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### Conference on 'Nutrition and healthy ageing' Symposium 1: Biology of ageing

## Biomarkers of healthy ageing: expectations and validation

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The challenge of devising a set of biomarkers capable of measuring the ageing rate in human subjects was articulated long ago. In recent years, progress in the basic biology of ageing suggests the realistic possibility of preventive or restaurative interventions that may extend healthy lifespan in mammals including human subjects. Specifically, frailty is being increasingly recognised as a clinically relevant syndrome that may be therapeutically addressed. This greatly enhances the need for sensitive and specific biomarkers of healthy ageing that are validated in both experimental animals and, importantly, in human subjects over the whole age range. Here, we will discuss the present challenges and requirements for biomarker of healthy ageing as: (i) better predictive power than chronological age for multiple dimensions of ageing; (ii) identification of the age range in which the marker is informative; (iii) establishment of sensitivity/specificity as indicators of its predictive power at the level of the individual; (iv) minimisation of methodological variation between laboratories.

Healthy ageing: Biomarkers: Human subjects: Human ageing: Biomarker validation

## Biomarkers and the stochastic nature of the ageing process

Over the last 30 years, biogerontology has moved from an observational to an interventional science with increasingly realistic potential for human interventions. This has generated an urgent need for markers that can precisely predict the biological age of populations, groups and individuals. Various approaches to define criteria for biomarkers of ageing, either in conjunction with or opposed to biomarkers of age-related disease have been published (1-4). The essential feature of a biomarker of ageing was defined by Baker and Sprott<sup>(2)</sup> as 'a biological parameter that either alone or in some multivariate composite will' ... 'better predict functional capability at some late age, than will chronological age', although the impact of age-related disease as originally excluded by Baker and Sprott is still a matter of debate<sup>(1-7)</sup>. Extensive programmes to validate marker candidates for intervention testing in mice<sup>(7)</sup> and non-human  $primates^{(5)}$  have been run, however, with limited success so  $far^{(6)}$ . This is to a large extent due to our still insufficient mechanistic understanding of the ageing process. Ageing is immensely complex. It is to a significant extent governed by chance, leading to stochastic distributions of all parameters that define the rate of ageing even in genetically identical individuals under (as much as possible) identical environmental influences<sup>(8)</sup>. While we know many of the gene products and environmental influences and their principal routes of interactions that determine the rate of ageing, the impact of any of these on the ageing process in a given individual can vary greatly due to chance events that may occur already during early development. In some cases this will be 'true' chance that is by its nature unpredictable (as described by the uncertainty principle in particle physics). In other cases, it will be randomness that arises from the sheer number of interactions each of which is essentially deterministic (and so can, in principle, be measured and assessed). Finally, experimental ignorance, not having

Abbreviations: SBP, systolic blood pressure; TL, telomere length. \*Corresponding author: T. von Zglinicki, email t.vonzglinicki@ncl.ac.uk

discovered the relevance of a gene or the existence of a pathway, is still a major cause of unexplained variance in ageing. For all these reasons, we are yet far from understanding ageing mechanistically. This is reflected by the fact that there is no definition of ageing as a process that occurs in an individual. Rather, the best available definition of biological ageing is a probabilistic one, by which ageing is identified by an ever increasing intrinsic probability of death with progressing time.

Accordingly, the perfect biomarker would allow the precise measurement of the probability of death at any given time. It is immediately clear from the above that a truly perfect biomarker of ageing cannot exist because there will always be true or apparent chance events in the future that change the ageing trajectory of an individual. In other words, biomarkers of ageing are by their nature probabilistic with a limited precision of a prediction at the level of the individual. Conversely, every improvement of biomarker prediction contributes to the reduction of apparent randomness and ignorance. Therefore, biomarkers of ageing not only have a utilitarian value, but do contribute greatly to the conceptual understanding of the ageing process.

# Ageing and disease or what is a biomarker of healthy ageing?

There has been a longstanding debate in the field whether biomarkers of ageing should (and could) specifically measure basic underlying processes independent of age-related disease (1-7). It has often been proposed that ageing, being a basic process underlying the development of disease and frailty, should be researched (and biomarkers of ageing should be validated) in disease-free subjects (for review see<sup>(9)</sup>). A biomarker of healthy ageing in this sense would indicate underlying ageing biology not modified by disease. This concept was highly relevant in the early days of biogerontology when the field struggled to prove that ageing was more than and distinct from the sum of age-related morbidities. However, it is now well established that common basic biological processes, typically triggered by molecular damage and modified by cellular and systemic responses drive ageing at the level of the organism and modify risks for multiple diseases in a tissue-, organ- and system-specific manner. In turn, disease will feed back into the underlying molecular networks and thus impact onto the rate of ageing as well as enhance the risk for additional disease. For instance, chronic inflammation, which is strongly associated with most age-related chronic diseases, including dementias, depression, atherosclerosis, cancers, and diabetes, aggravates cellular senescence, which in turn reduces tissue regenerative potential and enhances pro-inflammatory signals, potentially increasing the risk for additional diseases<sup>(10,11)</sup>. There is also good epidemiological evidence that prevalence of a chronic disease is a significant risk factor for incidence of additional, often multiple ageassociated degenerative diseases (see for instance<sup>(12)</sup>). Thus, both from an opportunistic and a conceptual point of view it seems appropriate to view 'basic' ageing and age-related multimorbidity and disability as a continuum, especially with respect to biomarker development and validation.

