Review Article

Role of sulfur amino acids in controlling nutrient metabolism and cell functions: implications for nutrition

Sophie Tesseraud¹*, Sonia Métayer Coustard¹, Anne Collin¹ and Iban Seiliez²

¹ INRA, UR83 Recherches Avicoles, F-37380 Nouzilly, France

² INRA, UMR1067 Nutrition Aquaculture et Génomique, F-64310 St Pée-sur-Nivelle, France

(Received 18 June 2008 - Revised 27 October 2008 - Accepted 28 October 2008 - First published online 15 December 2008)

Protein synthesis is affected when an insufficient level of sulfur amino acids is available. This defect may originate from dietary amino acid deficiency and/or excessive amino acid utilisation for other purposes such as the synthesis of glutathione and acute-phase proteins during catabolic stress. Sulfur amino acids are recognised to exert other significant functions since they are precursors of essential molecules, are involved in the methylation process, participate in the control of oxidative status, and may act as mediators affecting metabolism and cell functions. Despite this increased understanding of the role of sulfur amino acids, many questions still remain unanswered due to the complexity of the mechanisms involved. Moreover, surprising effects of dietary sulfur amino acids have been reported, with the development of disorders in cases of both deficiency and excess. These findings indicate the importance of defining adequate levels of intake and providing a rationale for nutritional advice. The aim of the present review is to provide an overview on the roles of sulfur amino acids as regulators of nutrient metabolism and cell functions, with emphasis placed on the implications for nutrition.

Methionine: Cysteine: Nutrient metabolism: Oxidative stress: Methylation process: Gene expression

Like other amino acids, sulfur amino acids affect protein metabolism. They are the components of tissue proteins and, when provided at insufficient levels, they lead to reduced protein synthesis. Methionine is, for example, the first limiting factor in classical diets used for growing chickens because of the high requirement of sulfur amino acids for the synthesis of feathers, whereas poultry diets based on maize and soyabean meal without supplementation are deficient in sulfur amino acids⁽¹⁾. A positive effect of methionine supplementation on muscle growth has been reported in this species, since the addition of methionine to a methionine-deficient diet that is otherwise balanced in terms of other amino acids increases the accretion and synthesis of protein in skeletal muscles. Like methionine, cysteine is used for protein synthesis. Cysteine can be produced through the metabolism of methionine (Fig. 1) and can thus be considered as a non-essential amino acid. It can become conditionally indispensable in particular situations such as stress conditions or inflammatory states. In such situations, cysteine may be used for acute-phase proteins but above all for the synthesis of glutathione, i.e. the most important intracellular antioxidant of the body. This results in an increased cysteine requirement, with a demand for cysteine thus exceeding the body's capacity for cysteine production⁽²⁻⁴⁾. Although insufficient intake of sulfur amino acids is clearly detrimental, negative effects of excess methionine or cysteine have also been reported⁽⁵⁻⁷⁾. It is therefore important to obtain a deeper understanding of the interaction between sulfur amino acid metabolism and nutritional demand, especially *in vivo*.

Optimising sulfur amino acid nutrition involves considering the different roles of these amino acids, which are recognised to exert several significant influences. They are precursors of essential molecules and act as mediators affecting metabolism and cell functions. For example, cysteine is required for the synthesis of glutathione and taurine, which play a crucial role in oxidative stress conditions since they have the capacity to affect cellular redox status. Some mechanisms through which sulfur amino acids control oxidative status, and potentially amino acid signalling, have been detailed in a recent $review^{(8)}$ and they will be not presented here. In the present review, we first address the implications of the roles related to oxidative stress and cell function for nutrition recommendations. Since sulfur amino acids may also be involved in the regulation of gene transcription and participate in methyl group metabolism, we then provide an overview of the roles of sulfur amino acids in controlling gene expression and methvlation processes, focusing particularly on the consequences for amino acid nutrition.

