A Meeting of the Nutrition Society, hosted by the Scottish Section was held at King's College Conference Centre, University of Aberdeen on 26–27 March 2012

Conference on 'Future food and health' Symposium 2: Diet–gene interactions; implications for future diets and health

Epigenetic consequences of a changing human diet

Paul Haggarty

Division of Lifelong Health, Rowett Institute of Nutrition and Health, University of Aberdeen, Greenburn Road, Bucksburn, Aberdeen AB21 9SB, UK

> The human diet has undergone profound changes over recent generations and this trend is likely to accelerate in the 21st century. Innovations in food technology, new ways of producing and processing foods and the increasing use of artificial vitamins and novel ingredients are changing the human diet in ways that our dietary monitoring systems struggle to keep pace with. There is a growing awareness of the importance of diet, but little understanding of how these changes may affect the health of current and future generations. Epigenetic programming, and specifically the persistence of functional epigenetic states following nutritional exposure, is particularly relevant to the issue of dietary change. Epigenetics is emerging as perhaps the most important mechanism through which diet and nutrition can directly influence the genome and there is now considerable evidence for nutritional epigenetic programming of health and the response to diet itself. A number of nutrients and food components that are changing in the human diet have been shown to produce epigenetic states that are stable across different timescales. We need to better understand the nutritional programming of epigenetic states, the persistence of these marks in time and their effect on biological function and the response to diet.

> > Fortification: Novel foods: Supplements: Methylation: Programming

The changing human diet

The human diet has undergone profound changes throughout human history and pre-history. The change in nutrient availability made possible by the use of fire in the preparation of food was so great that it has been proposed as a key driver for the evolution of the human brain and intelligence. Over the past 10000 years the human diet has changed significantly as mankind has moved from a hunter-gatherer subsistence to that of an agriculturalist⁽¹⁾. The astonishing pace of change in the development of global agriculture and food distribution systems over the past century has resulted in further changes to the human diet. In the USA in the 20th century, intakes of added oils, shortening, meat, cheese and frozen dairy products have increased significantly, with more recent increases in added sweeteners, fruit, fruit juices and vegetables⁽²⁾.

This has resulted in important changes in the intakes of individual nutrients; for example the ratio of *n*-6 linoleic acid to *n*-3 linolenic acid has increased while the intakes of total *n*-3 and *n*-6 long-chain polyunsaturates as a per cent of energy have fallen⁽³⁾. Rapid changes have also occurred in the diet in the UK over the past century (Fig. 1). Since 1940, when reliable figures were first collected, there has been a modest increase in the intake of meat but significant changes in the type of meat consumed, with a reduction in mutton and lamb and an increase in poultry⁽⁴⁾. Over the same period vegetable consumption has decreased, whereas fruit consumption has increased. These and other changes in food consumption patterns have resulted in significant changes in the UK during the 20th century.

More recent advances have increased the potential for deliberate modification of the nutritional composition

Corresponding author: Professor P. Haggarty, fax +44 1224 438433, email p.haggarty@abdn.ac.uk

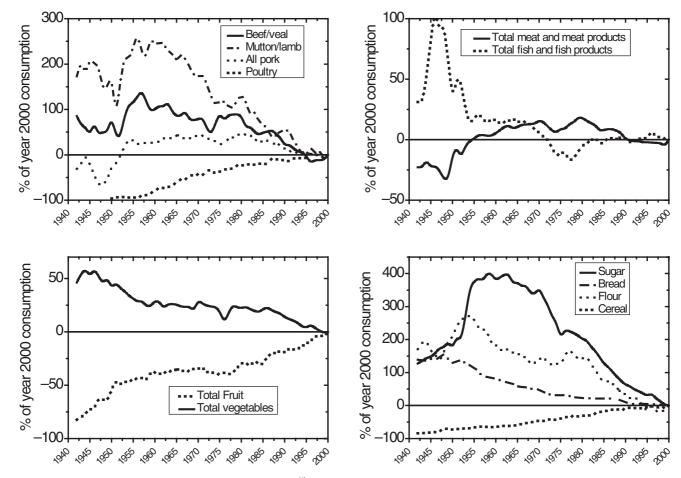


Fig. 1. Time trends in food intakes in the UK since 1940⁽⁴⁾. Data are presented as the percentage difference from intakes in the year 2000.

of the human diet. This trend is driven partly by pragmatism on the part of food producers and processors - efficiency and cost effectiveness of production; and partly by the desire to make claims, explicit and implicit, for the beneficial effects of foods and food products. These different goals tend to lead to socioeconomic stratification; highly processed low-cost foods v. high-value premium products defined by health claims. Innovations in food technology, new ways of producing and processing foods and the increasing use of artificial vitamins and novel ingredients are changing the human diet in ways that our dietary monitoring systems struggle to keep pace with. Since our nutritional monitoring systems are largely based on the estimated intakes of foods and food groups, they have a limited ability to detect changes in nutrient intake as a result of changes to the composition of foods. There is some monitoring of blood samples for nutrient status in national surveys, such as the National Health and Nutrition Examination Survey in the USA and the National Diet and Nutrition Survey in the UK, but the range of nutrients measured is relatively limited, the time spans over which change is measured are modest, and the changing methodologies used to measure nutrient status complicate the interpretation of trend data.

