Proceedings of the Nutrition Society (2019), **78**, 313–318 © The Author 2019 First published online 1 February 2019

The Nutrition Society Summer Meeting was held at the University of Leeds, UK, on 10-12 July 2018

Conference on 'Getting energy balance right' Symposium 2: Molecular mechanisms contributing to energy balance

Dietary modulation of energy homoeostasis and metabolic-inflammation

Helen M Roche^{1,2}

¹Nutrigenomics Research Group, School of Public Health, Physiotherapy and Sports Science, UCD Institute of Food and Health, Diabetes Complications Research Centre, University College Dublin, Dublin, Ireland ²Institute for Global Food Security, Queen's University Belfast, Northern Ireland, UK

> Dietary intake and nutritional status is an important environmental factor which can modulate metabolic-inflammation. In recent years, research has made significant advances in terms of understanding the impact of dietary components on metabolic-inflammation, within the context of obesity, type-2 diabetes (T2D) and CVD risk. Our work demonstrated that different fatty acids differentially modulate metabolic-inflammation, initially focusing on Nod-like receptor family, pyrin domain-containing three protein (NLRP3) inflammasome mediated IL-1 β biology and insulin signalling. However, the paradigm is more complex, wherein data from the immunology field clearly show that nature of cellular energy metabolism is a key determinant of inflammation. Whilst metabolic-inflammation is a critical biological interaction, there is a paucity of data in relation to the nature and the extent to which nutritional status affects metabolic-inflammation. The complex paradigm will be discussed within the context of if/how dietary components, in particular fatty acids, may modulate obesity, T2D and CVD risk, via inflammatory and metabolic processes.

Metabolic-inflammation: Fatty acids: Nod-like receptor family, pyrin domain-containing three protein: IL-1β: AMPK

Introduction to metabolic-inflammation

Early work suggested that sub-acute chronic inflammation that typified diet-related diseases including obesity, type-2 diabetes (T2D) and atherosclerosis disrupted metabolism, wherein immune cell mediators, particularly cytokines, impeded a range of metabolic pathways such as insulin signalling, glucose homoeostasis, lipid metabolism, reverse cholesterol transport, etc.^(1–3). The negative impact of inflammatory mediators on metabolism has been very well characterised and is relevant within a range of metabolic tissues, including adipose, skeletal muscle, liver, pancreas and the brain⁽⁴⁾. Thus sub-acute chronic inflammation is a classical hallmark of obesity, T2D and CVD (Fig. 1).

Interestingly more recent evidence shows that there is a much more dynamic reciprocal regulatory relationship between metabolism and inflammation, wherein the nature of cellular metabolism actually may determine immune cell functionality and thus inflammation. This review will explore the newer aspects of the interrelationships between metabolism and inflammation, with a view to understanding potential nutritional relevance. We will focus on IL-1 β mediated inflammation because of the strong inter-relationship between dietary fats, metabolic stressors and Nod-like receptor family, pyrin domain-containing three protein (NLRP3) inflammasome mediated inflammation. In terms of exploring the impact of dietary modulation of energy homoeostasis and metabolic-inflammation we will draw upon some simple examples to anticipate advances within this emerging and rapidly advancing field.

Metabolic stress and NLRP3 mediated IL-1ß inflammation

Metabolic stress, in the form of obesity, SFA, cholesterol, reactive oxygen species and/or uric acid, promote IL-1 β signalling⁽⁵⁾. It is now widely recognised that

Abbreviations: AMPK, AMP-activated protein kinase; HFD, high-fat diet; NLRP3, Nod-like receptor family, pyrin domain-containing three protein; OxPHOS, oxidative phosphorylation; PA, palmitic acid; T2D, type-2 diabetes; TLR, Toll-like receptor. Corresponding author: Helen Roche, email helen.roche@ucd.ie