Abbreviations: ATF4, activating transcription factor 4; CHOP, CCAAT/enhancer binding protein homologous protein; CpG, cytosine-guanine dinucleotide. * Corresponding author: Dr S. Tesseraud, fax + 33 2 47 42 77 78, email tesserau@tours.inra.fr

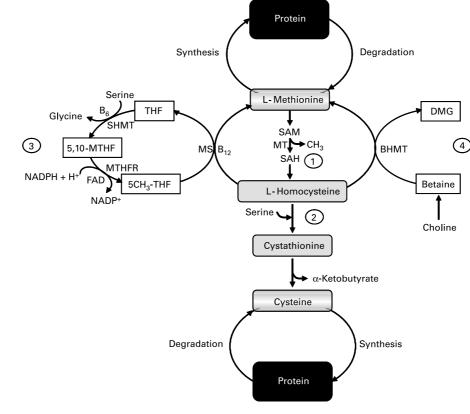


Fig. 1. Methionine–cysteine metabolic pathways. (1), Transmethylation; (2), trans-sulfuration; (3), folate-dependent remethylation; (4), folate-independent remethylation; B₆, vitamin B₆; B₁₂, vitamin B₁₂; BHMT, betaine homocysteine methyltransferase; 5-CH₃THF, 5-methyltetrahydrofolate; DMG, dimethylglycine; MS, methionine synthase; MT, methyl transferase; 5,10-MTHF, 5,10-methyl tetrahydrofolate; MTHFR, methylenetetrahydrofolate reductase; SAH, *S*-adenosylhomocysteine; SAM, *S*-adenosylmethionine; SHMT, serine hydroxymethyltransferase; THF, tetrahydrofolate.

Revisiting recommendations for methionine and cysteine in the light of their effects on oxidative stress and cell function

SS British Journal of Nutrition

Sulfur amino acids participate in the control of oxidative status since they are involved in the synthesis of intracellular antioxidants, particularly glutathione^(4,8,9), and in the methionine sulfoxide reductase antioxidant system^(8,10,11). Using cysteine derivatives (i.e. N-acetylcysteine) or cysteine-rich dietary proteins has been shown to increase blood and/or intracellular glutathione, which may affect the thiol redox status $^{(6,12-14)}$. Interestingly, dietary cysteine may also improve glucose control and alleviate sucrose-induced insulin resistance⁽¹⁵⁻¹⁷⁾. Moreover, several studies have reported a beneficial effect of cysteine supplementation at nutritional doses in the prevention of post-exercise oxidative stress in human subjects (18-21). There is also evidence that the cysteine/cystine redox couple acts as an extracellular thiol redox buffer (i.e. it represents the major plasma redox pool) and a modulator of cell function. In vitro studies have indicated that the extracellular cysteine/cystine redox potential affects proliferation and the p44/p42 mitogen-activated protein kinase pathway via a transforming growth factor α -dependent mechanism in enterocytes^(22,23), the production of reactive oxygen species and expression of cell-cell adhesion molecules in endothelial cells⁽²⁴⁾, and modulates reactive oxygen species-induced apop-tosis in retinal pigment epithelial cells⁽²⁵⁾ and Burkitt's lymphoma cells⁽²⁶⁾. In these studies, the effects of the cysteine/

cystine redox potential as a thiol/disulfide redox couple were independent of intracellular glutathione status, suggesting a specific role of the extracellular redox environment in cell function⁽²⁷⁾.

The cysteine/cystine redox potential in plasma has recently been shown to vary during the day, with a pattern influenced by meals which may reflect acute changes in plasma thiol redox state with dietary intake of sulfur amino acids⁽²⁸⁾. Interestingly, it has been found in rats that the cysteine/cystine redox potential was affected by both decrease and increase in the sulfur amino acid (L-methionine + L-cystine) content of the diet, in contrast to glutathione redox potential⁽²⁹⁾. Taken together, these studies provide new insights into the roles of cysteine and dietary sulfur amino acids in controlling cell function through modulation of the extracellular redox environment. These findings also highlight the complexity of the underlying mechanisms and do not support any systematic benefit from supplementing sulfur amino acids. In agreement with this assumption, decreasing methionine ingestion could be recommended in some situations. For example, due to the role of the methionine sulfoxide reductase antioxidant system, lowering dietary methionine levels appears to decrease the sensitivity of proteins to oxidative damage, oxidative stress and reactive oxygen species production^(30,31). Possible dietary implications for humans were discussed in a recent review⁽³²⁾. Since the intake of proteins, and therefore methionine, is higher than required in humans in developed countries, the authors suggest that decreasing excessive

consumption could be an efficient way to reduce tissue oxidative stress and improve healthy life span.