Technologies such as genetic modification have the potential to further increase the artificiality of the human diet. Such technologies are currently not allowed in Europe for foods destined for human consumption but the pressure is increasing to allow these technologies in order to alleviate food supply problems that will develop as a result of the linked issues of human population growth and climate change. An example of this dilemma is provided by 'Golden Rice'. This variety of rice has been genetically engineered to produce β -carotene to help combat vitamin A deficiency; a significant public health problem worldwide, particularly in developing countries⁽⁵⁾. This rice has recently been modified to further increase the content of β -carotene⁽⁶⁾. β -Carotene is a naturally occurring nutrient and while this food could help to reduce the prevalence of vitamin A deficiency worldwide there have been concerns over the potential for β -carotene to promote the development of lung cancer among high-risk individuals such as smokers and asbestos workers⁽⁷⁾.

Dietary supplements also have the potential to profoundly influence the intake of individual nutrients and there has been a steady increase in the use of supplements in industrialised countries over recent decades. In the USA, National Health and Nutrition Examination

364

NS Proceedings of the Nutrition Society

Survey II estimated supplement use at 28% among men and 38% among women aged 20 years and over in the period 1971-1975⁽⁸⁾. By 1976-1980 it was 32% among men and 43% among women in the same age group. By 1988-1994 over 40% of adults were using one or more dietary supplement and by 2003-2006 over 50% of adults were using supplements⁽⁸⁾. Dietary supplements can contain nutrients in amounts as high as or higher than the Institute of Medicine's Recommended Dietary Reference Intakes, and it is acknowledged that they may already be contributing substantially to total nutrient intake⁽⁸⁾. Multivitamins and multiminerals were the most frequently reported dietary supplement across all National Health and Nutrition Examination Survey years and the effect of supplements on total intake is likely to span a broad spectrum of nutrients. The UK is slightly behind the USA in terms of supplement use, but the trend is in the same direction. In the UK, it is estimated that 40% of women and 29% of men take dietary supplements⁽⁹⁾. There is also social stratification in supplement use in the UK with a higher frequency of supplement use in non-manual than manual groups. Cod liver oil and other fish-based supplements were the most commonly consumed supplements. Multivitamins and multiminerals were taken by 35% of those taking supplements with 12% taking multivitamins with no minerals, and the same proportion taking minerals with no vitamins⁽⁹⁾.

The nutrient content of foods may also be manipulated for a variety of reasons. Efforts to improve population health through diet have led to the introduction of a number of regulatory measures over the past century, which have required the addition of individual nutrients to certain foods. 'Restoration' is the term applied to the addition of nutrients to foods to ensure that any nutritional losses during storage, handling and manufacturing are made good. An example of this in the UK is the requirement that white and brown flour, unlike wholemeal flour, must be fortified with thiamin, niacin, calcium and iron. 'Substitution' is relevant to the production of substitute foods. An example of this is the substitution of margarine for butter and it has been a legal requirement in the UK since 1967 to fortify margarine with vitamins A and D, so that the levels are comparable with butter. 'Fortification' refers to the addition of vitamins or minerals irrespective of whether these nutrients were present originally and this may be mandatory or voluntary. An example of the former is the introduction in the USA, Canada and a number of other countries, of mandatory fortification of enriched grain products with folic acid. This is one of the most significant public health nutrition measures to be enacted in recent decades. Folic acid consumption by women is known to reduce the risk of neural tube defect in pregnancy and women who intend to become pregnant are currently recommended to take folic acid supplements periconceptionally and up until 12 weeks of gestation to reduce the risk of neural tube defect. However, many pregnancies are unplanned and fortification was introduced in order to reduce the incidence of neural tube defect in these pregnancies and in those groups

who were not following the advice on supplement use. This measure produced significant changes in population folate status. The introduction of mandatory fortification in the USA was completed in early 1998 and resulted in an estimated 215-240 µg/d increase in the intake of folates⁽¹⁰⁾ and a 144% increase in plasma folate concentration in the female population^(f1). Nutrients such as folic acid are also added, on a voluntary basis, to a wide range of foods such as breakfast cereals, spreads and a number of other product groups. When consumed in combination, or individually in large amounts, this can result in very high intakes. In some cases, the intake may exceed the upper level of folic acid intake considered to be safe $(1 \text{ mg/d for adults})^{(12)}$. In 2006 the UK Scientific Advisory Committee on Nutrition estimated that approximately 127000 people in the UK exceeded the upper limit for folic acid and 86% of these excess levels were attributed to consumption of foods, largely fat spreads and breakfast cereals, voluntarily fortified with folic $acid^{(12)}$.

