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MARINE ORGANIC CARBON AND RADIOCARBON—PRESENT AND FUTURE CHALLENGES

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ABSTRACT. We discuss present and developing techniques for studying radiocarbon in marine organic carbon (C). Bulk DOC (dissolved organic C) Δ^{14} C measurements reveal information about the cycling time and sources of DOC in the ocean, yet they are time consuming and need to be streamlined. To further elucidate the cycling of DOC, various fractions have been separated from bulk DOC, through solid phase extraction of DOC, and ultrafiltration of high and low molecular weight DOC. Research using ¹⁴C of DOC and particulate organic C separated into organic fractions revealed that the acid insoluble fraction is similar in ¹⁴C signature to that of the lipid fraction. Plans for utilizing this methodology are described. Studies using compound specific radiocarbon analyses to study the origin of biomarkers in the marine environment are reviewed and plans for the future are outlined. Development of ramped pyrolysis oxidation methods are discussed and scientific questions addressed. A modified elemental analysis (EA) combustion reactor is described that allows high particulate organic C sample throughput by direct coupling with the MIniCArbonDAtingSystem.

KEYWORDS: CSRA, dissolved organic carbon, methodology, organic carbon, radiocarbon.

INTRODUCTION

Organic carbon (C) is reduced C whose molecular structure is formed on skeletons of C atoms. There are two pools of organic C in seawater, dissolved organic C (DOC) that passes a 0.2 to 1.0 micron filter, and particulate organic C (POC) that is retained on the filter (Figure 1). The pool size of DOC is 682 GtC (Hansell 2013), similar in size to the organic C present in the upper one meter of oceanic sediments (1000 GtC, Hedges et al. 2000); the POC pool is much smaller (30–40 GtC). Early studies demonstrated that bomb ¹⁴C was incorporated into subsurface and deep organisms (technically a portion of the POC pool), such as decapods and fish, along with significant amounts of pre-bomb C, on decadal timescales (Williams et al. 1970; Williams and Linick 1975).

Organic C is primarily produced during photosynthetic reduction of dissolved inorganic C (DIC) by phototrophs in the surface ocean. The concentration of DOC in seawater is 2-4% that of DIC, which makes DOC more sensitive to changes in its sources and sinks than those for DIC. The processes controlling DIC in the ocean are CO₂ exchange with the atmosphere, photosynthesis, respiration and production and dissolution of carbonate. The processes controlling DOC in seawater are more complex, including river input, microbial production of recalcitrant DOC (Jiao et al. 2010), chemoautotrophy (Ingalls et al. 2006; Hansman et al. 2009), hydrothermal processes (McCarthy et al. 2011; Lang et al. 2006; Shah Walter et al. 2018; Estes et al. 2019), enhanced refractory DOC utilization by priming



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Figure 1 Size ranges of organic matter constituents in seawater and their relationships to the operationally defined pools of DOC (gray background) and POC (white background). (After Beaupré 2019.)

(Bianchi et al. 2011; Fang et al. 2020; Shen and Benner 2018), gel formation (Verdugo et al. 2004) and dissolution/aggregation of POC (Smith et al. 1992; Druffel et al. 2016).

The introduction of ¹⁴C analyses using AMS in the 1980s made it possible to study a broad range of organic C pools in the sea that were not possible with gas counting or liquid scintillation methods due to sample size restrictions. We discuss the present and developing advances for studying marine organic C using the mother of all isotopes, ¹⁴C.

Bulk DOC ¹⁴C

Methods for measuring bulk DOC Δ^{14} C in seawater were developed in the 1960s that required collection of 400–500 L of water in Gerard barrels (Figure 2), UV oxidation in 60 L batches, and gas counting of ¹⁴C in the CO₂ (Williams et al. 1969). Improvements to the method required UV oxidation of 5 L of seawater using AMS measurements of ¹⁴C (Williams and Druffel 1987). Beaupré et al. (2007) improved the method further by using UV oxidation of samples as small as 30 mL of seawater. Quantification of CO₂ lost during breakthrough of CO₂ from liquid nitrogen cooled traps during collection improved DOC concentrations reported with isotopic values (Walker et al. 2019). However, one day is required to process one sample, standard, or blank. New methods are critically needed to increase processing of DOC samples for ¹⁴C measurement. Batch processing of bulk DOC in 100 mL reactors improves throughput of high [DOC] samples, such as freshwater and sediment pore water, to several per day (Xue et al. 2015).

Radiocarbon in DIC has been used to measure the timescale of global deep ocean circulation. Deep water is produced in the northern North Atlantic, flows south to the Southern Ocean where it flows eastward, and then northward into the Indian and Pacific oceans, taking ~1500 ¹⁴C years to complete its journey (Broecker et al. 1960; Bien et al. 1965; Stuiver et al. 1983). This result agrees with pre-bomb bulk DOC Δ^{14} C values that decrease from the deep North Atlantic (-456‰, 4900 ¹⁴C years), to the South Atlantic (-475‰, 5200 ¹⁴C years), the Southern Ocean (-500‰, 5600 ¹⁴C years), and are lowest in the North Pacific (-560‰, 6600 ¹⁴C years) (Druffel et al. 2016, 2019; Bercovici et al. 2018; Druffel and Bauer 2000), increasing in ¹⁴C age by 1700 years. This remarkable similarity between the deep



Figure 2 Peter M. Williams on the *R/V Melville* with a Gerard barrel, a 270 L steel vessel used to collect large volume seawater samples. The barrel is covered to protect it from particles coming mostly from the ship's stack. Circa 1989. (Photo by Ellen Druffel.)

cycling time of DOC and DIC suggests that DOC Δ^{14} C is controlled primarily by circulation, indicating that deep DOC is refractory on millennial timescales (Hansell 2013). Yet the DOC ¹⁴C ages are much lower (~3000 ¹⁴C years) than those of DIC.

There are significant correlations between DOC and DIC Δ^{14} C values for seawater taken from the same niskin bottle at numerous locations in the world oceans (Beaupré and Aluwihare 2010; Beaupré et al. 2020). Similar to DIC Δ^{14} C results in the central Pacific (<1,000 m) that contained bomb ¹⁴C in the 1980s (Ostlund and Stuiver 1980) and >1990s (Key et al. 2004; McNichol et al. 2022), bomb ¹⁴C is seen in the DOC Δ^{14} C values in the upper 1000 m in the Pacific Ocean (Figure 3). The highest surface values are found in the mid-ocean gyres around 20°N and 20°S (>-250‰), and lower values are near the equator and polar



Figure 3 DOC Δ^{14} C values (‰) of water samples (indicated by black dots) collected from seven stations along 150°W on the P16N cruise in 2015 (Druffel et al. 2019), one station on the P06 cruise (stn 130, 32.5°S 144.7°W) in 2010 (Druffel and Griffin 2015) and one station from the Southern Ocean (SOce; 54.0°S 176.0°W) in 1995 (Druffel and Bauer 2000) using Ocean Data View (Schlitzer 2015). (After Druffel et al. 2019.)

regions (-260‰ to -400‰) where deeper waters upwell to the surface (Bercovici et al. 2018; Druffel and Bauer 2000). Subantarctic Mode Water, produced in the southwest Pacific, flows between 200 and 400 m to the Equatorial Undercurrent in the equatorial Pacific, and has Δ^{14} C values that range from -460 to -350‰ (Figure 3). Formed between 60° and 50°S, Antarctic Intermediate Water flows north at depths of ~700–1200 m and has Δ^{14} C values that range from -530 to -410‰ (Figure 3).