Sulfur amino acids, particularly cysteine, may also act on cell function as precursors of hydrogen sulfide $(H_2S)^{(33)}$. First considered as a toxic gas, H₂S is now recognised for its physiological function as the third (with NO and CO) gaseous transmitter in the body. In the brain, H₂S may act as a neuromodulator and protect neurons from oxidative stress^(34–38). H₂S has been shown to directly activate K_{ATP} channels in vascular smooth muscle cells and insulin-secreting cells⁽³⁹⁻⁴¹⁾, thus promoting vasorelaxation and inhibiting insulin secretion⁽³⁹⁻⁴⁵⁾. Although little is known regarding the production of endogenous H₂S in response to changes in cysteine concentrations, it is important to consider the possibility that sulfur amino acids might affect cell function through modulation of H₂S and not to ignore the possible consequences in terms of nutrition. Similarly, it is necessary to consider other products of sulfur amino acid metabolism such as sulfates. Sulfate is the predominant endproduct of sulfur amino acid catabolism, and a relationship has been reported between sulfate excretion and sulfur amino acid intake^(46,47). Sulfates contribute to glycosaminoglycan synthesis and detoxification processes through reactions depending on 3'-phosphoadenosine-5'-phosphosulfate, and these sulfation reactions account for 27 % of inorganic sulfate turnover in humans⁽⁴⁸⁾. In rats, ingesting a sulfur-deficient diet decreases plasma sulfate and elimination of the analgesic acetaminophen, with concomitant increase in its toxication⁽⁴⁹⁾, highlighting the importance of adequate methionine and/or cysteine intake for detoxification processes. Nevertheless, sulfates have long been thought to increase urinary excretion of Ca^(50,51); a recent study suggests that sulfur amino acids counterbalance the positive association between protein intake and bone mineral density⁽⁵²⁾. These positive and negative effects of sulfates should be taken into account when establishing recommendations for sulfur amino acid intake.

Potential function of sulfur amino acids in controlling gene expression

Amino acids are recognised to act as modulators of signal transduction pathways that control gene transcription and translation⁽⁵³⁻⁵⁵⁾. Some *in vitro* experiments performed in mammals^(56,57) and avian species⁽⁵⁸⁾ have suggested that methionine may have such a signal function by inducing the activation of an intracellular kinase involved in the control of mRNA translation, i.e. the p70 S6 kinase (p70S6K, also called S6K1). In addition, amino acid availability regulates the mammalian general control non-depressible 2/eukaryotic initiation factor 2α (eIF2 α) pathway that is thought to be affected by tRNA deacylation in the case of amino acid depletion⁽⁵⁹⁻⁶¹⁾. By influencing the phosphorylation of eIF2 α , this pathway affects (1) the first step in the initiation of mRNA translation through the formation of eIF2-GTP, and (2) amino acid-controlled gene expression through the induction of activating transcription factor 4 (ATF4). Indeed, there is evidence that ATF4 can bind to the amino acid response element in the promoters of genes such as CCAAT/enhancer binding protein (C/EBP) homologous protein (CHOP) and asparagine synthetase, thereby up-regulating their expression^(54,62). In relation to the present review, CHOP

and asparagine synthetase are over-expressed when cells are deprived of a particular amino acid, for instance leucine, lysine, phenylalanine and methionine^(63,64). Different mechanisms are probably involved in such regulation since CHOP expression is strongly induced in response to methionine deprivation but only slightly affected by cysteine, asparagine or histidine deprivation, whereas asparagine synthetase expression is induced equally whatever the amino acid depleted^(54,63). Cysteine restriction may induce the expression of several genes via the mammalian general control nondepressible 2 protein kinase/ATF4-dependent integrated stress response pathway to cope with oxidative and chemical stresses⁽⁶⁵⁾. It has also been reported that homocysteine (a metabolite of methionine) increases the expression of vascular endothelial growth factor by a mechanism involving ATF4⁽⁶⁶⁾. It is of note that CHOP can interfere with C/EBP α/β expression and function, thus regulating adipogenesis, which could provide a mechanism through which amino acids have an effect on the pathogenesis of insulin resistance, as proposed by Tremblay *et al.* ⁽⁶¹⁾. Moreover, methionine and cysteine are potent inhibitors of insulin-stimulated glucose transport in muscle cells⁽⁶⁷⁾. These results have been reported in non-physiological conditions, and the importance of a p70 S6 kinase-dependent regulatory loop in the development and/or the maintenance of insulin resistance remains to be determined⁽⁶⁸⁾. More information is therefore needed on the potential adverse effects of sulfur amino acid supplementation before use for nutritional purposes.