Most consumers can exercise choice in relation to the foods they consume but increasing sophistication of food manufacture and processing may actually reduce our ability in practice to regulate our dietary intake of nutrients. It is difficult, but feasible, to optimise a diet for a few nutrients (e.g. in an effort to follow current recommendations on saturated fat, salt and energy) but the widespread addition of nutrients and novel ingredients to foods makes the process of product selection a formidable mathematical optimisation task, which may well be impractical for most consumers. Even if individuals were prepared to spend time attempting this, the addition of individual nutrients and novel ingredients to foods in which they do not normally occur means that every single product has to be checked for a large number of nutrients. In addition, many in society (e.g. those in schools, care homes, hospitals, prisons, or even those who choose not to prepare their own food) have very little control over the products they eat. The net result is that in practice, consumers have only a limited ability to resist industry driven changes in the nutrient composition of foods even if they wished to.

The nutrient composition of the human diet varies between populations and ethnic groups, and across geographical regions. Mankind has been able to adapt to these diets over time but the current pace and nature of the change in the human diet is new. The dietary changes listed earlier, together with technological manipulation of foods and the increasing use of nutritional supplements, are resulting in mixtures of nutrients never before experienced in human evolutionary history and this trend to artificiality shows every sign of accelerating in the 21st century. The 'single nutrient - deficiency symptom' model has historically been very helpful in alleviating nutritional problems but it has had little success in relation to the complex diseases, which are now the main concern in industrialised countries. There is a growing awareness of the importance of nutrient mixtures and recognition that it may actually be multiple nutrient exposure and nutrient interactions that are the key to health. Whatever the mechanism, we have little understanding of how the profound changes in nutrient intakes already observed will affect the health of current and future generations. Our emerging understanding of the field of epigenetics, and the way it is affected by nutrition, make it particularly relevant to this issue.

Epigenetics

Epigenetics is emerging as perhaps the most important mechanism through which the diet and nutrition can directly influence the genome⁽¹³⁾. This is not surprising as the two key groups involved in epigenetic modification of the histones and DNA (methyl and acetyl groups) are at the heart of nutritional metabolism. Numerous studies have demonstrated effects on DNA methylation of alcohol^(14–20), the B vitamins^(21–29), protein^(30–33), micronutrients^(34–37), functional food components^(38–42) and general nutritional status^(43–45) (Table 1).

At its most fundamental level, epigenetics is about information, and specifically the information present in the genome over and above that coded in the DNA sequence. This epigenetic information determines how, when and where the sequence information is $used^{(13)}$. Epigenetics is also about time and the way in which exposures can result in metastable epigenetic marks that persist for variable amounts of time (Fig. 2) and can influence biological function and health^(13,46-50). It is this aspect of epigenetics that makes it particularly relevant to the rapid pace of change in the human diet.

Much of the work on basic epigenetic mechanisms has focused on reproduction and this has led to a particular interest in the possibility that epigenetic status may be influenced by specific environmental factors such as nutrition in the critical period before birth, and even before conception. Epigenetics has been defined as 'heritable changes in gene function that cannot be explained by changes in DNA sequence'⁽⁵¹⁾ and many studies have been carried out in pregnancy in animal models^(15,25,26,32,33,36,39,45) and human subjects^(21,29,34,43,44) looking at the effect on epigenetic status in the offspring of nutritional exposures during pregnancy. Nutritional factors at key life stages can result in relatively stable epigenetic marks that persist over decades, or even more than one lifetime, and have functional consequences for health. Most of the pregnancy studies have investigated nutrient exposure in mothers but transgenerational epigenetic programming is also relevant to fathers, and the nutrients they consume during the epigenetic programming of the sperm that provide one half of the DNA of the offspring^(20,52).

Epigenetic marking that is particularly relevant to the changing nutritional environment occurs in imprinted genes and the repeat elements. Imprinting refers to parent of origin specific regulation of gene expression^(53–55). The imprint is set early in development and passed down through the somatic cell lineage^(53–55). Some imprinted regions remain stable over decades^(56,57) but there is variation between individuals in the level of imprinting methylation^(29,56,58,59). This variation, and how it comes about, is of considerable interest as the process of imprinting, and imprinting status, is thought to be important in health and disease^(46,47). Human imprinting syndromes, where the normal process of imprinting is disrupted, result in a wide range of phenotypes^(60–62) including obesity⁽⁶³⁾ and diabetes⁽⁶⁴⁾. Loss of imprinting within the insulin-like growth factor 2 (IGF2) gene is characteristic of many cancers⁽⁶⁵⁾ and this even occurs in non-tumour tissue of individuals with cancer or at high risk of cancer⁽⁶⁵⁾.

Recent work from the encyclopaedia of DNA elements project have highlighted the importance of epigenetic control of the genome at large scale⁽⁶⁶⁾ and a large proportion (about 45%) of the genome is made up of repeat elements such as the long interspersed nuclear elements (LINE-1) and the short interspersed transposable nuclear elements (SINE), including the Alu family of human SINE elements^(49,50). These are frequently found in or near genes and the chromatin conformation formed at retrotransposons may spread and influence the transcription of nearby genes^(49,50). They can generate insertions, mutations and genomic instability and are responsible for sixty-five known genetic disorders^(49,50). Methylation has the effect of repressing transposition^(49,50). Like the imprinted genes, transposable elements are characterised by developmental stage dependent epigenetic marking and they are thought to play important roles in health and disease^(48–50). The epigenetic status of repeat elements such as intracisternal A particle (*IAP*) are resistant to reprogramming during primordial germ cell and pre-implantation development and this has been proposed as a mechanism by which epigenetic status may be passed between generations through the germline⁽⁶⁷⁾. Dietary intake of the phytoestrogen genisten during pregnancy in animals alters the methylation status of IAP and these changes appear to confer some protection against obesity in the offspring^(39,46).