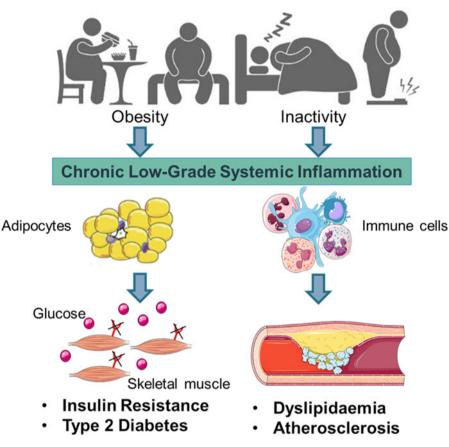


Fig. 1. (Colour online) Inflammation a classic hallmark of obesity, type-2 diabetes (T2D) and CVD. Excess energy intake and inactivity lead to the development of obesity, which is typified by sub-acute, chronic systemic inflammation. This pro-inflammatory phenotype develops in conjunction with expanding adipose tissue and immune cell activation. A range of immune cell mediators, including IL-1 β , disrupts adipogenesis, insulin signalling, glucose homoeostasis, lipid and cholesterol metabolism. Thus contributing to obesity related insulin resistance, T2D and atherosclerosis. Figure created using Servier Medical Art, https://smart.servier.com/

IL-1ß signalling plays a key role in obesity and T2D risk. Regulatory aspects of IL-1ß mediated inflammation are a very interesting aspect of metabolic-inflammation, by virtue of its two stage priming and activation process. As outlined in our recent reviews^(6,7), both stages of IL-1 β priming and NLRP3 mediated IL-1ß activation may be affected by the nutritional environment. As illustrated in Fig. 2, mature IL-1 β is produced and secreted by immune cells, following the initial process of priming, wherein the nascent form of pro-IL-1 β is formed, followed by the activation stage, wherein pro-IL-1 β is converted into mature IL-1ß by the NLRP3 inflammasome⁽⁸⁾. Structurally, the NLRP3 inflammasome is a multi-protein cytosolic complex composed of the regulatory subunit NLRP3, the effector subunit procaspase-1 and the adaptor molecule apoptosis-associated speck-like protein containing a caspase recruitment domain. Recruitment and activation of caspase-1 to the complex appears to be required for NLRP3 activation. Fig. 2 shows that both priming and activation stages of IL-1 β and NLRP3 biology are sensitive to nutritional, metabolic, as well as inflammatory, stress signals stemming from exogenous or endogenous sources, including fatty acids, glucose, reactive oxygen species, ATP and uric acid. For example, SFA induced Toll-like receptor (TLR)4 mediated metabolic stress, as well as inflammatory cytokines, prime pro-IL-1 β . Also in terms of the IL-1 β activation stage, a variety of metabolic factors, including SFA derivatives, ceramides, reactive oxygen species and ATP activate the NLRP3 inflammasome promoting the conversion of pro-IL-1 β into mature IL-1 β .

A substantial body of evidence has established that IL-1 β is key inflammatory signal involved in the development of obesity, insulin resistance and T2D⁽⁹⁾. IL-1 β disrupts cellular metabolism by interrupting a range of signalling pathways, including insulin sensitivity, lipid metabolism and adipogenesis^(10,11). We have demonstrated that IL-1R1 knockout (IL-1R1^{-/-}) mice are protected from high-fat diet (HFD)-induced insulin resistance, which was ascribed to attenuated adipose inflammation, early in life⁽¹¹⁾. Similarly, other groups have demonstrated that disrupting different components of the NLRP3 inflammasome complex, which regulates