Therefore, a biomarker of healthy ageing should not be confounded with a parameter that is informative or discriminatory for the healthy part of the ageing population only. Rather, it is a parameter that predicts the probability for maintenance of health with increasing age with high specificity and sensitivity.

Maintenance of health in an ageing population is a relative concept; at ages 85 years and above there is essentially no one free severe disease, and multimorbidity is  $common^{(13)}$ . It is also a multidimensional concept, including not only absence of disease/multimorbidity but also cognition, capability/dependency, frailty and, ultimately, longevity. In the old, correlations between most of these dimensions (especially those to multimorbidity) are weak, suggesting that individual biomarkers will have different predictive power for multiple dimensions of the ageing process. For instance, there has been a longstanding debate as to whether and to what extent lifespan and healthspan co-vary in the old<sup>(14-16)</sup>. Multimorbidity, disability and mortality are at best loosely associated in octa- and nonagenarians<sup>(17)</sup>.

However, all these dimensions are associated with and driven by the ageing process. We therefore proposed<sup>(17)</sup> that an informative biomarker of (healthy) ageing should be predictive for several of these dimensions. To better capture the multidimensionality of the ageing process, additional endpoint measures, prominently including measures of psychological and mental well-being, need to be considered for biomarker validation, as these deteriorate in significant subgroups of the population with important consequences for physiology and perception of the ageing process.

The concept of frailty requires special consideration in the context of ageing biomarker validation. Frailty is characterised by increased vulnerability to stress resulting in an increased risk of adverse health outcomes including disability, hospitalisation, institutionalisation and death. There is as yet no universally accepted definition of frailty. The two leading concepts are frailty as a clinical syndrome; a cluster of specific symptoms and signs including weight loss, exhaustion, low physical activity, muscle weakness and slow walking speed as developed by Fried et al.<sup>(18)</sup> or as a cumulative index of health deficits and indicator of biological age as proposed by Rockwood *et al.*<sup>(19,20)</sup>. These individual deficits can</sup> include diseases, symptoms, signs, function tests and laboratory tests. Provided enough deficits are included in the index, their exact nature seems  $unimportant^{(21)}$ . Thus, frailty can be regarded as a complex biomarker of ageing (the Rockwood model) or as a clinical definition of an ageing syndrome (the Fried model), clearly illustrating the ambivalence between endpoints and biomarkers in ageing research. Application of both models to the same population shows that they measure overlapping but not identical concepts with significant fractions of participants falling in one but not the other frailty category. Interestingly, on a cohort level, associations 424

to a large number of biomarker candidates, especially inflammation markers, were very similar for both frailty models<sup>(22)</sup>.

#### Use of birth cohorts for biomarker validation

Chronological age is the most universally available 'biomarker' of ageing (forensics being a notable exception). However, it is also a weak marker; the differences in survival between the longest- and the shortest-living member of a cohort are typically greater than mean or median lifespan of the cohort, even in genetically and environmentally homogeneous cohorts under protected conditions with very little impact of external causes of death. Therefore, the first requirement for a candidate biomarker of ageing is that it needs to have better predictive power than chronologic age. Many population-based studies include participants over a wide age range, and associations are adjusted for age. This might not always be a robust procedure, given that biomarker candidates and their predictive power are often non-linearly associated with chronologic age (see later). This problem is circumvented if associations between marker candidates and ageing 'outcomes' are analysed in birth cohorts, in which all cohort members fall within a narrow age range. This approach addresses directly the relevant question, namely: are (groups of) individuals that are of the same chronologic age different in their 'biological age' and if so, by how much?

The limitation of the birth cohort approach is that, even if the study group was representative for the whole population at that age, it answers the question only for a narrow age group. There is now ample evidence that the predictive power of multiple candidate biomarkers of ageing varies with age group. For instance, up to about age 70 years systolic blood pressure (SBP) increases continuously with age<sup>(23)</sup> and high SBP is a well-recognised risk factor for CVD and associated mortality<sup>(24–26)</sup>. At higher ages, however, SBP decreases rather than increases with  $age^{(23)}$  and higher blood pressure becomes protective in terms of all-cause mortality and cognition, whereas low SBP confers increased risks for mortality, cognitive impairment and disability<sup>(17,27–31)</sup>. Similarly, short peripheral blood telomere length (TL) is recognised as a risk factor for both mortality and (multi-) morbidity<sup>(32-37)</sup>. These associations are strongest in the age group up to about 75 years but tended to disappear in older populations<sup>(34,38)</sup>. Similar decreases of predictive power at higher age have been noted for other potential biomarkers<sup>(39,40)</sup>, although often there are not sufficient data on multiple age cohorts, especially the oldest old, available. Cohort and/or period effects may be partially responsible if there is a trend reversal or loss of predictive power at old age. Typically, later born cohorts are physically and cognitively healthier and show extended life expectancy as compared to earlier born cohorts at the same chronological age<sup>(26)</sup>. Increasingly widespread use of certain medication in the older population will influence biomarker associations. For instance, we found an association between high levels of vitamin D and cognitive

