of the travel-associated strains, 33% (259 strains) were isolated in summer, 25% (199) in autumn, 22% (171) in winter and 20% (156) in spring. Pen 1,44 and Pen 6,7 showed a peak in summer (44% and 62%), respectively), and the majority of the Pen 12 strains were isolated in summer (44%) and autumn (50%). The other serotypes in travel-associated strains were evenly distributed seasonally.

To study the demographical distribution of the serotypes, patients were divided into four age groups: 0–19 (83/88 isolates from domestic/imported infections), 20–39 (176/348), 40–59 (212/282) and ≥ 60 (151/62) years. In domestic infections, the strains of serotype Pen 12 were frequent (11–13%) in all age groups. Further, Pen 4-complex, Pen 6,7 and Pen 27 were evenly distributed (8–10%, 4–9% and 1–4%,

respectively). Pen 2 was the most common serotype in patients aged 0–59 years (13–18%) but accounted for only 7% of the isolates from older patients. Pen 1,44 was more common (7%) in the 0–19 years age group than in older patients (3–4%). Of the travel-related strains, the proportions of Pen 2 and Pen 1,44 were lower (6% and 3%, respectively) in patients ≥ 60 years than in younger patients (13–16% and 6–7%), whereas the proportion of Pen 4-complex was higher (16%) than in younger age groups (6–9%).

The nine clinical microbiology laboratories were grouped in three geographical regions, Southern (179/326 isolates from domestic/imported infections), Eastern (141/97) and Western (302/362) Finland, each with three laboratories to study the geographical distribution of the serotypes. In Southern Finland, the most common serotypes in domestic infections were Pen 12 (15%), Pen 2 (9%), Pen 1,44 (7%), and Pen 4-complex (6%); in Eastern Finland Pen 12 (12%), Pen 4-complex (12%), and Pen 2, Pen 6,7 and Pen 27 (6% of each) and in Western Finland Pen 2 (18%), Pen 12 (10%), Pen 4-complex (9%), and Pen 6,7 (8%). The lower frequency of Pen 2 in patients aged ≥ 60 years was observed in Southern and Western Finland (5% and 9%, respectively) compared to younger patients (8-13% and 20-23%). In Eastern Finland, the frequency of Pen 2 was 19% in the 0–19 years age group, and 4-6% in older patients. Of the travel-associated infections, there were only minor geographical differences in the distributions of the serotypes.

DISCUSSION

We derived data on 14361 human cases from NIDR, and serotyped results from a collection of 1407 *C. je-juni* strains isolated from patients with a known travel history. The aim was to investigate temporal, geographical and demographical trends of *Campylobacter* infections diagnosed in Finland in 2002–2005. In particular, we were interested in domestic cases, which were defined as not travelling abroad within 10 days prior to the onset of symptoms or 17 days prior to providing the specimen. The mean incubation period of *Campylobacter* infection is usually 2–5 days, with a range of 1–10 days [5, 6].

The mean incidence of Campylobacter infections was low in children (aged 0-14 years), but increased rapidly in the 15-29 years age group, and then decreased in older age groups. A peak in the incidence in young adults has been observed in many countries [7] and it has been proposed that it is related to increased foreign travel in this age group [8]. In our study, foreign travel-related and domestically acquired infections were analysed separately. In children aged <5years the incidence was higher than in older children although the proportion of travel-associated infections was lower. Previous studies have also shown a higher incidence in children aged < 5 years in many European countries, including Finland, USA and New Zealand [7, 9, 10]. This may be due to oversampling in this age group [8] or potential risk factors in young children [9, 11-13]. The number of both domestically acquired and travel-related cases almost doubled from the 15-19 years age group to the 20-24 years age group. Since the proportions of domestic and travel-related infections remained the same in these two age groups, increased foreign travel could only partly explain the increase in incidence. However, a higher rate of domestic infections in young adults in Finland was also reported in 1999 [10].

The annual number of domestic and travelassociated cases was about the same but the seasonal distribution was very different. The number of travelassociated cases was relatively constant throughout the year whereas domestic cases were infrequent in winter and increased considerably in summer. Our results showed that the seasonal peak observed in July and August in *Campylobacter* cases in Finland was mostly caused by domestic infections. Seasonal variations in human behaviour that may expose people to campylobacters, such as barbecue-prepared meals and attending outdoor parties, are similar to the seasonal distribution of *Campylobacter* infections [14, 15]. Moreover, the prevalence of *C. jejuni* in Finnish broiler flocks and cattle peak in July and August [16, 17]. The overall seasonal pattern of human cases and contamination of broiler flocks is similar in other Scandinavian countries as well [1, 18–20].