Proceedings of the Nutrition Society

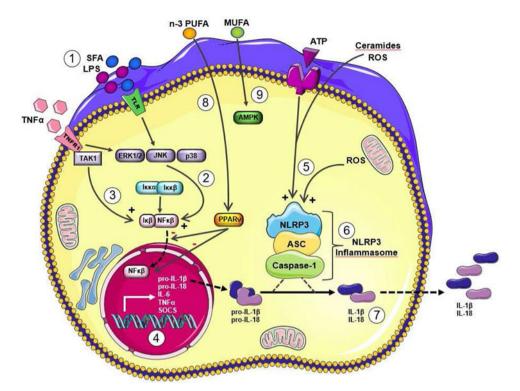


Fig. 2. (Colour online) Metabolic and inflammatory priming and activation of IL-1β via the Nod-like receptor family, pyrin domain-containing three protein (NLRP3) Inflammasome. Adapted from Ralston et al.⁽⁶⁾. Metabolic and inflammatory challenges that prime and activation IL-1β via NLRP3 inflammasome activation to modulate IL-1β secretion lipopolysaccharide (LPS) and SFA signal directly and indirectly via Toll-like receptor (TLR)4 (1) to activate stress kinases (2), which in turn activate NF- κ B with its translocation to the nucleus. TNF α also stimulates to NF- κ B activation (3). Upon translocation to the nucleus, NF-kB stimulates transcription of a range of inflammatory cytokines, including the nascent form of pro-IL-1 β (4). This stage represents the priming of pro-IL-1 β . Then the activation of pro-IL-1 β activation is dependent upon NLRP3 inflammasome activation. A wide range of metabolic stressor molecules including ATP, reactive oxygen species (ROS) and ceramides (derived from SFA) (5) have the ability to activate the NLRP3 inflammasome (6). A key stage of NLRP3 inflammasome activation includes the cleavage of pro-caspase-1 to active caspase-1. The resultant active IL-1ß is secreted from the cell (7). IL-18 has a very analogous biological pathway, wherein it is initially formed in a pro-IL-18 form and also requires NLRP3 mediated activation. Other dietary fatty acids may also modulate this process, for example MUFA maintain pAMPK activity, which in turn dampens NLRP3 inflammasome activation and thus reduces the IL-18/NLRP3 inflammasome mediated inflammatory response (8). Long chain n-3 PUFA may also attenuate these inflammatory pathways, possibly via PPARy activation, which in turn attenuates NF-κB activation (9). This figure was prepared using the Servier medical art website (http://www. servier.fr). ASC, apoptosis-associated speck-like protein containing a caspase-recruitment domain; TAK 1, transforming growth factor β activated kinase 1; ERK 1/2, extracellular signal-regulated kinase 1/2; JNK c-Jun N-terminal kinase; SOCS, suppressor of cytokine signaling.

IL-1 β activation, also protects against HFD-induced insulin resistance and inflammation^(12,13). Nevertheless the impact of NLRP3 mediated IL-1 β signalling on obesity and insulin sensitivity is probably more complex. Later in life IL-1R1^{-/-} mice ultimately develop mature-onset obesity, are profoundly insulin resistance in response to feeding HFD⁽¹⁴⁾. We speculate that IL-1 β biology may play a key role in adipogenesis which determines adipose morphology and functionality, which we discuss in the next section.

It is well accepted that SFA are potent pro-inflammatory fatty acids that promote obesity and insulin resistance. Initial lines of evidence suggested that SFA activate TLR, specifically TLR2 and TLR4⁽¹⁵⁾, which in turn activates the transcription factor NF- κ B, to promote and prime pro-IL-1 β production^(12,16). Interestingly more recent evidence shows that SFA are not in fact TLR4 ligands⁽¹⁷⁾. This very recent study showed that TLR4 is not a receptor for the common SFA palmitic acid (PA), which is derived from palm oil. PA does not bind to the TLR4 receptor to induce TLR4 dimerisation and activation of inflammation via NF- κ B. Instead PA mediates its pro-inflammatory effect by priming and re-configuring macrophage lipid metabolism, such that the cell is more responsive to SFA induced inflammatory and metabolic insults⁽¹⁷⁾.