Many other examples demonstrate that sulfur amino acids are able to regulate the expression of numerous genes, thereby controlling nutrient metabolism and cell functions. Methionine supply has been shown (1) to affect growth hormone-induced insulin-like growth factor I gene expression in ovine hepato $cytes^{(57)}$ and (2) to regulate the expression of an important gene controlling ubiquitin-proteasome-dependent proteolysis (i.e. E3 ubiquitin ligase atrogin-1) in avian fibroblasts⁽⁶⁹⁾. Cysteine and cysteine derivatives (for example, N-acetylcysteine, or N-acetylcysteine amide) can modulate the activity of NF- $\kappa B^{(70-73)},$ which induces the expression of many genes that are involved in cell survival and proliferation, and in the regulation of immune and inflammatory responses (for reviews, see Barnes & Karin⁽⁷⁴⁾, Chen et al.⁽⁷⁵⁾, Grimble⁽⁷⁶⁾ and Wek *et al.* ⁽⁶⁰⁾). The activation pathways of this transcription factor involve the thiol redox status that responds to antioxidants including cysteine (see the section 'Revisiting recommendations for methionine and cysteine in the light of their effects on oxidative stress and cell function' above). Despite the clear theoretical importance of sulfur amino acids in immune function, studies directly linking methionine and cysteine intake and immune function are still needed⁽⁷⁶⁾. Sulfur amino acids are also recognised to control genes involved in lipid metabolism, including cholesterol 7α -hydroxylase and apoA-I⁽⁷⁷⁾, which could be in favour of sulfur amino acid supplementation in cases of abnormal lipid metabolism. Nevertheless, high methionine intake can promote the development of atheromatous disorder⁽⁷⁸⁾, indicating that any such supplementation has to be undertaken with care to prevent atherosclerosis. The underlying mechanisms are still unclear. Methionine is the precursor of homocysteine (Fig. 1) and elevated levels of plasma homocysteine (hyperhomocysteinaemia) have been associated with occlusive

https://doi.org/10.1017/S0007114508159025 Published online by Cambridge University Pres.

vascular disease⁽⁷⁹⁾. However, Troen *et al.*⁽⁷⁸⁾ have shown in mice that the atherogenic effect of methionine exists even in the absence of hyperhomocysteinaemia. High plasma homocysteine levels appear not to be independently atherogenic as recently reported^(80,81). Vitamin B supplementation (its effect on methionine metabolism is discussed in the section 'Role of sulfur amino acids in the methylation processes' below) does not prevent CVD in humans despite lowering homocysteine^(82,83), similarly challenging the role of homocysteine as an independent cardiovascular risk factor.

Role of sulfur amino acids in the methylation processes

In addition to the above roles, sulfur amino acids have another major function since methionine is a source of the methyl groups needed for all biological methylation reactions, including methylation of DNA and histones, a process that influences chromatin structure and gene expression⁽⁸⁴⁻⁸⁹⁾. DNA methylation generally occurs at cytosines within cytosine-guanine dinucleotides (CpG). The CpG sites are dispersed throughout the genome, with specific CpGrich regions called CpG islands. The methylation status of cytosine residues within CpG dinucleotides and in the context of CpG islands determines which genes are active or not, and plays an important role in maintaining gene silencing that is needed for tissue- and development-specific gene expression, epigenetic mechanisms involved in the establishment and maintenance of gene expression patterns, and genomic imprinting.