Questions that remain unanswered about marine DOC include the sources and sinks of DOC from hydrothermal systems, chemoautotrophy, sediment porewaters, interaction with POC, and photooxidation of DOC in the surface ocean and aerosols (Beaupré et al. 2020). ¹⁴C and molecular identification methods (e.g., NMR and FT-ICR-MS) can help to determine the importance of these mechanisms.

SPE-DOC, and Size-Fractionated DOC

Solid-phase extraction (SPE) is a method of separating DOC (SPE-DOC) from bulk DOC using hydrophobic, silica or polystyrene resins. Seawater is acidified to enhance the relative abundance of protonated carboxyl groups, decreasing solubility and increasing sorption onto the resin (Mopper et al. 2007). XAD and C18 resins have been used to collect SPE-DOC and generally isolate humic substances from total DOC (Druffel et al. 1992; Green

and Blough 1994; Hedges 1992; Kieber et al. 1997; Peuravuori and Pihlaja 1997). C18 resins were especially effective in isolating chromophoric humic substances (Amador et al. 1990). More recently, styrene divinylbenzene polymer sorbents, such as PPL are more efficient at extracting DOC and leach less C than C18 resins. NMR analysis shows that SPE-DOC isolated using PPL is more representative of bulk DOC than that isolated using C18 resins (Dittmar et al. 2008). The ease of use, high recoveries, and ability to desalinate SPE-DOC samples have led to wide use of PPL resins in investigations of marine DOC. Subsequent methodological studies have compared PPL, C18, and other SPE resins in more detail, and show that DOC extraction can be increased up to 89% with the correct ratio of DOC/PPL (Li et al. 2016). Lewis et al. (2020) demonstrated that the optical and isotopic characteristics of SPE-DOC using PPL cartridges are heterogeneous during elution with methanol, which may be useful in further separation of compounds based on their relative polarities.

PPL has been used to isotopically and chemically characterize SPE-DOC in the east South Atlantic Ocean, the Bermuda Atlantic Time Series, Station ALOHA in the North Pacific, the Weddell Sea, and the deep North Pacific Ocean (Broek et al. 2020; Flerus et al. 2012; Hertkorn et al. 2013; Lechtenfeld et al. 2014; Zigah et al. 2017). SPE-DOC typically has lower Δ^{14} C and δ^{13} C values than those of total DOC (Broek et al. 2020; Coppola and Druffel 2016; Zigah et al. 2017; Lewis et al. 2021), is rich in aromatic C (Simjouw et al. 2005), and has elemental compositions indicative of thermogenic processes found throughout the water column (Dittmar and Koch 2006; Hertkorn et al. 2013). The presence of carboxyl-rich alicyclic molecules (CRAM), with low H/C and O/C ratios indicative of recalcitrance, is often found in SPE-DOC and tends to increase with depth in the water column (Broek et al. 2020; Hertkorn et al. 2013; Lechtenfeld et al. 2015). Lechtenfeld (2015) found that bacterial SPE-DOC isolated in laboratory experiments is similar in composition with natural marine DOC and the composition of recalcitrant molecules in the ocean. The link between SPE-DOC recalcitrance, low Δ^{14} C, and low δ^{13} C may all be indicative of the influence of marine heterotrophic bacteria on DOC degradation, transformation, cycling, and the production of refractory DOC (Flerus et al. 2012; Lechtenfeld et al. 2015; Osterholz et al. 2015). Solid-phase extraction will likely continue to advance our knowledge of DOC molecular composition as techniques to isolate and further characterize DOC develop further.

For several decades, ultrafiltration (UF) has been used as a method to isolate DOC from abundant salts in seawater. The technique has shaped much of our understanding of DOC cycling and molecular composition, and samples material that can be isolated without the chemical bias inherent to all resin chromatography methods. Early UF studies have shown that, despite low DOC recovery (10–30%), ultrafiltered DOC (UDOC), also referred to as high molecular weight (HMW DOC), was generally representative of total DOC in terms of its bulk elemental (C:N) and isotopic (δ^{13} C, δ^{15} N) values (Benner et al. 1992, 1997; McCarthy et al. 1993). However, several studies have shown that UDOC differs from bulk DOC in molecular composition (Skoog and Benner 1997; Dittmar et al. 2001) and Δ^{14} C values (Santschi et al. 1995; Loh et al. 2004; McNichol and Aluwihare 2007). Further parameterization of UDOC Δ^{14} C values as a function of UF experiment concentration factor (CF) reconciled >250‰ variability in reported surface and deep Pacific UDOC Δ^{14} C values (Figure 4a; Walker et al. 2011). Further work suggested that changes in CF could be used for the targeted isolation of semi-labile or even bioavailable UDOC (Walker et al. 2016a). Robust, size-age-composition relationships within both the DOC pool and



Figure 4 Size- Δ^{14} C and elemental composition (C:N) relationships of DOC in the Pacific Ocean. (A) DOC Δ^{14} C as a function of ultrafiltration concentration factor (CF) for the surface and deep Pacific ocean. Relationships between the C:N content of size-fractionated marine organic matter (B,D) and Δ^{14} C (C,E) in the surface and deep Pacific. (Figures adapted from Walker et al. 2011 and Walker et al. 2016b.)

total marine organic matter size continuum (Figure 4b,c,d) suggest a "precursor-product" relationship controlling the cycling (14 C age) and chemical composition (N-content) of marine organic matter. As a result, DOC C:N content and 14 C age can be generally predicted from molecular size (Walker et al. 2016b).

A recent advancement is the combination of UF and SPE techniques to view organic matter through various analytical windows simultaneously. By filtering out POC, then performing solid-phase extraction on UDOC permeate fractions, the comparison of up to three different sub-pools of DOC is possible (HMW-DOC, hydrophobic LMW-DOC, hydrophobic LMW-DOC) (Broek et al. 2017; Zigah et al. 2017; Broek et al. 2020). This method has provided new insights regarding the cycling of hydrothermal vent DOC in the open ocean, the possible addition of high molecular weight DOC to the deep ocean, and the selective utilization of some LMW-DOC during meridional overturning circulation (Broek et al. 2017; Zigah et al. 2017; Broek et al. 2020).

Compound Classes and ¹⁴C

Marine organic matter exists as molecules, some of which are characterizable. The characterizable organic C is made up mostly of amino acids/proteins, sugars/carbohydrates, lipids, and nucleic acids. However, more than half of the organic matter in seawater and sediments is not characterizable at the molecular level (Hedges et al. 2000). Whereas 85% of surface plankton and POC in the central Pacific is identified molecularly (Wakeham et al. 1997), less than 25% of deep POC is so identified, due to extensive remineralization as POC falls through the water column. The Δ^{14} C of sinking POC in the deep east North Pacific is lower than that of surface DIC that was photosynthesized to produce the organic C (Druffel et al. 1996). The surface sediment organic C has even lower Δ^{14} C values (Wang



Figure 5 Δ^{14} C of organic compound classes (THAA, open triangles; TCHO, open circles; lipids, solid triangles; acid insoluble fraction, solid circles). The range of DIC ¹⁴C values in surface waters at Stn M ranged from about 40 to 70‰. (After Hwang and Druffel 2003.)

et al. 1998). A correlation was found between Δ^{14} C and aluminum content of sinking POC, demonstrating that organic C associated with lithogenic material from sediment resuspension causes the observed low Δ^{14} C values (Hwang et al. 2010).