The ultimate methyl donor for epigenetic-methylation reactions is S-adenosylmethionine that is produced by the methylation cycle and it has been reported that periconceptional folic acid use alters the level of methylation within IGF2⁽²¹⁾. A larger study of human pregnancy also observed an effect of folic acid use on IGF2 methylation in the offspring but the effect was restricted to folic acid use after 12 weeks gestation when women are not recommended to take the supplement⁽²⁹⁾. Late gestation use of folic acid was also associated with reduced LINE-1 methylation and altered paternally expressed gene 3 (PEG3) methylation. Three of the four significant associations with folic acid use and folate status were negative and one was positive, suggesting that it may be naive to assume that this is a simple substrate limitation effect or that the supply of nutrients involved in the methylation cycle will affect all genes equally. Imprinting occurs before fertilisation but changes in imprinting methylation in animal models in response to nutritional exposures have been demonstrated into the early post-natal period for IGF2, after which the imprint is apparently fixed⁽⁶⁸⁾.

366

Table 1. Evidence for nutritional effects on epigenetic stat	for nutritional effects on epigenetic status
---	--

Exposure	Epigenetic process affected	Species/model	Reference
General nutrition			
Famine	Offspring imprinting (IGF2) methylation in blood	Human (pregnancy)	(43)
Dietary vitamin B ₁₂ , betaine, choline, folate, cadmium, zinc and iron	Male LINE-1 methylation affected by choline intake in early pregnancy	Human (pregnancy)	(44)
Late gestation undernutrition	Sex-specific changes in imprinting in tissues	Mouse (pregnancy)	(45)
Protein			
Low protein diet and folic acid	Hepatic gene expression	Rat	(30)
Low protein diet and folic acid	DNA methylation in Imprinting Control Region of Igf2/H19. Expression of Dnmt1 and Dnmt3a, and methyl CpG-binding domain 2 (Mbd2)	Rat	(31)
Low protein diet	Methylation of PPAR α and glucocorticoid receptor in liver of offspring	Rat (pregnancy)	(32)
Low protein diet	Some evidence of effect on expression. Little effect on methylation levels of imprinted genes or $PPARa$ promoter and enhancer, and B1 repetitive elements in the liver of offspring	Mouse (pregnancy)	(33)
Micronutrients			
Multi-micronutrient supplements in The Gambia	Reduced methylation levels in IGF2R in girls and GTL2–2 in boys	Human (pregnancy)	(34)
Methyl deficient diets plus arsenic	Increased global DNA methylation in female livers and decreased global DNA methylation in male livers	Mouse	(35)
Choline	Igf2 hypermethylated in the liver	Rat (pregnancy)	(36)
Arsenic	Wide range of effects (review)	Various	(37)
B vitamins			
Periconceptional folic acid supplement use	Imprinting (IGF2) methylation in blood of children	Human (pregnancy)	(21)
Folic acid supplement use at different stages of pregnancy, dietary folate, folate cycle genotype	Imprinting (IGF2, PEG3, SNRPN) and retrotransposon (LINE-1) methylation in cord blood	Human (pregnancy)	(29)
Folate genetics; methylenetetrahydrofolate reductase polymorphism	Global DNA methylation	Human	(22,23)
Biotin	Binding to histones, influence on retrotransposons	Human and mouse cells	(24)
Methyl-donor deficiency (methionine, choline, and folic acid)	Global DNA hypermethylation in the brain and hypomethylation in the liver	Rat	(28)
Folic acid, choline, betaine	DNA methylation of a long terminal repeat controlling expression of the agouti gene	Mouse	(25,26)
Niacin	Chromatin structure and function	Various	(27)
Alcohol			
Alcohol	Mouse embryo DNA methylation and gene expression	Mouse	(14)
Alcohol	DNA methylation in foetus	Mouse (pregnancy)	(15)
Alcohol	Human DNA methylation in blood	Human	(16,17)
Alcohol	DNA methylation in colonic mucosa	Rat	(18)
Alcohol	Hepatic myelocytomatosis oncogene (<i>Myc</i>) hypomethylation. Altered expression of the methionine adenosyltransferases	Rat	(19)
Alcohol	Sperm cytosine methyltransferase messenger RNA levels	Rat	(20)
Functional components	· · · · · ·		
Sulforaphane (found in cruciferous vegetables)	Histone deacetylase	Various	(38)
Genistein (phytoestrogen)	Methylation of retrotransposon upstream of the transcription start site of the Agouti gene	Mouse (pregnancy)	(39)
Polyphenols (found in green tea, coffee and soyabean)	DNA methylation and DNA methyltransferase activity	Various	(40–42)

IGF, insulin-like growth factor 2; LINE-1, long interspersed nuclear elements; PEG3, paternally expressed gene 3; SNRPN, small nuclear ribonucleoprotein polypeptide N.