Being converted to S-adenosylmethionine, the major biological methylating agent, methionine provides the methyl groups that are needed for cellular methylation reactions. However, in order to adapt dietary methionine intake effectively, it is important to keep in mind the specific features of the methyl metabolism that is more generally involved in methylation regulation. S-adenosylmethionine is converted to S-adenosylhomocysteine following methyl donation, and to homocysteine (reactions of transmethylation; Fig. 1). Homocysteine can be remethylated into methionine through two separate pathways: (1) the folate-dependent remethylation pathway that involves 5,10-methyl tetrahydrofolate; (2) the folate-independent remethylation pathway in which the methyl group is provided by betaine. Disorders of methyl balance lead to several diseases (liver disease, CVD, closure of the neural tube, synthesis of creatine, cancer), illustrating the key role played by methylation reactions^(90,91). Whether DNA methylation is increased by high levels of methionine is still unclear, since, for example, excess methionine may impair DNA methylation by inhibiting remethylation of homocysteine⁽⁹²⁾. These findings need further investigation to quantify their physiological relevance, particularly using more physiological amounts of methionine. In addition, based on the methionine cycle, nutritional factors affecting the supply of S-adenosylmethionine and/or removal of homocysteine (for example, dietary choline, betaine, folic acid and vitamin B₁₂) potentially affect DNA methylation and must therefore be taken into account when assessing the supply of methionine. Mathematical models have been developed to consider the complexity of methionine metabolic pathways, and they will be useful to complement and help guide laboratory studies^(93,94)

Recent in vivo studies have demonstrated opposite dietrelated changes in DNA methylation according to the tissue studied: long-term administration of a folate/methyl-deficient diet in rats results in progressive hypomethylation of DNA in the liver⁽⁹⁵⁾, but an increase in DNA methylation in the brain⁽⁹⁶⁾. Although the underlying mechanisms and significance of these findings are not understood, insufficient supply of the nutrients involved in methyl metabolism clearly affects methylation reactions and concomitantly alters patterns of gene expression⁽⁹⁶⁾. This function of methyl group donors represents another mechanism though which sulfur amino acids can regulate gene expression (see section 'Potential function of sulfur amino acids in controlling gene expression' above) with potentially both negative and positive consequences, for example, either unsuitable dysregulation of gene expression or conversely restoration of appropriate locus-specific DNA methylation using therapeutic pro-methylation diets⁽⁸⁹⁾. Epigenetic regulation thus plays an important role in the potential development (or in contrast prevention) of cancer, CVD, type 2 diabetes and obesity. For example, the availability of methyl group donors regulates the expression of genes involved in the development of the digestive tract through a mechanism dependent on DNA methylation, at least for the homeobox gene CDX1, which could explain the protective role attributed to folates in colon cancer⁽⁹⁷⁾.

An increasing number of reviews have highlighted the importance of perinatal nutrition (i.e. nutrition in early life), since aberrant methyl metabolism in utero is linked with certain disorders (for reviews, see Ulrey et al. (85), Waterland⁽⁸⁹⁾, Rees *et al.* ⁽⁹⁸⁾, Waterland & Michels⁽⁸⁷⁾ and Nafee et al.⁽⁸⁸⁾). For example, a murine metastable epiallele (axin fused, Axin^{Fu}) exhibits epigenetic plasticity to the maternal diet, since supplementation with methyl group donors before and during pregnancy increases offspring DNA methylation at Axin^{Fu(99)}. As discussed by some authors^(89,98,100), manipulating the sulfur amino acid content of the early diet may induce chronic changes in cell functions that have implications for long-term health. More recent findings have provided evidence that changes in the supply of methionine and specific B vitamins such as B_{12} during the periconception period can lead to widespread epigenetic changes in DNA methylation in offspring, and modify adult health-related phenotypes⁽¹⁰¹⁾. For instance, adult offspring exhibit changes in body composition (fatness), immune function and insulin response. Interestingly, these effects were obtained with modest early dietary intervention (changes within physiological ranges), indicating the importance of adequate dietary methyl group supply for metabolic programming and the need for further studies to define the level of intake. Such studies will certainly provide a rationale for nutritional advice in the future.