To understand these trends, separation of POC into four compound classes (or organic fractions), total hydrolyzable amino acids (THAA), total hydrolyzable carbohydrates (THCO), lipids and acid insoluble fraction were used to study ¹⁴C (Wang et al. 1996, 1998; Hwang and Druffel 2003). Dried POC, zooplankton, phytoplankton, detrital aggregates on surface sediment and sediment from Stn M (34°50'N, 123°0'W, 4100 m depth) were extracted with methylene chloride:methanol to extract lipids. Half of the residue was hydrolyzed with 6M HCl under N₂ for THAA and the other half with 72% and 0.6M H₂SO₄. The hydrolyzates were eluted through ion exchange columns to separate the two fractions, and the organic C leftover from the THAA extraction was used for the acid insoluble fractions. All THAA and THCO fractions had higher Δ^{14} C values than those for the lipid and acid insoluble fractions (Wang et al. 1996, 1998; Hwang and Druffel 2003) (Figure 5). The proportion of acid insoluble fractions increased with depth in the water column, and was highest in sediment samples (49–69%) and lowest in zooplankton and phytoplankton (<10%) from the upper 100 m. These results implied that the acid insoluble fraction was composed of lipid-like macromolecules.

Compound classes have also been measured in HMW-DOC. Results similar to those obtained for POC and sedimentary organic C were obtained, where THAA and TCHO had higher Δ^{14} C values than those for lipid and acid insoluble fractions (Loh et al. 2004).

Dissolved black C (DBC) is incompletely combusted organic matter produced from fossil fuel and biomass burning. DBC was measured in HMW-DOC from 4 samples from the Pacific and Atlantic Oceans and found to have Δ^{14} C values that were 470–860‰ lower than those for bulk DOC from the same water (Ziolkowski and Druffel 2010). DBC was also measured in SPE-DOC from 6 samples in the Atlantic, Pacific and Arctic Oceans and DBC Δ^{14} C values were 0– 560‰ lower than those of SPE-DOC (Coppola and Druffel 2016). They concluded that there

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are two distinct pools of DBC, that in the surface ocean and that in the deep ocean. Coppola et al. (2014) measured black C in sinking POC and sediments and found that the main transport mechanism of black C to sediments may be sorption of ancient DBC onto POC. Further studies of DBC in marine systems are needed to understand more fully the sources and sinks of DBC and its residence time in the ocean.

Compound Specific Radiocarbon Analyses (CSRA)

Organic matter in the ocean consists of a complex mixture of different compounds, both of terrigenous and marine origin. Greater understanding of C cycling processes in the marine realm can be obtained by isolating specific fractions of marine organic matter that can be ascribed to a known source (biomarkers), by investigating the ¹⁴C signature of individual organic compounds. Compound-specific ¹⁴C analyses (CSRA) of marine biomarkers have been used to elucidate processes affecting the fate of marine organic matter during transport (e.g., Ohkouchi et al. 2002; Mollenhauer et al. 2003, 2007; Ausín et al. 2019), the reactivity of different types of marine organic compounds (e.g., Shah et al. 2008; Kusch et al. 2010, 2016), and C sources utilized for biosynthesis (Ingalls et al. 2006; Hansmann et al. 2009). Improvement of stratigraphies for sediments that are otherwise difficult to date have also relied on CSRA of marine organic compounds (Ingalls et al. 2004; Ohkouchi and Eglinton 2008). These studies have revealed the long-term residence of marine organic matter in continuous re-suspension loops (e.g., Mollenhauer et al. 2007, 2011), which has also been shown for terrigenous biomarkers (e.g., Bröder et al. 2018; Wei et al. 2021), its preservation in association with fine-grained lithogenic material resulting in long-distance lateral transport and the millennial scale duration of these transport processes (Ohkouchi et al. 2002; Mollenhauer et al. 2005, 2007). Chemoautotrophic metabolism has been shown to prevail among mesopelagic microbial communities (Ingalls et al. 2006). Steroids and pigments and their derivatives, biomineral occluded compounds and as nucleic acids (Cherrier et al. 1999) have been investigated. Recent developments use CSRA of amino acids to study their sources and cycling (Bour et al. 2016; Blattmann et al. 2020).

All applications of CSRA on marine organic compounds require extraction and purification of sufficient mass of the compound from samples where they are typically present in trace amounts. As a consequence, most CSRA studies are conducted using sediments that have sufficient material available. In most cases, CSRA of compounds isolated from POC or DOC sampled from the water column is not possible, unless dedicated sampling campaigns that collect several thousands of liters of water are conducted (e.g., Ingalls et al. 2006). An exception was sugars collected from HMW-DOC in surface waters of the North Pacific that had post-bomb Δ^{14} C values (Repeta and Aluwihare 2006).

Commonly used methods for extraction and purification targeting mostly lipid compounds have been summarized by Mollenhauer et al. (2019). Often, a sequence of wet-chemical and chromatographic techniques is applied, and typically samples of 100 μ g C or less are obtained. The samples need to be combusted prior to ¹⁴C analysis and in many cases will be graphitized. AMS analyses of small samples are conducted using size-matched standards and blanks for normalization. For such small samples, accurate determination of the amount and Δ^{14} C value of extraneous C added to the sample during the various steps of sample preparation is of critical importance (Santos et al. 2010). Recently, isotope dilution techniques are increasingly used to obtain reliable estimates of this extraneous C



Figure 6 Schematic of the RPO reactor (left) and the results (right) from a POC sample collected from the Narayani River, Nepal (Hemingway 2017). The top half of the quartz reactor heats the sample while the gases are swept into a CO₂ analyzer before passing through a cryogenic trap. The gray shaded region is the thermogram, the variation in ppm CO₂ vs. temperature (units not shown), white circles show the δ^{13} C values, and bars show the fraction modern values for each temperature interval (Hemingway 2017). The data show that the high temperature, more refractory, fractions have dramatically less ¹⁴C, i.e., are older than the lower temperature fractions.

contribution and its ¹⁴C content, which can subsequently be used to correct CSRA results (e.g., Haghipour et al. 2019; Sun et al. 2020).

Taking advantage of the elemental-analyzer-accelerator-mass-spectrometer (EA-AMS) setup, recent methodological developments allow for direct injection of CO_2 gas produced from combustion of purified compounds into a gas accepting ion source (Haghipour et al. 2019; Mollenhauer et al. 2021). Purified compounds are transferred into small tin cups, dried and combusted in the EA (Elementar Elemental Analyzer) coupled to the AMS. Thereby, the graphitization step is omitted, and high-precision ¹⁴C data can be obtained for samples containing 50 µg C or less. For samples younger than approximately 5000 years, precise ¹⁴C analyses can be achieved for samples as small as 10 µg C (Haghipour et al. 2019).