Energy excess, metabolic-inflammation and adipose tissue health

Energy excess leads to obesity, with expansion of adipose tissue. Feeding HFD of course induce obesity, with concomitant sub-acute chronic inflammation and insulin resistance. However, it is important to appreciate that obese adipose tissue is a highly dynamic organ, not only reflecting energy storage potential. Adipose tissue expansion occurs through enlargement of adipocyte size (hypertrophy) or increased adipocyte number (hyperplasia). Interestingly there is a growing body of evidence that suggests that adipose morphology, hypertrophic v. hyperplasia, may determine adipose functionality and subsequent risk of T2D. Loss of white adipose tissue hyperplastic potential, resulting in hypertrophic adipose, is associated with increasing insulin concentrations and greater risk of insulin resistance and developing $T2D^{(18,19)}$. Therefore there has been great interest in determining the extent to which adipose tissue can be re-programmed such that its functional capacity is optimised to preserve insulin sensitivity,

attenuate T2D risk, despite obesity. Metabolic-inflammation plays a key role in adipogenesis. Adipose morphology reflects complex interplay between adipocyte programming and immune cell activation in adipose tissue. Dietary/metabolic stressors, including SFA prime pro-IL-1 β , which is then activated by the NLRP3 inflammasome complex. Several groups have shown that many cytokines, including IL-1B, potently impedes adipogenesis and insulin sensitivity in vitro⁽¹⁰⁾. It is possible that the induction of IL-1 β associated with feeding a HFD may prevent preadipocyte maturation and promote hyperplastic adipose morphology that typifies HFD-induced obesity. In contrast, mice lacking IL-1R1 signalling exhibit hyperplastic adipose morphology, with attenuated adipose inflammation and insulin resistance, despite obesity, in response to SFA-enriched HFD⁽¹⁴⁾. However the adipogenic potential of adipose tissue is lost with age and indeed the protective effect afforded by lack of IL-1ß signalling to preserve hyperplastic adipose morphology was also lost with age.

Interestingly, our recent work showed that obesogenic diets with different fatty acid composition induce different adipose morphology⁽²⁰⁾. Feeding SFA-rich HFD induced hypertrophic adipose morphology with greater NLRP3 mediated inflammation in adipose tissue concomitant with insulin resistance. In contrast, feeding a MUFA HFD was associated with a hyperplastic adipose morphology coincident with improved insulin sensitivity, despite equal adipose tissue weight gain. Detailed analysis of the adipogenic potential of the adipose stroma vascular fraction revealed that the MUFA HFD stroma vascular fraction had greater adipogenic potential, the potential of which was ascribed to lower NLRP3 mediated IL-1ß inflammation. This is only one potential example, it is likely that there is more to understand in relation to if and/how nutritional factors may modulate the metabolic-inflammation within the context of adipogenesis, which in turn may determine the physiological, metabolic and health impact of obesity.

Cellular energy homoeostasis and metabolic-inflammation

In the past number of years, the immunology field has demonstrated that metabolic reprogramming defines immune cell functionality⁽²¹⁾, as opposed to the initial understanding that inflammation disrupted metabolism. At the simplest level, cellular energy balance wherein cells rely upon oxidative phosphorylation (OxPHOS) or glucose dependent glycolytic metabolism to generate cellular ATP, defines the functional nature of immune cells and their inflammatory response. The metabolic pathway selected reflects the requirements and functionality of the immune cell. Activated pro-inflammatory immune cells require a shift wherein ATP is generated from anaerobic glycolysis, rather than deriving cellular energy from more efficient OxPHOS processes utilising the tricarboxylic acid cycle^(21,22). Jha et al. completed a very elegant combined transcriptomic and metabolomic analysis of the macrophage polarisation process⁽²¹⁾. Briefly, this systems approach showed that pro-inflammatory macrophages are less reliant upon mitochondrial OxPHOS, due to a disrupted tricarboxylic acid cycle, but depend upon glycolvsis as an ATP energy source. In contrast, antiinflammatory M2 macrophage or alternatively activated macrophage, are almost completely reliant upon OxPHOS with minimal glycolytic rates. Interestingly the nature of the energy substrates, glucose, fatty acids or amino acids, may affect the immune $response^{(23)}$. For example, there is an interesting switch in fatty acid metabolism, according to macrophage polarisation state. The ability and requirement to synthesise fatty acids, and probably also cholesterol, is a requirement of classically activated, pro-inflammatory M1 macrophage⁽²⁴⁾. In contrast, mitochondrial fatty acid β-oxidation is the predominant fuel source used by M2 macrophages.