Conclusion

Sulfur amino acids have a very significant place among amino acids due to their numerous roles. They are precursors of major components such as glutathione, taurine, H_2S and sulfates, and thus act on oxidative status and various signalling pathways. In addition, methionine plays a unique role in epigenetic regulation by affecting DNA methylation. Sulfur amino acids consequently affect metabolism, gene expression

https://doi.org/10.1017/S0007114508159025 Published online by Cambridge University Press

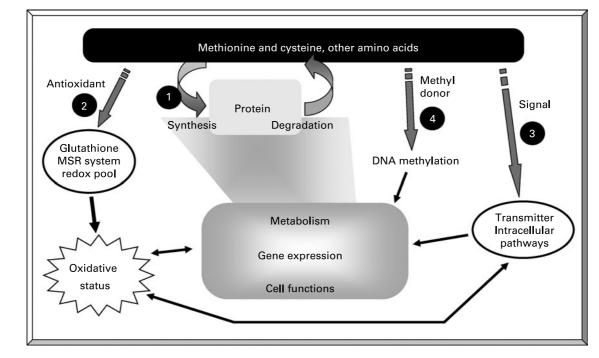


Fig. 2. Regulation of metabolism and cell functions, and possible roles of sulfur amino acids: (1) precursors of protein synthesis; (2) antioxidant function; (3) nutrient signal; (4) methyl group donor. The figure represents a summary of the findings discussed in the present review. MSR, methionine sulfoxide reductase.

and cell functions, as illustrated in Fig. 2, and a change in their dietary levels may have either beneficial or deleterious consequences. An increasing number of studies have been devoted to exploring the key roles of sulfur amino acids, but further experiments are still necessary to obtain direct evidence regarding some of their effects on nutrient metabolism, cell functions and more generally physiological responses in animals and humans. Understanding of the mechanisms of amino acid actions is essential to optimise dietary amino acid requirements whatever the conditions, whether physiological (pregnancy, early nutrition) or physiopathological (disease, ageing). In particular, the effects of sulfur amino acids should be analysed under circumstances affecting oxidative status and gene expression. Recommendations for methionine and cysteine intake have to be revisited in the light of their roles and the intricacies of sulfur amino acid metabolism. The availability of other dietary nutrients (for example, cysteine, vitamins) has of course to be taken into account when adjusting the supply of methionine, for instance. However, a major challenge for future experiments is to improve the integration of the various roles of sulfur amino acids at the whole-body level for rational use in nutrition.

Acknowledgements

The authors thank S. Kaushik (INRA, St Pée-sur-Nivelle, France) for helpful comments and discussions, and D. Raine (Surrey, UK) for revision of the English language. Studies on sulfur amino acids in the laboratories of the authors were funded by INRA and Adisseo France SA. The contribution of each author to the paper was as follows: S. T., analysis of the literature, writing of the manuscript and supervision of the review; S. M. C. and A. C., writing of the manuscript, production of the figures and referencing the literature cited; I. S., writing of the manuscript, and discussion to define the aims and structure of the review.

S. T., S. M. C., A. C. and I. S. declare no conflicts of interest.

References

- Hocquette JF, Tesseraud S, Cassar-Malek I, et al. (2007) Responses to nutrients in farm animals: implications for production and quality. Animal 1, 1297–1313.
- Obled C, Papet I & Breuille D (2004) Sulfur-containing amino acids and glutathione in diseases. In *Metabolic and Therapeutic Aspects of Amino Acids in Clinical Nutrition*, 2nd ed., pp. 667–687 [LA Cynober, editor]. Boca Raton, FL: CRC Press LLC.
- Le Floc'h N, Melchior D & Obled C (2004) Modifications of protein and amino acid metabolism during inflammation and immune system activation. *Livest Prod Sci* 87, 37–45.
- Soeters PB, van de Poll MC, van Gemert WG, *et al.* (2004) Amino acid adequacy in pathophysiological states. *J Nutr* 134, 1575S-1582S.
- van de Poll MC, Dejong CH & Soeters PB (2006) Adequate range for sulfur-containing amino acids and biomarkers for their excess: lessons from enteral and parenteral nutrition. *J Nutr* 136, 1694S-1700S.
- Stipanuk MH, Dominy JE Jr, Lee JI, et al. (2006) Mammalian cysteine metabolism: new insights into regulation of cysteine metabolism. J Nutr 136, 16528–16598.
- Garlick PJ (2006) Toxicity of methionine in humans. J Nutr 136, 1722S-1725S.
- Métayer S, Seiliez I, Collin A, *et al.* (2008) Mechanisms through which sulfur amino acids control protein metabolism and oxidative status. *J Nutr Biochem* 19, 207–215.