Ramped Pyrolysis Oxidation (RPO) and ¹⁴C

In the mid-2000s, attempts to understand the chronology of Antarctic sediments led John Hayes to develop the Ramped Pyrolysis Oxidation system (RPO) at the National Ocean Sciences AMS facility, a system that separates organic matter based on thermochemical properties. RPO ¹⁴C analysis involves the controlled step heating of samples containing organic C, with or without oxygen, typically at a heating rate of 5°C/minute (Figure 6). As heating proceeds, the most reactive organic components become volatile and are swept into an oxidizing reactor by a helium stream, allowing separation of CO₂ from other pyrolysis/ combustion components and collection of CO₂ over discrete temperature intervals for analysis of both stable and radiocarbon isotopes. A major advantage is the ability to analyze all of the organic matter in a sample, not just an isolated fraction. In sediments, the thermal separation allows the isolation of fresh detrital matter from eroded bedrock. The initial goal of improving the chronologies of Antarctic sediments was realized (Rosenheim et al. 2008) and has been expanded upon (Rosenheim et al. 2013; Subt et al. 2016).

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Building on this success, RPO has since been applied to a wide variety of biogeochemical questions in many environments including the oceans. It is being used to study the nature of the organic material transported to the oceans from rivers (Hemingway et al. 2019). Hemingway et al. (2017) developed an inverse method to calculate organic C reactivity from RPO results. They used the method on RPO results from 62 sediment organic C, riverine POC, soil organic C, and DOC samples to indicate that mineral interactions are more important for the preservation of organic C than selective preservation (Hemingway et al. 2019). RPO has been applied to studying the fate of petroleum hydrocarbons, both natural and pollutant, in marine and coastal ecosystems (Pendergraft et al. 2013; Pendergraft and Rosenheim 2014; Adikhari et al. 2016; Rogers et al. 2019). A time series study of the impact of the Deep Water Horizon oil spill on sediments from four sites demonstrated the range of impacts and the time evolution of the oil spill. RPO and C isotope analysis allowed the separation of the spilled petrocarbon from natural hydrocarbon seep C. Emerging work is using the technique to explore the nature of DOC, particularly the high molecular weight fraction.

EA-AMS for Suspended POC ¹⁴C

During oceanographic expeditions, suspended POC (POC_{susp}) for radiocarbon analysis is typically collected using in-situ water pumps with GF/F filters that are lowered to water depths where up to 1000 L are pumped (Griffith et al. 2012). Because water pumps are expensive, multiple casts per station are required to obtain full-depth water column ¹⁴C POC_{susp} profiles, a time consuming process that is also expensive.

Utilizing the EA-AMS setup can overcome in-situ water pump limitations, because the MICADAS (MIniCArbonDAtingSystem) allows routine, high precision analysis of small samples (> 100 μ gC). For EA-AMS, seawater is processed onboard directly from CTD-ROS (Conductivity Temperature Depth-Rosette) niskin bottles, where 2–40 L of water are collected to obtain sufficient C on GF/F filters, depending on the POC_{susp} concentration. It is possible to sample up to 24 depths during one deployment and generate high resolution depth profiles of POC_{susp} ¹⁴C (Figure 7). POC_{susp} is concentrated on pre-combusted GF/F filters using a vacuum supported glass filtration device that allows the sampling of filtrate to obtain the corresponding DOC fraction. In the lab, the filters are acidified, packed into tin capsules, and combusted in the EA. The generated CO₂ is measured in the MICADAS via direct injection into the gas ion source (Mollenhauer et al. 2021).

The conventional CuO oxidation reactor of the EA operating at 950°C is prone to cracking, due to halogen radicals reacting with CuO that attack the quartz reactor. Recently, a modified EA reactor has replaced the CuO with WO₃ and changed the position of the Ag wool. The WO₃ does not react with halogen radicals and Ag wool is positioned at low reaction temperatures further preventing unwanted side reactions. The reaction temperature is lowered to 850°C, and an Inconel liner and ash fingers are used to further protect the quartz reactor. This reactor setup withstands 150–200 samples, whereas a typical CuO reactor processing saline POC_{susp} filters cannot withstand more than 50 samples.

The disadvantage of this setup is that the C content of the blank increases significantly. The C content of a typical blank for a pre-combusted $\frac{1}{2}$ 45 mm GF/F analyzed on a CuO reactor is ~10 µg C, but for the modified WO₃ reactor it is ~20 µgC. The higher blank C originates from the WO₃ and can be reduced by carefully inspecting the material and removing incompletely



Figure 7 Preliminary F¹⁴C POC_{susp} values of samples collected along a shelf to open ocean transect off Cape Blanc (NW Africa, ~21°N) collected in November 2018 during research cruise MSM79 onboard the German R/V *Maria S. Merian* and processed according to the EA–AMS method described above (H. Grotheer unpublished data). Red dots indicate sampling depths. (The map was generated using Ocean Data View; Schlitzer 2015).

oxidized tungsten particles and foreign objects. Nonetheless, pre-combusted GF/F filters contain a significant amount of blank C and appropriate blank determination and subsequent correction is critical. The high blank C content has further implications for the minimum amount of seawater that needs to be filtered per sample in order to acquire an optimum blank C to POC_{susp} ratio.

In the first example of a high-resolution $F^{14}C$ (Fraction modern ${}^{14}C$) POC_{susp} profile (Figure 7, H. Grotheer unpublished data) was generated for a transect from the shelf to the open ocean off Cape Blanc (NW Africa, ~21°N). The Cape Blanc region is located in the southern part of the Canary Current Eastern Boundary Upwelling system off northwest Africa. Here, the Cape Blanc upwelling filament transports cold, nutrient-rich, upwelled waters for hundreds of kilometers offshore towards the oligotrophic ocean (Van Camp et al. 1991; Gabric et al. 1993; Àlvarez-Salgado et. al. 2001) and is an important transport pathway for particles (Karakas et al. 2006).

Fresh POC_{susp} (F¹⁴C 1.0) produced by primary production on the shelf is transported along the continental slope within the bottom layer and travels offshore at ~500 m water depth. Further offshore, underneath the photic zone, F¹⁴C POC_{susp} decreases to ~0.8 at 500 m depth. Below 500 m, F¹⁴C values show a heterogeneous distribution with pre-aged POC_{susp} (F¹⁴C ~0.6) separated by layers of POC_{susp} with intermediate F¹⁴C values, suggesting the presence of a refractory POC_{susp} pool, potentially originating from sediment resuspension. These preliminary data highlight the potential of the EA-AMS method to generate high-resolution, F¹⁴C POC_{susp} profiles that will advance our understanding of POC cycling, transport and sequestration processes in the ocean.

FUTURE WORK

Studies of marine organic C have benefitted from research conducted outside of the marine scientific community. Effort to continue this strong communication and collaboration is important to maintain and to nurture. New ways to unlock the secrets of the

biogeochemistry of the marine system will benefit humanity as we chart a course through the Anthropocene.