impairment in a population-based study of 85-year olds, which is most probably explained by vitamin D supplementation specifically in care homes<sup>(17)</sup>. An increased use of anti-hypertensive medication in this age group may partially explain the decrease of SBP. However, the general pattern remained even for participants not on antihypertensive medication and after adjustment for survivor bias<sup>(23)</sup>. Moreover, low SBP predicted increased mortality in 90-year olds without heart failure, defined by low levels of N-terminal prohormone of brain natriuretic peptide<sup>(30,41)</sup>.

Life history cohorts are the ideal test bed to establish the age dependency of candidate biomarkers of ageing. Life history cohorts are birth cohorts for which candidate marker information is available longitudinally over a large fraction of the complete life history and which have reached a sufficient age to be informative about age-related outcomes. There are at least 60-70 human ageing cohorts that have been studied longitudinally worldwide<sup>(42,43)</sup>; however, very few of these qualify as life history cohorts. Examples of the latter from the UK include the MRC National Survey of Health and Development<sup>(44)</sup> and the Lothian Birth Cohorts of 1921 and 1936<sup>(45,46)</sup>; see also www.halcyon.ac.uk. Some candidate biomarkers of physical capability (grip strength), cardiovascular function (SBP) and cognition have in fact been longitudinally assessed for long periods of time, with follow-ups spanning in some cases over 50 years in the same participants, enabling comprehensive validation of their predictive power over the life course<sup>(23,47–56)</sup>. However, for the vast majority of biomarker candidates, life-course longitudinal data are not available and will not be for a long while, if at all. For instance, TL as a biomarker of ageing was only introduced in  $2000^{(33)}$ . In this case, the best validation strategy follows the biomarker criteria derived by Nakamura *et al.*<sup>(3,5)</sup> by combining longitudinal analyses</sup>in multiple birth cohorts, in which the longitudinal change with age is expected to be consistent with the cross-sectional differences between the cohorts. However, in human subjects life history is strongly dependent on year of birth, and life expectation increases with time<sup>(57)</sup>. In addition, if biomarker data from multiple cohorts are pooled, technical variation between different laboratories becomes a concern. So far, longitudinal biomarker studies have seldom if ever been done in multiple cohorts performed by a single laboratory with no variation in methodology. For instance, we measured peripheral blood cell TL in about 7000 participants of six UK cohorts with consistent methodology (C Martin-Ruiz, T von Zglinicki and HALCYON Study team, unpublished results). However, technical variation in blood sampling and DNA extraction could not be avoided, and the observed cohort-specific differences could thus not definitely be attributed to variation of average biological age between cohorts. It should be noted that Nakamura's criteria<sup>(3)</sup> also request the rates of age-related change of a potential biomarker to be proportional to differences in the ageing rate or lifespan among the related species. It might be concluded that most of ageing marker candidates in present use have not been sufficiently validated.

# How to validate a candidate biomarker of healthy ageing?

As discussed earlier, the main problem arising from the complex and stochastic nature of the ageing process is that there is no good single process or parameter to test biomarker prediction against. In other words, there is no gold standard. In most animal and a large number of human ageing biomarker studies, survival or lifespan is used as the closest approximation to an estimate of ageing rates. While this is in keeping with the definition of biological ageing, it has two major disadvantages: it rapidly loses power in small cohorts, and as discussed earlier, its association to 'health span' is uncertain. We believe that four steps are necessary for validation of a candidate biomarker of healthy ageing: (i) Prove better predictive power than chronological age for multiple dimensions of ageing; (ii) Identify the informative age range; (iii) Establish sensitivity/specificity; (iv) Assess (and minimise) methodological variation between laboratories.

We discussed earlier the essentiality to cover the multidimensionality of the ageing process, the advantages of birth cohorts for biomarker candidate validation and the problems associated with use of multiple birth cohorts to establish the informative age range. However, the two latter points deserve some further comments.

So far, the majority of studies in the ageing biomarker field do not go beyond establishing correlations at the cohort level. However, even a highly significant correlation between a biomarker and an outcome in a large cohort does not necessarily imply that the biomarker in question will predict the outcome with any degree of certainty for an individual. To give an example, we tested the ability of measuring TL in stroke survivors immediately after the stroke to predict incidence of dementia and 2-year survival<sup>(58)</sup>. In a linear model, every 1000 bp of TL resulted in a decreased hazard ratio (HR) for incidence of dementia (HR=0.19, 95% CI 0.19, 0.54, P=0.002). However, performing a receiver operator curve analysis, it was found that despite these strong associations a telomere setting that would result in the correct prediction for 80% of those that developed dementia would also predict 44% false positives. This resulted in a total of only 59% correct predictions (as compared to 50% by pure chance) because only a minority of patients did develop dementia $^{(58)}$ . Both the advance of preventive and restorative interventions into ageing processes (e.g. frailty) and the increasing commercialisation of biomarker services (e.g. telomeres) make it essential to generate this type of information as part of the validation process of biomarkers of ageing.