The most common serotypes in domestic C. jejuni strains were Pen 2, Pen 12, Pen 4-complex, Pen 6,7, Pen 1,44 and Pen 27, each with a proportion of $\ge 3\%$. In a previous study, the same serotypes were found to predominate [10]. The seasonal distribution of the infections caused by these serotypes followed the overall seasonal pattern of domestic Campylobacter infections in Finland. Of these serotypes, Pen 2, Pen 4-complex and Pen 1,44 were among the five most common types of travel-related strains, whereas Pen 12, Pen 6,7 and Pen 27 were significantly associated with domestic infections. The overall proportions of the two most common serotypes Pen 2 and Pen 12 in all domestic strains was about the same but their distribution differed. Pen 12, Pen 4-complex and Pen 6,7 were evenly distributed both geographically and across the age groups of the patients. Pen 2 was less common in patients aged ≥ 60 years. A lower mean age of Finnish patients infected with a strain of serotype Pen 2 has been reported previously [13]. It has been suggested that people may develop immunity to the most common serotypes of C. jejuni, which leads to a reduced frequency of common serotypes in older age groups [21, 22]. Another interesting feature was that the frequency of Pen 2 varied geographically between 18% in Western Finland and 6% in Eastern Finland. Moreover, Pen 2 strains were more prevalent than other serotypes in winter. For comparison, the seasonal, demographical and geographical distributions of the serotypes of strains isolated from travel-associated infections were also analysed. Of travel-associated strains, Pen 2 was evenly distributed seasonally whereas Pen 12, Pen 1,44 and Pen 6,7 peaked in summer and/or autumn. However, the differences found in the travel-associated strains may be of limited value because the destination countries varied both seasonally and depending on the age of the traveller.

The serotype distribution of the domestic human strains was quite similar but not identical to the serotypes found in Finnish cattle and broilers. In Finnish cattle, Pen 2 and Pen 4-complex are the most prevalent serotypes followed by Pen 12, while Pen 1,44 has also been reported [17]. Pen 6,7 is rarely observed in cattle [17] but has been reported as the predominant serotype in Finnish poultry [16]. Pen 12, Pen 4 complex and Pen 27 strains have also been found in poultry [16]. Serotypes Pen 3 and Pen 37, that had the strongest association with imported infections, have not been reported in Finnish foodproduction animals. Although Campylobacter infections in humans and the prevalence of Campylobacter strains in broiler flocks and cattle follow the same seasonal pattern, it is not clear how much these animals contribute to human illness. The temporal overlap in serotypes and genotypes found in Finnish patients and chicken flocks at slaughter, suggest common environmental sources for both human infection and flock contamination [23]. An association between C. jejuni isolates from cattle, chicken and humans has been shown in other countries in studies exploiting serotyping and different genotyping methods and suggests a common environmental source of infection [24–26].

In conclusion, >40% of the *C. jejuni* infections in Finland seem to be domestically acquired, and the number of domestic infections is highest in July and August. The serotype distribution of strains isolated from domestic and travel-associated infections differed. The demographic and geographic distributions of the serotypes of the domestic strains were serotype specific. Some serotypes were evenly distributed, whereas others varied both demographically and geographically. Both age-related acquired immunity and age- or area-related infection sources may play a role but the exact factors behind these differences remain to be determined.

ACKNOWLEDGEMENTS

We are grateful to the clinical microbiology laboratories for collection of isolates. The postgraduate studies of U.-M. Nakari were funded by the Finnish Graduate School on Applied Bioscience: Bioengineering, Food & Nutrition, Environment.

DECLARATION OF INTEREST

None.

REFERENCES

 Nylen G, et al. The seasonal distribution of campylobacter infection in nine European countries and New Zealand. *Epidemiology and Infection* 2002; 128: 383–390.