REFERENCES

- Adhikari PL, Maiti K, Overton EB, Rosenheim BE, Marx BD. 2016. Distributions and accumulation rates of polycyclic aromatic hydrocarbons in the northern Gulf of Mexico sediments. Environmental Pollution 212:413–423. doi: 10. 1016/j.envpol.2016.01.064.
- Ålvarez-Salgado XA, Doval MD, Borges AV, Joint I, Frankignoulle M, Woodward M, Figueiras FG. 2001. Off-shelf fluxes of labile materials by an upwelling filament in the NW Iberian upwelling system. Progress in Oceanography 51(2–4):321– 337. doi: 10.1016/S0079-6611(01)00073-8.
- Amador J, Milne PJ, Moore CA, Zika RG. 1990. Extraction of chromophoric humic substances from seawater. Marine Chemistry 29:1–17. doi: 10.1016/0304-4203(90)90002-T.
- Ausín B, Magill C, Haghipour N, Fernández A, Wacker L, Hodell D, Baumann K-H, Eglinton TI. 2019. (In)coherent multiproxy signals in marine sediments: Implications for highresolution paleoclimate reconstruction. Earth and Planetary Science Letters 515:38–46. doi: 10.1016/j.epsl.2019.03.003.
- Beaupré SR, Aluwihare L. 2010. Constraining the 2component model of marine dissolved organic radiocarbon. Deep Sea Research Part II: Topical Studies in Oceanography 57(16):1494– 1503. doi: 10.1016/j.dsr2.2010.02.017.
- Beaupré SR, Walker BD, Druffel ERM. 2020. The two-component model coincidence: Evaluating the validity of marine dissolved organic radiocarbon as a stable-conservative tracer at Station M. Deep-Sea Research II 173:104737.
- Beaupré SR, Druffel ERM, Griffin S. 2007. A low– blank photochemical extraction system for concentration and isotopic analyses of marine dissolved organic carbon. Limnology and Oceanography: Methods 5:174–184.
- Beaupré et al. 2019. Oceanic efflux of ancient marine dissolved organic carbon in primary marine aerosol. Science Advances 5:eaax6535.
- Benner R, Biddanda B, Black B, McCarthy M. 1997. Abundance, size distribution, and stable carbon and nitrogen isotopic compositions of marine organic matter isolated by tangential-flow ultrafiltration. Marine Chemistry 57:243–263.
- Benner R, Paculksy J, McCarthy M, Hedges J, et al. 1992. Bulk chemical characteristics of dissolved organic matter in the ocean. Science 255:1561– 1564.
- Bercovici SK, McNichol AP, Xu L, Hansell DA. 2018. Radiocarbon content of dissolved organic carbon in the South Indian Ocean. Geophysical Research Letters 45(2):872–879. doi: 10.1002/ 2017gl076295.

- Blattmann TM, Montluçon DB, Haghipour N, Ishikawa NF, Eglinton TI. 2020. Liquid chromatographic isolation of individual amino acids extracted from sediments for radiocarbon analysis. Front. Mar. Sci. 7:174. doi: 10.3389/ fmars.2020.00174.
- Bianchi TS. 2011. The role of terrestrially derived organic carbon in the coastal ocean: a changing paradigm and the priming effect. Proc. National Academy Sciences 108(49):19473–19481.
- Bien G, Rakestraw NW, Suess HE. 1965. Radiocarbon in the Pacific and Indian Oceans and its relation to deep water movements. Limnology and Oceanography 10:R25–R36.
- Bour AL, Walker BD, Broek TAB, McCarthy MD. 2016. Radiocarbon analysis of individual amino acids: carbon blank quantification for a smallsample high-pressure liquid chromatography purification method Analytical Chemistry 88(7):3521–3528. doi: 10.1021/acs.analchem. 5b03619.
- Bröder L, Tesi T, Andersson A, Semiletov I, Gustafsson Ö. 2018. Bounding cross-shelf transport time and degradation in Siberian-Arctic land-ocean carbon transfer. Nature Communications 9(1):806. doi: 10.1038/s41467-018-03192-1.
- Broecker WS, Gerard R, Ewing M, and Heezen BC. 1960. Natural radiocarbon in the Atlantic Ocean: Jour. Geophys. Research 55:2903–2931.
- Broek TAB, Walker BD, Guilderson TP, McCarthy MD. 2017. Coupled ultrafiltration and solid phase extraction approach for the targeted study of semi-labile high molecular weight and refractory low molecular weight dissolved organic matter. Marine Chemistry 194:146–157.
- Broek TA, Walker BD, Guilderson TP, Vaughn JS, Mason HE, McCarthy MD. 2020. Low molecular weight dissolved organic carbon: aging, compositional changes, and selective utilization during global ocean circulation. Global Biogeochemical Cycles 34(6):1–20. doi: 10.1029/2020GB006547.
- Cherrier J, Bauer J, Druffel ERM, Coffin R, Chanton J. 1999. Radiocarbon in marine bacteria: Evidence for the ages of assimilated carbon. Limnology and Oceanography 44(3):730–736.
- Coppola AI, Ziolkowski LA, Masiello CA, Druffel ERM. 2014. Aged black carbon in marine sediments and sinking particles. Geophys. Res. Lett. 41. doi: 10.1002/2013GL059068.
- Coppola AI, Druffel ERM. 2016. Cycling of black carbon in the ocean. Geophysical Research Letters 43(9):4477–4482. doi: 10.1002/ 2016GL068574.

- Dittmar T, Koch BP. 2006. Thermogenic organic matter dissolved in the abyssal ocean. Marine Chemistry 102(3–4):208–217. doi: 10.1016/j. marchem.2006.04.003.
- Dittmar T, Fitznar HP, Kattner G. 2001. Origin and biogeochemical cycling of organic nitrogen in the eastern Arctic Ocean as evident from D- and Lamino acids. Geochim. Cosmochim. Acta 65:4103–4114.
- Dittmar T, Koch B, Hertkorn N, Kattner G. 2008. A simple and efficient method for the solid-phase extraction of dissolved organic matter (SPE-DOM) from seawater. Limnology and Oceanography: Methods 6(6):230–235.
- Druffel ERM, Bauer JE. 2000. Radiocarbon distributions in Southern Ocean dissolved and particulate organic matter. Geophysical Research Letters 27(10):1495–1498. doi: 10. 1029/1999GL002398.
- Druffel ERM, Griffin S. 2015. Radiocarbon in dissolved organic carbon of the South Pacific Ocean, Geophy. Res. Lett. 42:4096–4101. doi: 10.1002/2015GL063764.
- Druffel ERM, Bauer JE, Williams PM, Griffin S, Wolgast D. 1996. Seasonal variability of radiocarbon in particulate organic carbon in the Northeast Pacific Ocean. Journal of Geophysical Research 101:20543–20552.
- Druffel ERM, Griffin S, Coppola AI, Walker BD. 2016. Radiocarbon in dissolved organic carbon of the Atlantic Ocean. Geophysical Research Letters 43. doi: 10.1002/2016GL068746.
- Druffel ERM, Griffin S, Garcia NG, Wang N, McNichol AP, Key R, Walker BD. 2019. Dissolved organic radiocarbon in the central Pacific Ocean, Geophysical Research Letters 46. doi: 10.1029/2019GL083149.
- Druffel ERM, Williams PM, Bauer JE, Ertel JR. 1992. Cycling of dissolved and particulate organic matter in the open ocean. Journal of Geophysical Research 97(C10):15639. doi: 10. 1029/92JC01511
- Estes ER, Berti D, Coffey NR, Hochella MF, Wozniak AS, Luther GW. 2019. Abiotic synthesis of graphite in hydrothermal vents. Nature Communications 10. doi: 10.1038/ s41467-019-13216-z.
- Fang L, Lee SH, Lee S-A, Hahm D, Kim G, Druffel ERM, Hwang J. 2020. Removal of refractory dissolved organic carbon in the Amundsen Sea, Antarctica. Scientific Reports. doi: 10.1038/ s41598-020-57870-6.
- Flerus R, Lechtenfeld OJ, Koch BP, McCallister SL, Schmitt-Kopplin P, Benner R, Kaiser K, Kattner G. 2012. A molecular perspective on the ageing of marine dissolved organic matter. Biogeosciences 9(6):1935–1955. doi: 10.5194/bg-9-1935-2012
- Gabric AJ, Garcia L, Van Camp L, Nykjaer L, Eifler W, Schrimpf W. 1993. Offshore export of shelf production in the Cape Blanc (Mauritania)

giant filament as derived from coastal zone color scanner imagery. Journal of Geophysical Research 98:4697–4712. doi: 10.1029/92jc01714.