It is generally assumed that the relation between a biomarker and the outcome it stands for can be described by a linear function (possibly after some simple mathematical transformation of the marker values). Due to the complexity of the ageing process and the uncertain associations between its multiple domains discussed earlier, this will often not be true for candidate biomarkers of ageing. This is illustrated in Fig. 1, showing the



**Fig. 1.** Distribution of telomere length in a population-based birth cohort sample of 2660 participants aged 53 years. The box plot on top shows upper and lower percentiles, quartiles and median. Assuming a linear relationship between telomere length and age as estimated from multiple birth cohorts (see text) estimates of 'biological age' have been calculated for the upper and lower percentiles and the median.

distribution of TL in peripheral blood in a populationbased birth cohort sample of 2660 participants aged 53 years. From a total of eleven cohorts comprising 9929 participants and spanning an age range from 50 to 88 years, we obtained a linear regression between age and TL with very narrow confidence intervals as

$$TL = 8359 \text{ (sem 77)} - 58 \text{ (sem 1)} \times Age$$

Assuming that the same equation would also describe the distribution of 'biological age' according to TL within the 53-year-old cohort, we would find a 'biological age' for the participants with a TL representing the median of reasonable 48 years. For the upper and lower quartiles of the distribution 'biological ages' of 28.6 and 70.2 years, respectively, would be calculated. However, the upper and lower deciles of the telomere distribution, still representing 266 participants each, would end up with calculated 'biological ages' of less than 4 or more than 85.5 years, which is clearly unrealistic. Thus, the association between the biomarker TL and biological age is definitely different from the one between average TL and chronological age, most probably nonlinear and possibly not even a continuous function (e.g. only a small part of the biomarker variation between individuals may be informative for any given domain of ageing).

Finally, the use of any potential biomarker is severely restricted as long as methodological variation between laboratories has not been independently assessed and NS Proceedings of the Nutrition Society

minimised. Again using peripheral blood TL as an example for many candidate biomarkers of ageing, it is clear that at present no general applicable reference ranges for a 'normal' TL at any given age can be defined because data differ so widely between laboratories. This prohibits direct pooling of data from different laboratories to increase power as necessary for genotype-phenotype association studies. It also implies that measurements of this biomarker in individuals are useless as long as they cannot compared to a reference set established in the same laboratory.

#### Some recent biomarker examples

Recent progress in high-throughput, '-omics' technologies has enabled unbiased searches for novel candidates for biomarkers of healthy ageing to start. However, despite the clear heritability of longevity, genome-wide association studies have so far largely failed to deliver novel marker candidates<sup>(59)</sup>, probably both due to the very complex genomics involved in the ageing process and to the problems of defining the phenotype of healthy ageing clearly. A number of interesting associations with metabolomic and lipidomic parameters have been found<sup>(60–64)</sup>; however, in general these possible biomarker candidates or combinations thereof still await validation.

A number of potential biomarker candidates have been suggested by recent developments in the cell and molecular biology of ageing and, especially, cell senescence. Recent data show cell senescence as an important driver of ageing in mammals<sup>(65,66)</sup>. Peripheral blood TL was the first senescence marker to be suggested as a biomarker of human ageing<sup>(33)</sup>. This suggestion has been confirmed in a large number of independent studies but the marker suffers from low specificity/sensitivity and large methodological variation between laboratories as discussed earlier. High levels of p16 (CDKN2A) are also an indicator of cell senescence. p16 expression was first suggested as a biomarker of ageing in mice in  $2004^{(67)}$  and first applied to human subjects in  $2006^{(68)}$ . Its informative age range and whether it is actually better than chronological age still needs to be established. Persistent DNA damage may trigger either apoptosis or cell senescence, both of which may be associated with ageing. DNA damage also induces the phosphorylation of the histone variant H2AX (then called  $\gamma$ H2AX), which forms foci at sites of DNA damage, especially double-strand breaks<sup>(69)</sup>. yH2AX has recently been put forward as a potential candidate biomarker of ageing with clinical potential  $(^{70-72})$ . The capacity to repair DNA double-strand breaks is impaired with age $ing^{(73-75)}$ ,  $\gamma$ H2AX foci have been used for detection and clinical assessment of tumours in human subjects (for review see<sup>(76)</sup>), they increase with age<sup>(77)</sup> and they have been used as a marker for morbidity and age-related diseases<sup>(77)</sup>; however, from an epidemiological point of view this marker candidate is yet far from being validated as most of the studies are small and the methodologies applied are not completely compatible<sup>(70,71)</sup>. Cell senescence is now known to be mechanistically integrated with inflammation<sup>(66)</sup> and with oxidative stress<sup>(78)</sup>. However, so far a few markers of molecular oxidative damage or inflammation appear useful and consistent as biomarkers of ageing in human cohorts<sup>(71,79)</sup>.