A range of studies have demonstrated that disrupting different points within metabolic pathways can alter the nature of the inflammatory response. For example, impeding macrophage lipid metabolism by inhibiting fatty acid synthase, which catalyses the endogenous synthesis of SFA, suppresses the pro-inflammatory response, via NLRP3-inflammasome mediated IL-1 β activa-tion⁽²⁵⁾. In contrast, inhibiting glycolysis via pyruvate kinase inhibition promotes resolution of inflammation⁽²⁶⁾. These represent only two of several other points in metabolism that can be modulated, in order to affect the inflammatory response. Furthermore, it is also evident that metabolites, as well as metabolic derivatives of altered metabolism, can also re-configure the nature of the inflammatory phenotype. For example, the classical inflammatory insult, lipopolysaccharide which disrupts the tricarboxylic acid cycle, leads to the production of tricarboxylic acid intermediates, including citrate and succinate, which in turn play a functional biological role⁽²⁷⁾. Citrate promotes lipogenesis and is pro-inflammatory⁽²⁸⁾. Succinate acts as a pro-inflammatory metabolic signal by increasing Hif-1 α dependent IL-1 β expression⁽²⁹⁾. The above examples represent a brief snapshot wherein immune cells utilise different energy

substrates and metabolic pathways depending on their pro-inflammatory, anti-inflammatory or resolving role.

Dietary modulation of metabolic-inflammation

Going forward, it will be interesting to determine the extent to which dietary fatty acids, carbohydrate and protein sources may modulate the above processes given that the diet is an important exogenous source of some endogenous metabolites and/or substrates for the metabolic pathways eluded to above. Essentially the extent to which diet can (or cannot) re-programme metabolicinflammation in adipose tissue is relatively unknown. Kratz *et al.* demonstrated that metabolic insults such as the SFA PA, glucose and insulin; induced more complex inflammatory-metabolic phenotypes in adipose tissue macrophage compared to the classical lipopolysaccharide agonist, with greater lipid transport protein configuration in association with IL-1 β activation⁽³⁰⁾. Very recently, Febbraio's team confirmed that PA induces distinct inflammatory pathways different to lipopolysaccharide, not as a TLR4 agonist, but through TLR4 priming and altered macrophage lipid metabolism⁽¹⁷⁾. Thus metabolic re-programming is required for SFA induced inflammation in vitro. It will be interesting over the next number of years to determine the extent to which this paradigm hold true in vivo and also translates to man.

In a recent study we investigated the impact of different dietary fatty acids on metabolic-inflammation in an obese setting⁽²⁰⁾. This work provided interesting data with respect to the interaction and reciprocal regulation of NLRP3 mediated IL-1B activation and AMP-activated protein kinase (AMPK). Targeting AMPK mediated immunometabolism in obesity is attractive because it is a critical hub of both metabolism and IL-1 β inflammation⁽³¹⁾. AMPK is primarily known for its critical role in cellular energy homoeostasis, regulating metabolism⁽³¹⁾. AMPK activation promotes glucose-sparing, OxPHOS to generate maximal ATP from cellular energy substrates, a process utilised by most quiescent immune cells. This is converse to the rapid glucose uptake and glycolysis typical of activated immune cells. On the inflammatory side, AMPK activation attenuates IL-1ß inflammation via NLRP3^(23,32). Briefly, we showed that feeding a SFA enriched HFD to induce obesity was associated with significant NLRP3 mediated IL-16 inflammation, insulin resistance and adipose hypertrophy⁽²⁰⁾. In contrast, a MUFA-enriched HFD induced equivalent obesity, but adipose IL-1ß inflammation and insulin resistance was markedly attenuated compared to the SFA-HFD group. Interestingly we observed that adipose AMPK expression was not reduced following the MUFA-HFD, a common phenomenon associated with obesity. Given AMPK dual role linking metabolism and inflammation, we explored whether the different fatty acids mediated their pro- v. antiinflammatory effects in an AMPK dependent fashion. Using a combination of AMPK activators, inhibitors and shRNA approaches we were able to show that the MUFA oleic acid not only failed to prime IL-1ß but also blocked NLRP3 mediated IL-1ß activation in an AMPK

dependent manner⁽²⁰⁾. This was opposed to the SFA PA which both primed pro-IL-1 β and promoted NLRP3-mediated activation, via down-regulation of AMPK.