- Green SA, Blough NV. 1994. Optical absorption and fluorescence properties of chromophoric dissolved organic matter in natural waters. Limnology and Oceanography 39(8):1903–1916. doi: 10.4319/lo.1994.39.8.1903.
- Griffith DR, McNichol AP, Xu L, McLaughlin FA, Macdonald RW, Brown KA, Eglinton TI. 2012. Carbon dynamics in the western Arctic Ocean: insights from full-depth carbon isotope profiles of DIC, DOC, and POC. Biogeosciences 9. doi: 10.5194/bg-9-1217-2012.
- Haghipour N, Ausin B, Usman MO, Ishikawa N, Wacker L, Welte C, Ueda K, Eglinton TI. 2019. Compound-Specific Radiocarbon Analysis by Elemental Analyzer–Accelerator Mass Spectrometry: precision and limitations. Analytical Chemistry 91(3):2042–2049. doi: 10. 1021/acs.analchem.8b04491.
- Hansell DA. 2013. Recalcitrant dissolved organic carbon fractions. Annual Reviews of Marine Science 5:421–448.
- Hansman RL, Griffin S, Watson JT, Druffel ERM, Ingalls AE, Pearson A, Aluwihare LI. 2009. The radiocarbon signature of microorganisms in the mesopelagic ocean. Proceedings of the National Academy of Sciences 106:6513–6518. doi: 10.1073/Pnas.0810871106.
- Hedges JI. 1992. A comparison of dissolved humic substances from seawater with Amazon River counterparts by 13C-NMR spectroscopy. Geochemica et Cosmochimica Acta 56(4):1753– 1757.
- Hedges JI, et al. 2000. The molecularlyuncharacterized component of nonliving organic matter in natural environments. Organic Geochemistry 31:945–958.
- Hemingway JD, Galy VV, Gagnon AR, Grant KE, Rosengard, SZ, Soulet G, Zigha PK, McNichol AP. 2017. Assessing the blank carbon contribution, isotope mass balance and kinetic isotope fractionation of the Ramped Pyrolysis/ Oxidation Instrument at NOSAMS. Radiocarbon 59:179–193. doi: 10.1017/RDC.2017.3.
- Hemingway, JD, Rothman DH, Rosengard SZ, Galy VV. 2017. Technical note: An inverse method to relate organic carbon reactivity to isotope composition from serial oxidation. Biogeosciences 14(22):5099–5114. doi: 10.5194/ bg-14-5099-2017.
- Hemingway JD, Rothman DH, Grant KE, Rosengard SZ, Eglinton TI, Derry LA, Galy VV. 2019. Mineral protection regulates longterm global preservation of natural organic carbon. Nature 570(7760):228–231. doi: 10. 1038/s41586-019-1280-6.
- Hertkorn N, Harir M, Koch BP, Michalke B, Schmitt-Kopplin P. 2013. High-field NMR

spectroscopy and FTICR mass spectrometry: Powerful discovery tools for the molecular level characterization of marine dissolved organic matter. Biogeosciences 10(3):1583–1624. doi: 10. 5194/bg-10-1583-2013.

- Hwang J, Druffel ERM. 2003. Lipid–like material as the source of the uncharacterized organic carbon in the ocean? Science 299:881–884.
- Hwang J, Druffel ERM, Eglinton TI. 2010. Widespread influence of resuspended sediments on oceanic particulate organic carbon: Insights from radiocarbon and aluminum contents in sinking particles, Global Biogeochemical Cycles 24:GB4016. doi: 10.1029/2010GB003802.
- Ingalls A, Anderson RF, Pearson A. 2004. Radiocarbon dating of diatom-bound organic compounds. Marine Chemistry 92:91–105.
- Ingalls AE, Shah SR, Hansman RL, Aluwihare LI, Santos GM, Druffel ERM, Pearson A. 2006. Quantifying archeal community autotrophy in the mesopelagic ocean using natural radiocarbon. Proceedings of the National Academy of Sciences 103(17):6442–6447. doi: 10.1073/pnas.0510157103.
- Jiao N, Herndl GJ, Hansell DA, Benner R, Kattner G, Wilhelm SW, Kirchman DL, Weinbauer TL, Chen F, Azam F. 2010. Microbial production of recalcitrant dissolved organic matter: longterm carbon storage in the global ocean. Nature Reviews Microbiology 8:593–599. doi: 10.1038/ nrmicro2386.
- Karakas G, Nowald N, Blaas M, Marchesiello P, Frickenhaus S, Schlitzer R. 2006. Highresolution modeling of sediment erosion and particle transport across the northwest African shelf. Journal of Geophysical Research 111: C06025. doi: 10.1029/2005JC003296.
- Key RM, Kozyr A, Sabine CL, Lee K, Wanninkhof R, Bullister JL, et al. 2004. A global ocean carbon climatology: Results from Global Data Analysis Project (GLODAP). Global Biogeochemical Cycles 18:GB4031. doi: 10.1029/ 2004GB002247.
- Kieber RJ, Hydro LH, Seaton PJ. 1997. Photooxidation of triglycerides and fatty acids in seawater: Implication toward the formation of marine humic substances. Limnology and Oceanography 42(6):1454–1462. doi: 10.4319/lo. 1997.42.6.1454.
- Kusch S, Kashiyaman, Ogawa AN, Altabet M, Butzin M, Friedrich J, Ohkouchi N, Mollenhauer G. 2010. Implications for chloroand pheopigment synthesis and preservation from combined compound-specific d¹³C, d¹⁵N, and Δ^{14} C analysis. Biogeosciences 7(12):4105– 4118. doi: 10.5194/bg-7-4105-2010.
- Kusch S, Rethemeyer J, Hopmans EC, Wacker L, Mollenhauer G. 2016. Factors influencing ¹⁴C concentrations of algal and archaeal lipids and their associated sea surface temperature proxies

in the Black Sea. Geochimica et Cosmochimica Acta 188:35–57. doi: 10.1016/j.gca.2016.05.025.