A candidate biomarker of ageing should only be regarded as fully validated if it fulfils the requirements stated earlier. These are very stringent criteria that are only just met by some of the longest established biomarkers of ageing, hand grip strength being a notable example<sup>(80–86)</sup>. This test has strong potential as a screening tool because of its simplicity and its robust association with disability, mortality, and health care indicators such as longer hospitalisation or risk of post-surgery complications<sup>(85)</sup>. In healthy adults, lower hand grip associates with all-cause mortality and increased risk of disability in later life over a wide age range<sup>(80-84)</sup>. However, the association between mortality and hand grip strength becomes weaker in cohorts aged 60 years and over and, as evident in studies with long-term follow up, might be modified by cohort  $effects^{(86,87)}$ . Other tests of physical performance such as 'Functional reach', 'Timed Up and Go', and 'One-leg stance', also predict frailty, disability<sup>(88-91)</sup> and mortality in the short term on those over  $65^{(90)}$  but appear to lose power in the very old<sup>(92)</sup>. Even for such relatively simple assessments of physical performance, comparability between studies is limited by methodological concerns<sup>(93,94)</sup>. In any case, the predictive power of these and other 'classical' biomarkers of ageing is low and their sensitivity/specificity does not reach the limits required for a biomarker with diagnostic power, fuelling the ongoing need for novel marker candidates.

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### **Conflicts of Interest**

None.

#### Authorship

T. von Z. and C. M. M. designed and wrote the paper.

#### References

- 1. Reff ME & Schneider EL (1982) *Biological Markers of Aging*. Washington, DC: US Department of Health and Human Sciences.
- Baker GT III & Sprott RL (1988) Biomarkers of aging. Exp Gerontol 23, 223–239.
- 3. Nakamura E, Lane MA, Roth GS *et al.* (1994) Evaluating measures of hematology and blood chemistry in male rhesus monkeys as biomarkers of aging. *Exp Gerontol* **29**, 151–177.
- Butler RN, Sprott R, Warner H et al. (2004) Biomarkers of aging: from primitive organisms to humans. J Gerontol A Biol Sci Med Sci 59, B560–B567.
- 5. Ingram DK, Nakamura E, Smucny D *et al.* (2001) Strategy for identifying biomarkers of aging in long-lived species. *Exp Gerontol* **36**, 1025–1034.
- 6. Sprott RL (2010) Biomarkers of aging and disease: introduction and definitions. *Exp Gerontol* **45**, 2–4.
- Warner HR (2004) Current status of efforts to measure and modulate the biological rate of aging. J Gerontol A Biol Sci Med Sci 59, 692–696.
- 8. Finch CE & Kirkwood TBL (2000) *Chance, Development, and Aging.* New York, USA: Oxford University Press.
- 9. Holloszy JO (2000) The biology of aging. *Mayo Clin Proc* 75, Suppl., S3–S8.
- Salminen A, Kauppinen A & Kaarniranta K (2012) Emerging role of NF-κB signaling in the induction of senescence-associated secretory phenotype (SASP). *Cell* Signal 24, 835–845.
- 11. Freund A, Orjalo AV, Desprez P-Y *et al.* (2010) Inflammatory networks during cellular senescence: causes and consequences. *Trends Mol Med* **16**, 238–246.
- 12. Hillege HL, Nitsch D, Pfeffer MA *et al.* (2006) Renal function as a predictor of outcome in a broad spectrum of patients with heart failure. *Circulation* **113**, 671–678.
- Collerton J, Davies K, Jagger C *et al.* (2009) Health and disease in 85 year olds: baseline findings from the Newcastle 85+ cohort study. *BMJ* 339, b4904.
- 14. Fries JF (1980) Aging, natural death, and the compression of morbidity. *N Engl J Med* **303**, 130–135.
- 15. Crimmins EM & Beltran-Sanchez H (2011) Mortality and morbidity trends: is there compression of morbidity? *J Gerontol B Psychol Sci Soc Sci* **66**, 75–86.
- Andersen SL, Sebastiani P, Dworkis DA et al. (2012) Health span approximates life span among many supercentenarians: compression of morbidity at the approximate limit of life span. J Gerontol A Biol Sci Med Sci 67, 395–405.
- Martin-Ruiz C, Jagger C, Kingston A *et al.* (2011) Assessment of a large panel of candidate biomarkers of ageing in the Newcastle 85+ study. *Mech Ageing Dev* 132, 496–502.
- Fried LP, Tangen CM, Walston J et al. (2001) Frailty in older adults: evidence for a phenotype. J Gerontol A Biol Sci Med Sci 56, M146–M156.
- 19. Rockwood K & Mitnitski A (2007) Frailty in relation to the accumulation of deficits. *J Gerontol A Biol Sci Med Sci* 62, 722–727.
- 20. Searle SD, Mitnitski A, Gahbauer EA *et al.* (2008) A standard procedure for creating a frailty index. *BMC Geriatr* **8**, 24.
- 21. Rockwood K, Mitnitski A, Song X *et al.* (2006) Long-term risks of death and institutionalization of elderly people in relation to deficit accumulation at age 70. *J Am Geriatr Soc* **54**, 975–979.