- 9. Wu G, Fang YZ, Yang S, *et al.* (2004) Glutathione metabolism and its implications for health. *J Nutr* **134**, 489–492.
- Moskovitz J, Rahman MA, Strassman J, et al. (1995) Escherichia coli peptide methionine sulfoxide reductase gene: regulation of expression and role in protecting against oxidative damage. J Bacteriol 177, 502–507.
- Levine RL, Mosoni L, Berlett BS, et al. (1996) Methionine residues as endogenous antioxidants in proteins. Proc Natl Acad Sci U S A 93, 15036–15040.
- Mariotti F, Simbelie KL, Makarios-Lahham L, *et al.* (2004) Acute ingestion of dietary proteins improves post-exercise liver glutathione in rats in a dose-dependent relationship with their cysteine content. *J Nutr* **134**, 128–131.
- Diniz YS, Rocha KK, Souza GA, *et al.* (2006) Effects of N-acetylcysteine on sucrose-rich diet-induced hyperglycaemia, dyslipidemia and oxidative stress in rats. *Eur J Pharmacol* 543, 151–157.
- Dröge W (2006) Redox regulation in anabolic and catabolic processes. Curr Opin Clin Nutr Metab Care 9, 190–195.
- Blouet C, Mariotti F, Azzout-Marniche D, et al. (2007) Dietary cysteine alleviates sucrose-induced oxidative stress and insulin resistance. *Free Radic Biol Med* 42, 1089–1097.
- Blouet C, Mariotti F, Mikogami T, *et al.* (2007) Meal cysteine improves postprandial glucose control in rats fed a high-sucrose meal. *J Nutr Biochem* 18, 519–524.
- Mariotti F, Hermier D, Sarrat C, *et al.* (2008) Rapeseed protein inhibits the initiation of insulin resistance by a high-saturated fat, high-sucrose diet in rats. *Br J Nutr* **100**, 984–991.
- Tsakiris S, Parthimos T, Parthimos N, *et al.* (2006) The beneficial effect of L-cysteine supplementation on DNA oxidation induced by forced training. *Pharmacol Res* 53, 386–390.
- Parthimos T, Tsopanakis C, Angelogianni P, et al. (2007) L-Cysteine supplementation prevents exercise-induced alterations in human erythrocyte membrane acetylcholinesterase and Na⁺,K⁺-ATPase activities. *Clin Chem Lab Med* 45, 67–72.
- Schulpis KH, Reclos GJ, Parthimos T, et al. (2006) L-Cysteine supplementation protects the erythrocyte glucose-6-phosphate dehydrogenase activity from reduction induced by forced training. Clin Biochem 39, 1002–1006.
- Lands LC, Grey VL & Smountas AA (1999) Effect of supplementation with a cysteine donor on muscular performance. J Appl Physiol 87, 1381–1385.
- Jonas CR, Ziegler TR, Gu LH, *et al.* (2002) Extracellular thiol/ disulfide redox state affects proliferation rate in a human colon carcinoma (Caco2) cell line. *Free Radic Biol Med* 33, 1499–1506.
- Nkabyo YS, Go YM, Ziegler TR, et al. (2005) Extracellular cysteine/cystine redox regulates the p44/p42 MAPK pathway by metalloproteinase-dependent epidermal growth factor receptor signaling. Am J Physiol Gastrointest Liver Physiol 289, G70–G78.
- Go YM & Jones DP (2005) Intracellular proatherogenic events and cell adhesion modulated by extracellular thiol/disulfide redox state. *Circulation* 111, 2973–2980.
- Jiang S, Moriarty-Craige SE, Orr M, et al. (2005) Oxidantinduced apoptosis in human retinal pigment epithelial cells: dependence on extracellular redox state. *Invest Ophthalmol* Vis Sci 46, 1054–1061.
- Banjac A, Perisic T, Sato H, *et al.* (2008) The cystine/cysteine cycle: a redox cycle regulating susceptibility *versus* resistance to cell death. *Oncogene* 27, 1618–1628.
- 27. Go YM & Jones DP (2008) Redox compartmentalization in eukaryotic cells. *Biochim Biophys Acta* **1780**, 1273–1290.
- Blanco RA, Ziegler TR, Carlson BA, *et al.* (2007) Diurnal variation in glutathione and cysteine redox states in human plasma. *Am J Clin Nutr* 86, 1016–1023.