- Lang SQ, Butterfield DA, Lilley MD, Paul Johnson H, Hedges JI. 2006. Dissolved organic carbon in ridge-axis and ridge-flank hydrothermal systems. Geochimica et Cosmochimica Acta 70:3830–3842. doi: 10.1016/j.gca.2006.04.031.
- Lechtenfeld OJ, Hertkorn N, Shen Y, Witt M, Benner R. 2015. Marine sequestration of carbon in bacterial metabolites. Nature Communications 6(1):8. doi: 10.1038/ncomms7711.
- Lechtenfeld OJ, Kattner G, Flerus R, McCallister SL, Schmitt-Kopplin P, Koch BP. 2014. Molecular transformation and degradation of refractory dissolved organic matter in the Atlantic and Southern Ocean. Geochimica et Cosmochimica Acta 126:321–337. doi: 10.1016/j.gca.2013.11. 009.
- Lewis CB, Walker BD, Druffel ERM. 2020. Isotopic and optical heterogeneity of solid phase extracted marine dissolved organic carbon. Marine Chemistry 219:103752. doi: 10.1016/j.marchem. 2020.103752
- Lewis CB, Walker BD, Druffel ERM. 2021. Constraining the cycling of global solid-phase extracted dissolved organic carbon using radiocarbon measurements, Geophysical Research Letters. doi: 10.1002/(ISSN)1944-8007.
- Li Y, Harir M, Lucio M, Kanawati B, Smirnov K, Flerus R, Koch BP, Schmitt-Kopplin P, Hertkorn N. 2016. Proposed guidelines for solid phase extraction of Suwannee River dissolved organic matter. Analytical Chemistry 88(13): 6680–6688. doi: 10.1021/acs.analchem.5b04501.
- Li Y, Harir M, Uhl J, Kanawati B, Lucio M, Smirnov KS, Koch BP, Schmitt-Kopplin P, Hertkorn N. 2017. How representative are dissolved organic matter (DOM) extracts? A comprehensive study of sorbent selectivity for DOM isolation. Water Research 116:316–323. doi: 10.1016/j.watres. 2017.03.038.
- Loh AN, Bauer JE, Druffel ERM. 2004. Variable aging and storage of dissolved organic components in the open ocean. Nature 430(7002):877–881.
- McCarthy M, Hedges J, Benner R. 1993. The chemical composition of dissolved organic matter in seawater. Chemical Geology 107(3– 4):503–507.
- McCarthy MD, Beaupré SR, Walker BD, Voparil I, Guilderson TP, Druffel ERM. 2011. Chemosynthetic origin of ¹⁴C-depleted dissolved organic matter in a ridge-flank hydrothermal system. Nature Geoscience 4:32–36. doi: 10. 1038/ngeo1015.
- McNichol AP, Aluwihare LI. 2007. The power of radiocarbon in biogeochemical studies of the marine carbon cycle: insights from studies of dissolved and particulate organic carbon (DOC and POC). Chem. Rev. 107:443–466.

- McNichol AP, Key RM, Guilderson TO. 2022. Global ocean radiocarbon programs. Radiocarbon. In press (this issue).
- Mollenhauer G, Eglinton TI, Ohkouchi N, Schneider RR, Müller PJ, Grootes PM, Rullkötter J. 2003. Asynchronous alkenone and foraminifera records from the Benguela Upwelling System. Geochimica et Cosmochimica Acta 67(12): 2157–2171.
- Mollenhauer G, Grotheer H, Gentz T, Bonk E, Hefter J. 2021. Standard operation procedures and performance of the MICADAS radiocarbon laboratory at Alfred Wegener Institute (AWI), Germany. Nuclear Instruments and Methods in Physics Research Section B: Beam Interactions with Materials and Atoms 496:45–51. doi: 10.1016/j.nimb.2021.03.016.
- Mollenhauer G, Inthorn M, Vogt T, Zabel M, Sinninghe Damsté JS, Eglinton TI. 2007. Aging of marine organic matter during cross-shelf lateral transport in the Benguela upwelling system revealed by compound-specific radiocarbon dating, Geochemistry, Geophysics, Geosystems G3 8(9):Q09004. doi: 10.1029/ 2007GC001603.
- Mollenhauer G, Kienast M, Lamy F, Meggers H, Schenider RR, Hayes JM, Eglinton TI. 2005. An evaluation of ¹⁴C age relationships between co-occurring foraminifera, alkenones, and total organic carbon in continental margin sediments. Paleoceanography 20:PA1016. doi: 10.1029/ 2004PA001103.
- Mollenhauer G, Kusch S, Eglinton TI, Pearson A. 2019. Compound-specific radiocarbon measurements. In: Kirk CJ, Bokuniewicz JH, Yager LP, editors. Encyclopedia of ocean sciences. 3rd edition. Vol. 1:235–244. Elsevier. ISBN: 978-0-12-813081-0.
- Mollenhauer G, McManus JF, Wagner T, McCave IN, Eglinton TI. 2011. Radiocarbon and ²³⁰Th data reveal rapid redistribution and temporal changes in sediment focussing at a North Atlantic drift. Earth and Planetary Science Letters 301:373–382. doi: 10.1016/j.epsl.2010.11. 022.
- Mopper K, Stubbins A, Ritchie JD, Bialk HM, Hatcher PG. 2007. Advanced instrumental approaches for characterization of marine dissolved organic matter: extraction techniques, mass spectrometry, and nuclear magnetic resonance spectroscopy. Chemical Reviews 107(2):419–442. doi: 10.1021/cr050359b
- Ohkouchi N, Eglinton TI. 2006. Radiocarbon constraint on relict organic carbon contributions to Ross Sea sediments, Geochemistry, Geophysics, Geosystems G3 7(4):Q04012. doi: 10.1029/2005GC001097.
- Ohkouchi N, Eglington TI. 2008. Compound-specific radiocarbon dating of Ross Sea sediments: a prospect for constructing chronologies in

high-latitude oceanic sediments. Quaternary Geochronology 3(3):235–243.

- Ohkouchi N, Eglinton TI, Keigwin LD, Hayes JM. 2002. Spatial and temporal offsets between proxy records in a sediment drift. Science 298:1224–1227.
- Osterholz H, Niggemann J, Giebel HA, Simon M, Dittmar T. 2015. Inefficient microbial production of refractory dissolved organic matter in the ocean. Nature Communications 6:7422. doi: 10.1038/ncomms8422.
- Ostlund G, Stuiver M. 1980. GEOSECS Pacific radiocarbon. Radiocarbon 22(1):25–53.
- Pendergraft MA, Dincer Z, Sericano JL, Wade TL, Kolasinski J, Rosenheim BE. 2013. Linking ramped pyrolysis isotope data to oil content through PAH analysis. Environmental Research Letters 8(4). doi: 10.1088/1748-9326/8/ 4/044038.
- Pendergraft MA, Rosenheim BE. 2014. Varying relative degradation rates of oil in different forms and environments revealed by ramped pyrolysis. Environmental Science, Technology 48(18):10966–10974. doi: 10.1021/es501354c.
- Peuravuori J, Pihlaja K. 1997. Molecular size distribution and spectroscopic properties of aquatic humic substances. Analytica Chimica Acta 337(2):133–149.
- Repeta DJ, Aluwihare LI. 2006. Radiocarbon analysis of neutral sugars in high-molecularweight dissolved organic carbon: implications for organic carbon cycling. Limnology and Oceanography 51(1):1045–1053.
- Rogers KL, Bosman SH, Lardie-Gaylord M, McNichol A, Rosenheim BE, Montoya JP, et al. 2019. Petrocarbon evolution: ramped pyrolysis/oxidation and isotopic studies of contaminated oil sediments from the Deepwater Horizon oil spill in the Gulf of Mexico. PLoS ONE 14(2):e0212433. doi: 10.1371/journal.pone. 0212433.
- Rosenheim BE, Day MB, Domack E, Schrum H, Benthien A, Hayes JM. 2008. Antarctic sediment chronology by programmedtemperature pyrolysis: Methodology and data treatment, Geochem. Geophys. Geosyst. 9(4): Q04005 doi: 10.1029/2007GC001816.
- Rosenheim BE, Santoro JA, Gunter M, Domack EW. 2013. Improving Antarctic sediment ¹⁴C dating using ramped pyrolysis: an example from the Hugo Island Trough. Radiocarbon 55(1):115–126.
- Santos GM, Southon JS, Drenzek NJ, Ziolkowski L, Druffel ERM, Xu X, Zhang D, Trumbore SE, Eglinton TI, Hughen KA. 2010. Blank assessment for ultra-small radiocarbon samples: chemical extraction and separation vs. AMS. Radiocarbon 52(3):1322–1335.
- Santschi PH, Guo LD, Baskaran M, Trumbore S, Southon J, Bianchi TS, Honeyman B, Cifuentes L. 1995. Isotopic evidence for the contemporary

origin of high-molecular-weight organic-matter in oceanic environments. Geochim. Cosmochim. Acta 59:625–631.