- Iivonen J, et al. Infectious Diseases in Finland 1995–2004. Publications of the National Public Health Institute KTL, Helsinki, Finland, 2005.
- Nakari UM, Puhakka A, Siitonen A. Correct identification and discrimination between *Campylobacter jejuni* and *C. coli* by a standardized hippurate test and speciesspecific polymerase chain reaction. *European Journal of Clinical Microbiology and Infectious Diseases* 2008; 27: 513–518.
- Penner J, Hennessy J. Passive hemagglutination technique for serotyping *Campylobacter fetus* subsp. *jejuni* on the basis of soluble heat-stable antigens. *Journal of Clinical Microbiology* 1980; 12: 732–737.
- The Blue Book. Guidelines for the control of infectious diseases 2005. Victorian Government Department of Human Services, Communicable Diseases Section, Melbourne, Victoria, Australia.
- Skirrow M, Blaser M. Clinical aspects of *Campylobacter* infection. In: Nachamkin I, Blaser M, eds. *Campylobacter*. Washington, DC: American Society for Microbiology, 2000, pp. 69–88.
- Friedman C, et al. Epidemiology of Campylobacter jejuni infections in the United States and other industrialized nations. In: Nachamkin I, Blaser M, eds. Campylobacter. Washington, DC: American Society for Microbiology, 2000, pp. 121–138.
- Kapperud G, Aasen S. Descriptive epidemiology of infections due to thermotolerant *Campylobacter* spp. in Norway, 1979–1988. *Acta Pathologica, Microbiologica et Immunologica Scandinavica* 1992; 100: 883– 890.
- Koehler K, et al. Population-based incidence of infection with selected bacterial enteric pathogens in children younger than five years of age, 1996–1998. *Pediatric Infectious Disease Journal* 2006; 25: 129–134.
- Vierikko A, et al. Domestically acquired Campylobacter infections in Finland. Emerging Infectious Diseases 2004; 10: 127–130.
- 11. **Tenkate T, Stafford R.** Risk factors for campylobacter infection in infants and young children: a matched case-control study. *Epidemiology and Infection* 2001; **127**: 399–404.
- Fullerton K, et al. Sporadic campylobacter infection in infants: a population-based surveillance case-control study. *Pediatric Infectious Disease Journal* 2007; 26: 19–24.
- Schönberg-Norio D, et al. Strain and host characteristics of Campylobacter jejuni infections in Finland. Clinical Microbiology and Infection 2006; 12: 754–760.
- Sopwith W, et al. Campylobacter jejuni multilocus sequence types in humans, northwest England, 2003– 2004. Emerging Infectious Diseases 2006; 12: 1500– 1507.
- Eberhart-Phillips J, et al. Campylobacteriosis in New Zealand: results of a case-control study. Journal of Epidemiology and Community Health 1997; 51: 686– 691.
- Perko-Mäkelä P, et al. Prevalence of campylobacters in chicken flocks during the summer of 1999 in Finland. *Epidemiology and Infection* 2002; 129: 187–192.

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- Hakkinen M, Heiska H, Hänninen M-L. Prevalence of Campylobacter spp. in cattle in Finland and antimicrobial susceptibilities of bovine Campylobacter jejuni strains. Applied and Environmental Microbiology 2007; 73: 3232–3238.
- Hansson I, et al. Surveillance programme for Campylobacter species in Swedish broilers, July 2001 to June 2002. Veterinary Record 2004; 155: 193–196.
- Bang D, et al. A one-year study of campylobacter carriage by individual Danish broiler chickens as the basis for selection of *Campylobacter* spp. strains for a chicken infection model. *Epidemiology and Infection* 2003; 130: 323–333.
- 20. Hofshagen M, Kruse H. Reduction in flock prevalence of *Campylobacter* spp. in broilers in Norway after implementation of an action plan. *Journal of Food Protection* 2005; **68**: 2220–2223.
- Miller G, et al. Does age acquired immunity confer selective protection to common serotypes of Campylobacter jejuni? BMC Infectious Diseases 2005; 5: 66.
- 22. Linneberg A, et al. IgG antibodies against microorganisms and atopic disease in Danish adults: the

Copenhagen Allergy Study. *Journal of Allergy and Clinical Immunology* 2003; **111**: 847–853.

- 23. Kärenlampi R, et al. Temporal and geographical distribution and overlap of Penner heat-stable serotypes and pulsed-field gel electrophoresis genotypes of *Campylobacter jejuni* isolates collected from humans and chickens in Finland during a seasonal peak. *Journal of Clinical Microbiology* 2003; 41: 4870–4872.
- 24. Schouls L, et al. Comparative genotyping of Campylobacter jejuni by amplified fragment length polymorphism, multilocus sequence typing, and short repeat sequencing: strain diversity, host range, and recombination. Journal of Clinical Microbiology 2003; 41: 15–26.
- Nielsen E, Engberg J, Madsen M. Distribution of serotypes of *Campylobacter jejuni* and *C. coli* from Danish patients, poultry, cattle and swine. *FEMS Immunology* and Medical Microbiology 1997; 19: 47–56.
- Nielsen E, et al. Evaluation of phenotypic and genotypic methods for subtyping *Campylobacter jejuni* isolates from humans, poultry, and cattle. *Journal of Clinical Microbiology* 2003; 8: 3800–3810.