Conclusions

Nutritional modulation of metabolic-inflammation is a fast evolving area. No doubt our understanding of the complex interactions between what were considered two independent pathways, but now very much inter-connected, will be advanced. The real challenge will be the extent to which these cellular processes translate to man, so that we can fully appreciate if/how/when dietary constituents either promote or attenuate metabolic-inflammation.

Acknowledgements

The author thanks Marcella O'Reilly who designed Fig. 1.

Financial Support

The research presented in this review work was supported by Science Foundation Ireland principal investigator award (11/PI/1119); Joint Programming Healthy Life for a Healthy Diet (JPI HDHL) funded EU Food Biomarkers Alliance 'FOODBALL' (14/JP-HDHL/ B3076) and the Irish Department of Agriculture, Food and the Marine, 'ImmunoMet - dietary manipulation of microbiota diversity for controlling immune function' (14/F/828) programmes.

Conflict of Interest

None.

Authorship

The author had sole responsibility for all aspects of preparation of this paper.

References

- 1. Gregor MF & Hotamisligil GS (2011) Inflammatory mechanisms in obesity. *Annu Rev Immunol* **29**, 415–445.
- Samuel VT & Shulman GI (2016) The pathogenesis of insulin resistance: integrating signaling pathways and substrate flux. J Clin Invest 126, 12–22.
- 3. O'Reilly M, Dillon E, Guo W, *et al.* (2016) High-density lipoprotein proteomic composition, and not efflux capacity, reflects differential modulation of reverse cholesterol transport by saturated and monounsaturated fat diets. *Circulation* **133**, 1838–1850.
- 4. Osborn O & Olefsky JM (2012) The cellular and signaling networks linking the immune system and metabolism in disease. *Nat Med* **18**, 363–374.
- 5. Tack CJ, Stienstra R, Joosten LA, *et al.* (2012) Inflammation links excess fat to insulin resistance:

NS Proceedings of the Nutrition Society

the role of the interleukin-1 family. *Immunol Rev* 249, 239–252.

- 6. Ralston JC, Lyons CL, Kennedy EB, *et al.* (2017) Fatty acids and NLRP3 inflammasome-mediated inflammation in metabolic tissues. *Annu Rev Nutr* **37**, 77–102.
- Kirwan AM, Lenighan YM, O'Reilly ME, et al. (2017) Nutritional modulation of metabolic inflammation. Biochem Soc Trans 45, 979–985.
- Netea MG, Nold-Petry CA, Nold MF, et al. (2009) Differential requirement for the activation of the inflammasome for processing and release of IL-1beta in monocytes and macrophages. *Blood* 113, 2324–2335.
- Stienstra R, Joosten LA, Koenen T, *et al.* (2010) The inflammasome-mediated caspase-1 activation controls adipocyte differentiation and insulin sensitivity. *Cell Metab* 12, 593–605.
- Lagathu C, Yvan-Charvet L, Bastard JP, et al. (2006) Long-term treatment with interleukin-1beta induces insulin resistance in murine and human adipocytes. *Diabetologia* 49, 2162–2173.
- 11. McGillicuddy FC, Harford KA, Reynolds CM, *et al.* (2011) Lack of interleukin-1 receptor I (IL-1RI) protects mice from high-fat diet-induced adipose tissue inflammation coincident with improved glucose homeostasis. *Diabetes* **60**, 1688–1698.
- Wen H, Gris D, Lei Y, et al. (2011) Fatty acid-induced NLRP3-ASC inflammasome activation interferes with insulin signaling. *Nat Immunol* 12, 408–415.
- 13. Stienstra R, van Diepen JA, Tack CJ, *et al.* (2011) Inflammasome is a central player in the induction of obesity and insulin resistance. *Proc Natl Acad Sci USA* **108**, 15324–15329.
- McGillicuddy FC, Reynolds CM, Finucane O, et al. (2013) Long-term exposure to a high-fat diet results in the development of glucose intolerance and insulin resistance in interleukin-1 receptor I-deficient mice. Am J Physiol Endocrinol Metab 305, E834–E844.
- Nguyen MT, Favelyukis S, Nguyen AK, *et al.* (2007) A subpopulation of macrophages infiltrates hypertrophic adipose tissue and is activated by free fatty acids via Toll-like receptors 2 and 4 and JNK-dependent pathways. *J Biol Chem* 282, 35279–35292.
- 16. Reynolds CM, McGillicuddy FC, Harford KA, *et al.* (2012) Dietary saturated fatty acids prime the NLRP3 inflammasome via TLR4 in dendritic cells-implications for diet-induced insulin resistance. *Mol Nutr Food Res* **56**, 1212–1222.
- 17. Lancaster GI, Langley KG, Berglund NA, *et al.* (2018) Evidence that TLR4 is not a receptor for saturated fatty acids but mediates lipid-induced inflammation by reprogramming macrophage metabolism. *Cell Metab* 27, 1096–110.e5.