Schlitzer R. 2015. Ocean data view. http://odv.awi.de

- Shah SR, Mollenhauer G, Ohkouchi N, Eglinton TI, Pearson A. 2008. Origins of archaeal tetraether lipids in sediments: Insights from radiocarbon analysis. Geochimica et Cosmochimica Acta 72(18):4577–4594.
- Shaw Walter SR, Jaekel U, Osterholz H, Fisher AT, Huber JA, Pearson A, et al. 2018. Microbial decomposition of marine dissolved organic matter in cool oceanic crust. Nature Geoscience 11. doi: 10.1038/s41561-018-0109-5.
- Shen Y, Benner R. 2018. Mixing it up in the ocean carbon cycle and the removal of refractory dissolved organic carbon. Scientific Reports. doi: 10.1038/s41598-018-20857-5.
- Simjouw J-P, Minor EC, Mopper K. 2005. Isolation and characterization of estuarine dissolved organic matter: comparison of ultrafiltration and C18 solid-phase extraction techniques. Marine Chemistry 96(3–4):219–235. doi: 10. 1016/j.marchem.2005.01.003.
- Skoog A, Benner R. 1997. Aldoses in various size fractions of marine organic matter: Implications for carbon cycling. Limnology And Oceanography 42:1803–1813.
- Smith KLJ, Baldwin RJ, Williams PM. 1992. Reconciling particulate organic carbon flux and sediment community oxygen consumption in the deep North Pacific. Nature 359:313–315. doi: 10.1038/359313a0
- Stuiver M, Quay PD, Ostlund HG. 1983. Abyssal water ¹⁴C distribution and the age of the world oceans. Science 219(4586):849–851. doi: 10. 1126/science.219.4586.849.
- Subt C, Fangman KA, Wellner JS, Rosenheim BE. 2016. Sediment chronology in Antarctic deglacial sediments: reconciling organic carbon ¹⁴C ages to carbonate ¹⁴C ages using ramped PyrOx. The Holocene 26(2):265–273. doi: 10. 1177/0959683615608688.
- Sun S, et al. 2020. ¹⁴C blank assessment in small-scale compound-specific radiocarbon analysis of lipid biomarkers and lignin phenols. Radiocarbon 62(1):207–218. doi: 10.1017/RDC.2019.108.
- Van Camp L, Nykjaer L, Mittelstaedt E, Schlittenhardt P. 1991. Upwelling and boundary circulation off northwest Africa as depicted by infrared and visible satellite observations. Prog. Oceanogr. 26:357–402.
- Verdugo P, Alldredge AL, Azam F, Kirchmand DL, Passowe U, Santschif PH. 2004. The oceanic gel phase: a bridge in the DOM–POM continuum. Marine Chemistry 92:67–85. doi: 10.1016/j. marchem.2004.06.017
- Walker B, Beaupre S, Guilderson T, Druffel ERM, McCarthy M. 2011. Large-volume ultrafiltration for the study of radiocarbon signatures and size vs. age relationships in

marine dissolved organic matter. Geochimica et Cosmochimica Acta 75:5187–5202.

- Walker BD, et al. 2016a. Linked changes in marine dissolved organic carbon molecular size and radiocarbon age. Geophysical Research Letters 43:10,385–10,393.
- Walker BD, Beaupre SR, Guilderson TP, McCarthy MD, Druffel ERM. 2016b. Pacific carbon cycling constrained by organic matter size, age and composition relationships. Nature Geoscience 9:888–891.
- Walker BD, Beaupré SR, Griffin S, Druffel ERM. 2019. UV photochemical oxidation and extraction of marine dissolved organic carbon at UC Irvine: status, surprises, and methodological recommendations. Radiocarbon 61:1603–1618. doi: 10.1017/RDC.2019.9.
- Wakeham SG, Lee C, Hedges JI, Hermes PJ, Peterson ML. 1997. Fate of major biochemicals in water column particles and sediments of the central Equatorial Pacific Ocean. Geochimica et Cosmochimica Acta 61:5363–5369.
- Wang X-C, Druffel ERM, Lee C. 1996. Evidence for sorption of old carbon to particulate material in the deep sea from radiocarbon in organic compound classes. Geophysical Research Letters 23:3583–3586.
- Wang X-C, Druffel ERM, Lee C. 1998. Radiocarbon studies of organic compound classes in plankton and sediment of the NE Pacific Ocean. Geochim Cosmochim Acta 62:1365–1378.
- Wei B, Mollenhauer G, Hefter J, Kusch S, Grotheer H, Schefuß E, Jia G. 2021. The nature, timescale, and efficiency of riverine export of terrestrial organic carbon in the (sub)tropics: insights at the molecular level from the Pearl River and adjacent coastal sea. Earth and Planetary Science Letters 565:116934. doi: 10.1016/j.epsl. 2021.116934.
- Williams PM, Druffel ERM. 1987. Radiocarbon in dissolved organic carbon in the central North Pacific Ocean. Nature 330:246–248.
- Williams PM, Oeschger H, Kinney P. 1969. Natural radiocarbon activity of the dissolved organic carbon in the Northeast Pacific. Nature 224:256–258.
- Williams PM, McGowan JA, Stuiver M. 1970. Bomb carbon-14 in deep sea organisms. Nature 227(5256):375–376.
- Williams PM, Linick TW. 1975. Cycling of organic carbon in the ocean: Use of naturally occurring radiocarbon as a long and short term tracer. IAEA, Vienna. Symposium on isotope ratios as pollutant source and behavior indicators. Vienna, Austria, 18 Nov 1974. IAEA-SM–191/26.
- Van Camp L, Nykjaer L, Mittelstaedt E, Schlittenhardt P. 1991. Upwelling and boundary circulation off Northwest Africa as depicted by infrared and visible satellite observations. Progress in Oceanography 26:357–402. doi: 10. 1016/0079-6611(91)90012-B.

- Xue Y, Tiantian GE, Xuchen W. 2015. An effective method of UV-oxidation of dissolved organic carbon in natural waters for radiocarbon analysis by accelerator mass spectrometry. J. Ocean Univ. China. doi: 10.1007/s11802-015-2935-z.
- Zigah PK, McNichol, AP, Xu L, Johnson C, Santinelli C, Karl DM, Repeta DJ. 2017. Allochthonous sources and dynamic cycling

of ocean dissolved organic carbon revealed by carbon isotopes. Geophysical Research Letters 44:2407–2415. doi: 10.1002/2016GL07 1348.

Ziolkowski LA, Druffel ERM. 2010. Aged black carbon identified in marine dissolved organic carbon, Geophysical Research Letters 37: L16601. doi: 10.1029/2010GL043963.