Imprinting requires removal of the epigenetic mark of the previous generation followed by sex-specific epigenetic marking in the gametes^(62,69,70) although it is thought that some repetitive elements, such as *LINE1* and *IAP*, may only be partially demethylated in the primordial germ cells⁽⁶²⁾. Such retention of epigenetic information could be one way in which maternal exposures during key stages of development result in epigenetic changes in the offspring. There are also differences in timing with some paternal alleles acquiring methylation before maternal alleles in the male germline and vice versa in the female germ $line^{(62)}$ and variation in

Proceedings of the Nutrition Society

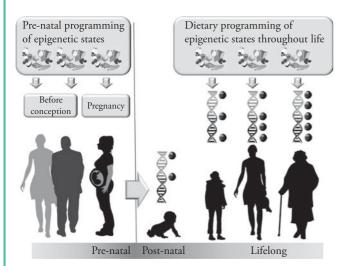


Fig. 2. Dietary programming of metastable functional epigenetic states before and after birth.

the process of imprinting by $gene^{(62,71)}$. There are also significant differences in the stage of development at which male and female gametes acquire imprints^(62,70,72). There are reports that the balance of maternal and paternal imprints in the offspring may have functional significance⁽⁷³⁾. The difference in the timing of maternal and paternal imprinting and other epigenetic processes in relation to life stage is another way in which changing nutritional exposures could influence biological function and health.

Life stage-specific epigenetic marking is not restricted to the period before birth. There is evidence that it changes with age across the life-course. There is a loss of global DNA methylation with $age^{(74,75)}$ and this is reflected in a fall in methylation in some repeat elements^(75–77) but not all⁽⁷⁷⁾. There are also reports of increases in CpG island methylation, and decreases in methylation in regions out with CpG islands, with age in solid tissues and blood-derived DNA⁽⁷⁸⁾ The picture in relation to individual genes is complicated⁽⁷⁹⁾, with some increasing^(75,80,81) and others decreasing⁽⁷⁶⁾ with age.

Implications

The human diet has undergone profound changes over recent generations and this trend is likely to accelerate in the 21st century. There is a growing awareness of the importance of diet and nutrition to human health but little understanding of how these temporal changes in diet are likely to affect the health of current and future generations. One problem is that our understanding of nutrient effects on health is largely based on observational studies in populations consuming diets representative of a particular time and location. Even intervention studies are carried out on a background intake of nutrients that may not be wholly relevant to future populations.

Epigenetic change has been demonstrated in response to a wide range of foods and nutrients and epigenetic status is emerging as a critical determinant of the response of the organism to the environment and its biological function and disease susceptibility. Dietary change may act directly on the epigenetic processes that result in health/disease but it can also programme metabolism and the future response to nutrition itself. There is a growing body of evidence, largely based on animal studies, demonstrating that nutritional exposures during particular life stages, and developmental windows, can influence epigenetic status, biology and physiology throughout life. Supporting evidence is also beginning to emerge from studies in human subjects.

Transgenerational programming is proposed to have developed in human subjects to confer flexibility of response to the environment: the hypothesis is that it allows the offspring genome to be optimally programmed in response to the maternal environment before birth to make it better fitted to respond metabolically to the environment it will experience. However, the profound dietary changes already occurring within less than a human life span, and the apparent acceleration of that change, mean that the nutritional environment experienced by the mother during pregnancy may not reflect the one in which the offspring will live. The concept of epigenetic programming is not only limited to the period before birth, it also applies to nutritional effects across the life course.

We need better monitoring of changing nutrient intakes in the population, particularly in vulnerable sub-groups, but the rapid pace of change in food reformulation, fortification and the increasing use of novel ingredients, presents a challenge to our current food based national monitoring systems. We need to understand better the consequences of intakes of novel mixtures of nutrients and their effect on health. Epigenetic programming, and specifically the concept of persistence of functional epigenetic states following a nutritional exposure, is particularly relevant to the issue of dietary change. We need to better understand the susceptibility of the genome to epigenetic marking, the critical temporal windows when this occurs, the persistence of these marks in time, and their effect on biological function and the response to diet.

Acknowledgements

The author is grateful to the Scottish Government (RESAS).

Financial Support

The Scottish Government (RESAS) provided support.

Conflicts of Interest

None.