- Arner E, Westermark PO, Spalding KL, et al. (2010) Adipocyte turnover: relevance to human adipose tissue morphology. *Diabetes* 59, 105–109.
- Kim SM, Lun M, Wang M, et al. (2014) Loss of white adipose hyperplastic potential is associated with enhanced susceptibility to insulin resistance. *Cell Metab* 20, 1049–1058.
- Finucane OM, Lyons CL, Murphy AM, et al. (2015) Monounsaturated fatty acid-enriched high-fat diets impede adipose NLRP3 inflammasome-mediated IL-1beta secretion and insulin resistance despite obesity. *Diabetes* 64, 2116– 2128.
- Jha AK, Huang SC, Sergushichev A, et al. (2015) Network integration of parallel metabolic and transcriptional data reveals metabolic modules that regulate macrophage polarization. *Immunity* 42, 419–430.
- MacIver NJ, Michalek RD & Rathmell JC (2013) Metabolic regulation of T lymphocytes. Annu Rev Immunol 31, 259–283.
- O'Neill LA & Hardie DG (2013) Metabolism of inflammation limited by AMPK and pseudo-starvation. *Nature* 493, 346–355.
- Carroll RG, Zaslona Z, Galvan-Pena S, et al. (2018) An unexpected link between fatty acid synthase and cholesterol synthesis in proinflammatory macrophage activation. J Biol Chem 293, 5509–5521.
- Moon JS, Nakahira K & Choi AMK (2015) Fatty acid synthesis and NLRP3-inflammasome. *Oncotarget* 6, 21765– 21766.
- Palsson-McDermott EM, Curtis AM, Goel G, et al. (2015) Pyruvate kinase M2 regulates Hif-1alpha activity and IL-1beta induction and is a critical determinant of the warburg effect in LPS-activated macrophages. Cell Metab 21, 65–80.
- Williams NC & O'Neill LAJ (2018) A role for the Krebs cycle intermediate citrate in metabolic reprogramming in innate immunity and inflammation. *Front Immunol* 9, 141.
- Iacobazzi V & Infantino V (2014) Citrate new functions for an old metabolite. *Biol Chem* 395, 387–399.
- 29. Tannahill GM, Curtis AM, Adamik J, *et al.* (2013) Succinate is an inflammatory signal that induces IL-1beta through HIF-1alpha. *Nature* **496**, 238–242.
- Kratz M, Coats BR, Hisert KB, et al. (2014) Metabolic dysfunction drives a mechanistically distinct proinflammatory phenotype in adipose tissue macrophages. Cell Metab 20, 614–625.
- Herzig S & Shaw RJ (2018) AMPK: guardian of metabolism and mitochondrial homeostasis. *Nat Rev Mol Cell Biol* 19, 121–135.
- 32. Jeong HW, Hsu KC, Lee JW, *et al.* (2009) Berberine suppresses proinflammatory responses through AMPK activation in macrophages. *Am J Physiol Endocrinol Metab* **296**, E955–E964.