- 22. Collerton J, Martin-Ruiz C, Davies K *et al.* (2012) Frailty and the role of inflammation, immunosenescence and cellular ageing in the very old: cross-sectional findings from the Newcastle 85+ Study. *Mech Ageing Dev* **133**, 456–466.
- 23. Wills AK, Lawlor DA, Matthews FE *et al.* (2011) Life course trajectories of systolic blood pressure using longitudinal data from eight UK cohorts. *PLoS Med* **8**, e1000440.
- Borghi C, Dormi A, L'Italien G et al. (2003) The relationship between systolic blood pressure and cardiovascular risk results of the Brisighella Heart Study. J Clin Hypertens (Greenwich) 5, 47–52.
- 25. Paultre F & Mosca L (2006) The relation of blood pressure to coronary heart mortality in different age groups varies by ethnicity. *Am J Hypertens* **19**, 179–183.
- Kannel WB, Gordon T & Schwartz MJ (1971) Systolic versus diastolic blood pressure and risk of coronary heart disease: the Framingham study. *Am J Cardiol* 27, 335–346.
- 27. Euser SM, van Bemmel T, Schram MT *et al.* (2009) The effect of age on the association between blood pressure and cognitive function later in life. *J Am Geriatr Soc* **57**, 1232–1237.
- 28. Sabayan B, van Vliet P, de Ruijter W *et al.* (2013) High blood pressure, physical and cognitive function, and risk of stroke in the oldest old: the Leiden 85-plus study. *Stroke* 44, 15–20.
- 29. Sabayan B, Oleksik AM, Maier AB *et al.* (2012) High blood pressure and resilience to physical and cognitive decline in the oldest old: the Leiden 85-plus Study. *J Am Geriatr Soc* **60**, 2014–2019.
- Poortvliet RK, de Ruijter W, de Craen AJ et al. (2013) Blood pressure trends and mortality: the Leiden 85-plus Study. J Hypertens 31, 63–70.
- 31. Blom JW, de Ruijter W, Witteman JC *et al.* (2013) Changing prediction of mortality by systolic blood pressure with increasing age: the Rotterdam study. *Age (Dordr)* **35**, 431–438.
- von Zglinicki T & Martin-Ruiz CM (2005) Telomeres as biomarkers for ageing and age-related diseases. *Curr Mol Med* 5, 197–203.
- 33. von Zglinicki T, Serra V, Lorenz M et al. (2000) Short telomeres in patients with vascular dementia: an indicator of low antioxidative capacity and a possible risk factor? *Lab Invest* 80, 1739–1747.
- Cawthon RM, Smith KR, O'Brien E et al. (2003) Association between telomere length in blood and mortality in people aged 60 years or older. *Lancet* 361, 393– 395.
- 35. Hoffmann J & Spyridopoulos I (2011) Telomere length in cardiovascular disease: new challenges in measuring this marker of cardiovascular aging. *Future Cardiol* 7, 789–803.
- Mather KA, Jorm AF, Parslow RA et al. (2011) Is telomere length a biomarker of aging? A review. J Gerontol A Biol Sci Med Sci 66, 202–213.
- von Zglinicki T (2012) Will your telomeres tell your future? BMJ 344, e1727.
- Martin-Ruiz CM, Gussekloo J, van Heemst D *et al.* (2005) Telomere length in white blood cells is not associated with morbidity or mortality in the oldest old: a populationbased study. *Aging Cell* 4, 287–290.
- 39. Gerstorf D, Ram N, Hoppmann C *et al.* (2011) Cohort differences in cognitive aging and terminal decline in the Seattle Longitudinal Study. *Dev Psychol* **47**, 1026–1041.
- Nybo H, Petersen HC, Gaist D et al. (2003) Predictors of mortality in 2249 nonagenarians – the Danish 1905-Cohort Survey. J Am Geriatr Soc 51, 1365–1373.