Authorship

P. H. conceived the paper, carried out the analysis, and wrote the manuscript.

368

References

- Jew S, AbuMweis SS & Jones PJ (2009) Evolution of the human diet: linking our ancestral diet to modern functional foods as a means of chronic disease prevention. J Med Food 12, 925–934.
- 2. Barnard ND (2010) Trends in food availability, 1909–2007. *Am J Clin Nutr* **91**, 1530S–1536S.
- 3. Blasbalg TL, Hibbeln JR, Ramsden CE *et al.* (2011) Changes in consumption of omega-3 and omega-6 fatty acids in the United States during the 20th century. *Am J Clin Nutr* **93**, 950–962.
- DEFRA (2012) Family Food Datasets. Department for Environment Food and Rural Affairs; available at http:// archive.defra.gov.uk/evidence/statistics/foodfarm/food/ familyfood/.
- 5. Ye X, Al-Babili S, Kloti A *et al.* (2000) Engineering the provitamin A (beta-carotene) biosynthetic pathway into (carotenoid-free) rice endosperm. *Science* **287**, 303–305.
- 6. Paine JA, Shipton CA, Chaggar S *et al.* (2005) Improving the nutritional value of Golden Rice through increased pro-vitamin A content. *Nat Biotechnol* **23**, 482–487.
- Sommer A & Vyas KS (2012) A global clinical view on vitamin A and carotenoids. *Am J Clin Nutr* 96, 1204S– 1206S.
- Gahche J, Bailey R, Burt V, Hughes J, Yetley E, Dwyer J, Piccano MF, McDowell M & Sempos C (2011) *Dietary* Supplement use Among U.S. Adults has Increased Since NHANES III (1988–1994) no. 61. Hyattsville, MD: National Center for Health Statistics.
- 9. Henderson L, Gregory J & Swan G (2002) The National Diet and Nutrition Survey: Adults Aged 19 to 64 Years. Vol 1; Types and Quantities of Foods Consumed. London: HMSO.
- Quinlivan EP & Gregory JF III (2003) Effect of food fortification on folic acid intake in the United States. Am J Clin Nutr 77, 221–225.
- Pfeiffer CM, Caudill SP, Gunter EW et al. (2005) Biochemical indicators of B vitamin status in the US population after folic acid fortification: results from the National Health and Nutrition Examination Survey 1999–2000. Am J Clin Nutr 82, 442–450.
- 12. Scientific Advisory Committee on Nutrition (2006) Folate and Disease Prevention. London: TSO.
- 13. Haggarty P (2012) Nutrition and the epigenome. *Prog Mol Biol Transl Sci* 108, 427–446.
- 14. Liu Y, Balaraman Y, Wang G *et al.* (2009) Alcohol exposure alters DNA methylation profiles in mouse embryos at early neurulation. *Epigenetics* **4**, 500–511.
- Garro AJ, McBeth DL, Lima V et al. (1991) Ethanol consumption inhibits fetal DNA methylation in mice: implications for the fetal alcohol syndrome. Alcohol Clin Exp Res 15, 395–398.
- 16. Bonsch D, Lenz B, Reulbach U *et al.* (2004) Homocysteine associated genomic DNA hypermethylation in patients with chronic alcoholism. *J Neural Transm* **111**, 1611–1616.
- 17. Bonsch D, Lenz B, Kornhuber J *et al.* (2005) DNA hypermethylation of the alpha synuclein promoter in patients with alcoholism. *Neuroreport* **16**, 167–170.
- Choi SW, Stickel F, Baik HW *et al.* (1999) Chronic alcohol consumption induces genomic but not p53-specific DNA hypomethylation in rat colon. *J Nutr* **129**, 1945–1950.
- Lu SC, Huang ZZ, Yang H et al. (2000) Changes in methionine adenosyltransferase and S-adenosylmethionine homeostasis in alcoholic rat liver. Am J Physiol Gastrointest Liver Physiol 279, G178–G185.

- Bielawski DM, Zaher FM, Svinarich DM et al. (2002) Paternal alcohol exposure affects sperm cytosine methyltransferase messenger RNA levels. *Alcohol Clin Exp Res* 26, 347–351.
- Steegers-Theunissen RP, Obermann-Borst SA, Kremer D et al. (2009) Periconceptional maternal folic acid use of 400 microg per day is related to increased methylation of the IGF2 gene in the very young child. PLoS ONE 4, e7845.
- 22. Friso S, Choi SW, Girelli D et al. (2002) A common mutation in the 5,10-methylenetetrahydrofolate reductase gene affects genomic DNA methylation through an interaction with folate status. *Proc Natl Acad Sci USA* **99**, 5606–5611.
- 23. Stern LL, Mason JB, Selhub J et al. (2000) Genomic DNA hypomethylation, a characteristic of most cancers, is present in peripheral leukocytes of individuals who are homozygous for the C677T polymorphism in the methylenetetrahydrofolate reductase gene. Cancer Epidemiol Biomarkers Prev 9, 849–853.
- Zempleni J, Chew YC, Bao B *et al.* (2009) Repression of transposable elements by histone biotinylation. *J Nutr* 139, 2389–2392.
- 25. Cooney CA, Dave AA & Wolff GL (2002) Maternal methyl supplements in mice affect epigenetic variation and DNA methylation of offspring. *J Nutr* **132**, 23938–2400S.
- 26. Wolff GL, Kodell RL, Moore SR *et al.* (1998) Maternal epigenetics and methyl supplements affect agouti gene expression in Avy/a mice. *FASEB J* **12**, 949–957.
- Kirkland JB (2009) Niacin status impacts chromatin structure. J Nutr 139, 2397–2401.
- Pogribny IP, Karpf AR, James SR *et al.* (2008) Epigenetic alterations in the brains of Fisher 344 rats induced by longterm administration of folate/methyl-deficient diet. *Brain Res* 1237, 25–34.
- 29. Haggarty P, Hoad G, Campbell DM *et al.* (2013) Folate in pregnancy and imprinted gene and repeat element methylation in the offspring. *Am J Clin Nutr* **97**, 94–99.
- Lillycrop KA, Phillips ES, Jackson AA et al. (2005) Dietary protein restriction of pregnant rats induces and folic acid supplementation prevents epigenetic modification of hepatic gene expression in the offspring. J Nutr 135, 1382–1386.
- Gong L, Pan YX & Chen H (2010) Gestational low protein diet in the rat mediates IGF2 gene expression in male offspring via altered hepatic DNA methylation. *Epigenetics* 5, 619–626.
- 32. Burdge GC, Slater-Jefferies J, Torrens C *et al.* (2007) Dietary protein restriction of pregnant rats in the F0 generation induces altered methylation of hepatic gene promoters in the adult male offspring in the F1 and F2 generations. *Br J Nutr* **97**, 435–439.
- 33. Ivanova E, Chen JH, Segonds-Pichon A et al. (2012) DNA methylation at differentially methylated regions of imprinted genes is resistant to developmental programming by maternal nutrition. *Epigenetics* 7, 1200–1210.
- 34. Cooper WN, Khulan B, Owens S *et al.* (2012) DNA methylation profiling at imprinted loci after periconceptional micronutrient supplementation in humans: results of a pilot randomized controlled trial. *FASEB J* 26, 1782–1790.
- 35. Nohara K, Baba T, Murai H *et al.* (2011) Global DNA methylation in the mouse liver is affected by methyl deficiency and arsenic in a sex-dependent manner. *Arch Toxicol* **85**, 653–661.

P. Haggarty

- 36. Kovacheva VP, Mellott TJ, Davison JM *et al.* (2007) Gestational choline deficiency causes global and IGF2 gene DNA hypermethylation by up-regulation of Dnmt1 expression. *J Biol Chem* **282**, 31777–31788.
- 37. Reichard JF & Puga A (2010) Effects of arsenic exposure on DNA methylation and epigenetic gene regulation. *Epigenomics* **2**, 87–104.
- Ho E, Clarke JD & Dashwood RH (2009) Dietary sulforaphane, a histone deacetylase inhibitor for cancer prevention. J Nutr 139, 2393–2396.
- 39. Dolinoy DC, Weidman JR, Waterland RA *et al.* (2006) Maternal genistein alters coat color and protects Avy mouse offspring from obesity by modifying the fetal epigenome. *Environ Health Perspect* **114**, 567–572.
- Lee WJ & Zhu BT (2006) Inhibition of DNA methylation by caffeic acid and chlorogenic acid, two common catechol-containing coffee polyphenols. *Carcinogenesis* 27, 269–277.
- 41. Fang M, Chen D & Yang CS (2007) Dietary polyphenols may affect DNA methylation. *J Nutr* **137**, 223S–228S.
- 42. Fang MZ, Wang Y, Ai N *et al.* (2003) Tea polyphenol (–)-epigallocatechin-3-gallate inhibits DNA methyltransferase and reactivates methylation-silenced genes in cancer cell lines. *Cancer Res* **63**, 7563–7570.
- 43. Heijmans BT, Tobi EW, Stein AD *et al.* (2008) Persistent epigenetic differences associated with prenatal exposure to famine in humans. *Proc Natl Acad Sci USA* **105**, 17046–17049.
- 44. Boeke CE, Baccarelli A, Kleinman KP *et al.* (2012) Gestational intake of methyl donors and global LINE-1 DNA methylation in maternal and cord blood: prospective results from a folate-replete population. *Epigenetics* 7, 253–260.
- 45. Radford EJ, Isganaitis E, Jimenez-Chillaron J *et al.* (2012) An unbiased assessment of the role of imprinted genes in an intergenerational model of developmental programming. *PLoS Genet* **8**, e1002605.
- 46. Jirtle RL & Skinner MK (2007) Environmental epigenomics and disease susceptibility. *Nat Rev Genet* **8**, 253–262.
- 47. Waterland RA & Jirtle RL (2004) Early nutrition, epigenetic changes at transposons and imprinted genes, and enhanced susceptibility to adult chronic diseases. *Nutrition* **20**, 63–68.
- 48. Walter J, Hutter B, Khare T *et al.* (2006) Repetitive elements in imprinted genes. *Cytogenet Genome Res* **113**, 109–115.
- 49. Levin HL & Moran JV (2011) Dynamic interactions between transposable elements and their hosts. *Nat Rev Genet* **12**, 615–627.
- 50. Cordaux R & Batzer MA (2009) The impact of retrotransposons on human genome evolution. *Nat Rev Genet* **10**, 691–703.
- 51. Russo VEA, Martienssen RA & Riggs AD (1996) Epigenetic Mechanisms of Gene Regulation. Woodbury: Cold Spring Harbor Laboratory Press.
- 52. Ferguson-Smith AC & Patti ME (2011) You are what your dad ate. *Cell Metab* **13**, 115–117.
- 53. Ferguson-Smith AC & Surani MA (2001) Imprinting and the epigenetic asymmetry between parental genomes. *Science* **293**, 1086–1089.
- 54. Ferguson-Smith AC (2011) Genomic imprinting: the emergence of an epigenetic paradigm. *Nat Rev Genet* 12, 565–575.
- 55. Reik W & Walter J (2001) Genomic imprinting: parental influence on the genome. *Nat Rev Genet* **2**, 21–32.