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- 41. Poortvliet RK, Blom JW, de Craen AJ *et al.* (2013) Low blood pressure predicts increased mortality in very old age even without heart failure: the Leiden 85-plus Study. *Eur J Heart Fail* **15**, 528–533.
  - 42. Birnie K, Cooper R, Martin RM *et al.* (2011) Childhood socioeconomic position and objectively measured physical capability levels in adulthood: a systematic review and meta-analysis. *PLoS ONE* **6**, e15564.
  - 43. Seematter-Bagnoud L & Santos-Eggimann B (2006) Population-based cohorts of the 50s and over: a summary of worldwide previous and ongoing studies for research on health in ageing. *Eur J Ageing* 3, 41–59.
  - 44. Wadsworth M, Kuh D, Richards M *et al.* (2006) Cohort Profile: the 1946 National Birth Cohort (MRC National Survey of Health and Development). *Int J Epidemiol* **35**, 49–54.
  - 45. Gow AJ, Johnson W, Pattie A *et al.* (2008) Mental ability in childhood and cognitive aging. *Gerontology* **54**, 177–186.
  - 46. Deary IJ, Gow AJ, Taylor MD *et al.* (2007) The Lothian Birth Cohort 1936: a study to examine influences on cognitive ageing from age 11 to age 70 and beyond. *BMC Geriatr* 7, 28.
  - 47. Stenholm S, Harkanen T, Sainio P *et al.* (2012) Long-term changes in handgrip strength in men and women accounting the effect of right censoring due to death. *J Gerontol A Biol Sci Med Sci* **67**, 1068–1074.
  - 48. Stenholm S, Tiainen K, Rantanen T *et al.* (2012) Long-term determinants of muscle strength decline: prospective evidence from the 22-year mini-Finland follow-up survey. J Am Geriatr Soc **60**, 77–85.
  - 49. Xue QL, Beamer BA, Chaves PH *et al.* (2010) Heterogeneity in rate of decline in grip, hip, and knee strength and the risk of all-cause mortality: the Women's Health and Aging Study II. *J Am Geriatr Soc* 58, 2076–2084.
  - Hart CL, Taylor MD, Smith GD et al. (2005) Childhood IQ and all-cause mortality before and after age 65: prospective observational study linking the Scottish Mental Survey 1932 and the Midspan studies. Br J Health Psychol 10, 153–165.
  - 51. Holsinger T, Helms M & Plassman B (2007) Intelligence in early adulthood and life span up to 65 years later in male elderly twins. *Age Ageing* **36**, 286–291.
  - 52. Deary IJ, Whiteman MC, Starr JM *et al.* (2004) The impact of childhood intelligence on later life: following up the Scottish mental surveys of 1932 and 1947. *J Pers Soc Psychol* **86**, 130–147.
  - 53. Calvin CM, Deary IJ, Fenton C *et al.* (2011) Intelligence in youth and all-cause-mortality: systematic review with meta-analysis. *Int J Epidemiol* **40**, 626–644.
  - Andersen UO, Marott JL & Jensen GB (2011) Decreasing systolic blood pressure and declining mortality rates in an untreated population: results from the Copenhagen City Heart Study. *Eur J Cardiovasc Prev Rehabil* 18, 248–253.
  - 55. Arbeev KG, Ukraintseva SV, Akushevich I *et al.* (2011) Age trajectories of physiological indices in relation to healthy life course. *Mech Ageing Dev* **132**, 93–102.
  - 56. Andersen UO & Jensen GB (2010) Trends and determinant factors for population blood pressure with 25 years of follow-up: results from the Copenhagen City Heart Study. *Eur J Cardiovasc Prev Rehabil* 17, 655–659.
  - 57. Wang H, Dwyer-Lindgren L, Lofgren KT et al. (2012) Age-specific and sex-specific mortality in 187 countries, 1970–2010: a systematic analysis for the

Global Burden of Disease Study 2010. Lancet 380, 2071–2094.

- Martin-Ruiz C, Dickinson HO, Keys B *et al.* (2006) Telomere length predicts poststroke mortality, dementia, and cognitive decline. *Ann Neurol* 60, 174–180.
- Sebastiani P, Solovieff N, DeWan AT *et al.* (2012) Genetic signatures of exceptional longevity in humans. *PLoS ONE* 7, e29848.
- 60. Ruhaak LR, Koeleman CA, Uh HW *et al.* (2013) Targeted biomarker discovery by high throughput glycosylation profiling of human plasma alpha1-antitrypsin and immunoglobulin A. *PLoS ONE* **8**, e73082.
- 61. Mishur RJ & Rea SL (2012) Applications of mass spectrometry to metabolomics and metabonomics: detection of biomarkers of aging and of age-related diseases. *Mass Spectrom Rev* **31**, 70–95.
- 62. Hulbert AJ (2010) Metabolism and longevity: is there a role for membrane fatty acids? *Integr Comp Biol* **50**, 808–817.
- 63. Jove M, Naudi A, Aledo JC *et al.* (2013) Plasma long-chain free fatty acids predict mammalian longevity. *Sci Rep* **3**, 3346.
- 64. Hausman DB, Fischer JG & Johnson MA (2012) Protein, lipid, and hematological biomarkers in centenarians: definitions, interpretation and relationships with health. *Maturitas* **71**, 205–212.
- 65. Baker DJ, Wijshake T, Tchkonia T *et al.* (2011) Clearance of p16Ink4a-positive senescent cells delays ageing-associated disorders. *Nature* **479**, 232–236.
- 66. Tchkonia T, Zhu Y, van Deursen J *et al.* (2013) Cellular senescence and the senescent secretory phenotype: therapeutic opportunities. *J Clin Invest* **123**, 966–972.
- Krishnamurthy J, Torrice C, Ramsey MR et al. (2004) Ink4a/Arf expression is a biomarker of aging. J Clin Invest 114, 1299–1307.
- Ressler S, Bartkova J, Niederegger H *et al.* (2006) p16INK4A is a robust *in vivo* biomarker of cellular aging in human skin. *Aging Cell* 5, 379–389.
- 69. Rogakou EP, Boon C, Redon C *et al.* (1999) Megabase chromatin domains involved in DNA double-strand breaks *in vivo. J Cell Biol* **146**, 905–916.
- Valdiglesias V, Giunta S, Fenech M et al. (2013) gammaH2AX as a marker of DNA double strand breaks and genomic instability in human population studies. *Mutat Res* **753**, 24–40.
- 71. Jacob KD, Noren Hooten N, Trzeciak AR *et al.* (2013) Markers of oxidant stress that are clinically relevant in aging and age-related disease. *Mech Ageing Dev* **134**, 139–157.
- Mah LJ, El-Osta A & Karagiannis TC (2010) GammaH2AX as a molecular marker of aging and disease. *Epigenetics* 5, 129–136.
- 73. Mayer PJ, Lange CS, Bradley MO *et al.* (1989) Age-dependent decline in rejoining of X-ray-induced DNA double-strand breaks in normal human lymphocytes. *Mutat Res* 219, 95–100.
- 74. Singh NP, Danner DB, Tice RR *et al.* (1990) DNA damage and repair with age in individual human lymphocytes. *Mutat Res* **237**, 123–130.
- Garm C, Moreno-Villanueva M, Burkle A *et al.* (2013) Age and gender effects on DNA strand break repair in peripheral blood mononuclear cells. *Aging Cell* 12, 58–66.
- Ivashkevich A, Redon CE, Nakamura AJ *et al.* (2012) Use of the gamma-H2AX assay to monitor DNA damage and repair in translational cancer research. *Cancer Lett* 327, 123–133.