- 56. Sandovici I, Leppert M, Hawk PR *et al.* (2003) Familial aggregation of abnormal methylation of parental alleles at the IGF2/H19 and IGF2R differentially methylated regions. *Hum Mol Genet* **12**, 1569–1578.
- 57. Woodfine K, Huddleston JE & Murrell A (2011) Quantitative analysis of DNA methylation at all human imprinted regions reveals preservation of epigenetic stability in adult somatic tissue. *Epigenetics Chromatin* 4, 1.
- 58. Heijmans BT, Kremer D, Tobi EW *et al.* (2007) Heritable rather than age-related environmental and stochastic factors dominate variation in DNA methylation of the human IGF2/H19 locus. *Hum Mol Genet* **16**, 547– 554.
- 59. Kaminsky ZA, Tang T, Wang SC *et al.* (2009) DNA methylation profiles in monozygotic and dizygotic twins. *Nat Genet* **41**, 240–245.
- Owen CM & Segars JH Jr. (2009) Imprinting disorders and assisted reproductive technology. *Semin Reprod Med* 27, 417–428.
- 61. Robertson KD (2005) DNA methylation and human disease. *Nat Rev Genet* 6, 597–610.
- 62. Trasler JM (2006) Gamete imprinting: setting epigenetic patterns for the next generation. *Reprod Fertil Dev* 18, 63–69.
- 63. Horsthemke B & Buiting K (2008) Chapter 8 genomic imprinting and imprinting defects in humans. *Adv Genet* 61, 225–246.
- 64. Temple IK & Shield JP (2002) Transient neonatal diabetes, a disorder of imprinting. J Med Genet **39**, 872–875.
- 65. Feinberg AP, Ohlsson R & Henikoff S (2006) The epigenetic progenitor origin of human cancer. *Nat Rev Genet* 7, 21–33.
- 66. Bernstein BE, Birney E, Dunham I *et al.* (2012) An integrated encyclopedia of DNA elements in the human genome. *Nature* **489**, 57–74.
- 67. Reik W (2007) Stability and flexibility of epigenetic gene regulation in mammalian development. *Nature* 447, 425–432.
- Waterland RA, Lin JR, Smith CA *et al.* (2006) Post-weaning diet affects genomic imprinting at the insulinlike growth factor 2 (IGF2) locus. *Hum Mol Genet* 15, 705– 716.
- 69. Allegrucci C, Thurston A, Lucas E *et al.* (2005) Epigenetics and the germline. *Reproduction* **129**, 137–149.
- Sasaki H & Matsui Y (2008) Epigenetic events in mammalian germ-cell development: reprogramming and beyond. *Nat Rev Genet* 9, 129–140.
- 71. Lucifero D, Mertineit C, Clarke HJ *et al.* (2002) Methylation dynamics of imprinted genes in mouse germ cells. *Genomics* **79**, 530–538.
- 72. Kerjean A, Dupont JM, Vasseur C *et al.* (2000) Establishment of the paternal methylation imprint of the human H19 and MEST/PEG1 genes during spermatogenesis. *Hum Mol Genet* **9**, 2183–2187.
- 73. Badcock C & Crespi B (2006) Imbalanced genomic imprinting in brain development: an evolutionary basis for the aetiology of autism. *J Evol Biol* **19**, 1007–1032.
- 74. Fraga MF, Agrelo R & Esteller M (2007) Cross-talk between aging and cancer: the epigenetic language. Ann N Y Acad Sci 1100, 60–74.
- 75. Gentilini D, Mari D, Castaldi D *et al.* (2012) Role of epigenetics in human aging and longevity: genome-wide DNA methylation profile in centenarians and centenarians' offspring. *Age (Dordr)* [Epublication ahead of print].



- 76. Heyn H, Li N, Ferreira HJ et al. (2012) Distinct DNA methylomes of newborns and centenarians. Proc Natl Acad Sci USA 109, 10522–10527.
- 77. Jintaridth P & Mutirangura A (2010) Distinctive patterns of age-dependent hypomethylation in interspersed repetitive sequences. *Physiol Genomics* **41**, 194–200.
- Christensen BC, Houseman EA, Marsit CJ et al. (2009) Aging and environmental exposures alter tissue-specific DNA methylation dependent upon CpG island context. PLoS Genet 5, e1000602.
- 79. Madrigano J, Baccarelli A, Mittleman MA *et al.* (2012) Aging and epigenetics: longitudinal changes in genespecific DNA methylation. *Epigenetics* **7**, 63–70.
- Post WS, Goldschmidt-Clermont PJ, Wilhide CC *et al.* (1999) Methylation of the estrogen receptor gene is associated with aging and atherosclerosis in the cardiovascular system. *Cardiovasc Res* 43, 985–991.
- Kwabi-Addo B, Chung W, Shen L *et al.* (2007) Age-related DNA methylation changes in normal human prostate tissues. *Clin Cancer Res* 13, 3796–3802.