- 77. Schurman SH, Dunn CA, Greaves R *et al.* (2012) Age-related disease association of endogenous gamma-H2AX foci in mononuclear cells derived from leukapheresis. *PLoS ONE* **7**, e45728.
- 78. Passos JF, Nelson G, Wang C *et al.* (2010) Feedback between p21 and reactive oxygen production is necessary for cell senescence. *Mol Syst Biol* **6**, 347.
- 79. Wiley L, Ashok D, Martin-Ruiz CM *et al.* (2014) Reactive oxygen species production and mitochondrial dysfunction in white blood cells are not valid biomarkers of ageing in the very old. *PLos ONE* **9**, e91005.
- Rantanen T, Harris T, Leveille SG et al. (2000) Muscle strength and body mass index as long-term predictors of mortality in initially healthy men. J Gerontol A Biol Sci Med Sci 55, M168–M173.
- 81. Rantanen T, Avlund K, Suominen H *et al.* (2002) Muscle strength as a predictor of onset of ADL dependence in people aged 75 years. *Aging Clin Exp Res* **14**, 10–15.
- 82. Gale CR, Martyn CN, Cooper C et al. (2007) Grip strength, body composition, and mortality. Int J Epidemiol 36, 228–235.
- 83. Sasaki H, Kasagi F, Yamada M *et al.* (2007) Grip strength predicts cause-specific mortality in middle-aged and elderly persons. *Am J Med* **120**, 337–342.
- 84. Newman AB, Kupelian V, Visser M et al. (2006) Strength, but not muscle mass, is associated with mortality in the health, aging and body composition study cohort. J Gerontol A Biol Sci Med Sci 61, 72–77.
- 85. Bohannon RW (2008) Hand-grip dynamometry predicts future outcomes in aging adults. J Geriatr Phys Ther **31**, 3–10.

- Cooper R, Kuh D & Hardy R (2010) Objectively measured physical capability levels and mortality: systematic review and meta-analysis. *BMJ* 341, c4467.
- Shamliyan T, Talley KMC, Ramakrishnan R *et al.* (2013) Association of frailty with survival: a systematic literature review. *Ageing Res Rev* 12, 719–736.
- Wennie Huang W-N, Perera S, VanSwearingen J et al. (2010) Performance measures predict onset of activity of daily living difficulty in community-dwelling older adults. J Am Geriatr Soc 58, 844–852.
- Lin M-R, Hwang H-F, Hu M-H *et al.* (2004) Psychometric comparisons of the timed up and go, one-leg stand, functional reach, and Tinetti balance measures in communitydwelling older people. *J Am Geriatr Soc* 52, 1343–1348.
- Davis DHJ, Rockwood MRH, Mitnitski AB *et al.* (2011) Impairments in mobility and balance in relation to frailty. *Arch Gerontol Geriatr* 53, 79–83.
- Idland G, Pettersen R, Avlund K et al. (2013) Physical performance as long-term predictor of onset of activities of daily living (ADL) disability: a 9-year longitudinal study among community-dwelling older women. Arch Gerontol Geriatr 56, 501–506.
- Takata Y, Ansai T, Soh I *et al.* (2012) Physical fitness and 6.5-year mortality in an 85-year-old community-dwelling population. *Arch Gerontol Geriatr* 54, 28–33.
- Michikawa T, Nishiwaki Y, Takebayashi T *et al.* (2009) One-leg standing test for elderly populations. *J Orthop Sci* 14, 675–685.
- 94. Taekema DG, Gussekloo J, Westendorp RG *et al.* (2012) Predicting survival in oldest old people. *Am J Med* **125**, 1188–1194.e1.