- 29. Nkabyo YS, Gu LH, Jones DP, *et al.* (2006) Thiol/disulfide redox status is oxidized in plasma and small intestinal and colonic mucosa of rats with inadequate sulfur amino acid intake. *J Nutr* **136**, 1242–1248.
- Sanz A, Caro P, Ayala V, *et al.* (2006) Methionine restriction decreases mitochondrial oxygen radical generation and leak as well as oxidative damage to mitochondrial DNA and proteins. *FASEB J* 20, 1064–1073.
- Caro P, Gómez J, López-Torres M, *et al.* (2008) Forty percent and eighty percent methionine restriction decrease mitochondrial ROS generation and oxidative stress in rat liver. *Biogerontology* 9, 183–196.
- 32. López-Torres M & Barja G (2008) Lowered methionine ingestion as responsible for the decrease in rodent mitochondrial oxidative stress in protein and dietary restriction. Possible implications for humans. *Biochim Biophys Acta* **1780**, 1337–1347.
- Stipanuk MH (2004) Sulfur amino acid metabolism: pathways for production and removal of homocysteine and cysteine. *Annu Rev Nutr* 24, 539–577.
- Abe K & Kimura H (1996) The possible role of hydrogen sulfide as an endogenous neuromodulator. *J Neurosci* 16, 1066–1071.
- Eto K, Asada T, Arima K, *et al.* (2002) Brain hydrogen sulfide is severely decreased in Alzheimer's disease. *Biochem Biophys Res Commun* 293, 1485–1488.
- Han Y, Qin J, Chang X, et al. (2005) Hydrogen sulfide may improve the hippocampal damage induced by recurrent febrile seizures in rats. Biochem Biophys Res Commun 327, 431–436.
- Kimura H (2000) Hydrogen sulfide induces cyclic AMP and modulates the NMDA receptor. *Biochem Biophys Res Commun* 267, 129–133.
- Kimura Y & Kimura H (2004) Hydrogen sulfide protects neurons from oxidative stress. FASEB J 18, 1165–1167.
- Zhao W, Zhang J, Lu Y, *et al.* (2001) The vasorelaxant effect of H₂S as a novel endogenous gaseous KATP channel opener. *EMBO J* 20, 6008–6016.
- Tang G, Wu L, Liang W, et al. (2005) Direct stimulation of KATP channels by exogenous and endogenous hydrogen sulfide in vascular smooth muscle cells. *Mol Pharmacol* 68, 1757–1764.
- Yang W, Yang G, Jia X, *et al.* (2005) Activation of KATP channels by H₂S in rat insulin-secreting cells and the underlying mechanisms. *J Physiol* 569, 519–531.
- Zhao W & Wang R (2002) H₂S-induced vasorelaxation and underlying cellular and molecular mechanisms. *Am J Physiol Heart Circ Physiol* 283, H474–H480.
- Cheng Y, Ndisang JF, Tang G, *et al.* (2004) Hydrogen sulfideinduced relaxation of resistance mesenteric artery beds of rats. *Am J Physiol Heart Circ Physiol* 287, H2316–H2323.
- Hosoki R, Matsuki N & Kimura H (1997) The possible role of hydrogen sulfide as an endogenous smooth muscle relaxant in synergy with nitric oxide. *Biochem Biophys Res Commun* 237, 527–531.
- 45. Kaneko Y, Kimura Y, Kimura H, *et al.* (2006) L-Cysteine inhibits insulin release from the pancreatic β -cell: possible involvement of metabolic production of hydrogen sulfide, a novel gasotransmitter. *Diabetes* **55**, 1391–1397.
- 46. Hamadeh MJ & Hoffer LJ (2001) Use of sulfate production as a measure of short-term sulfur amino acid catabolism in humans. *Am J Physiol Endocrinol Metab* 280, E857–E866.
- Hamadeh MJ & Hoffer LJ (2003) Effect of protein restriction on sulfur amino acid catabolism in insulin-dependent diabetes mellitus. *Am J Physiol Endocrinol Metab* 284, E382–E389.
- Hoffer LJ, Hamadeh MJ, Robitaille L, et al. (2005) Human sulfate kinetics. Am J Physiol Regul Integr Comp Physiol 289, R1372-R1380.
- Gregus Z, Kim HJ, Madhu C, *et al.* (1994) Sulfation of acetaminophen and acetaminophen-induced alterations in sulfate and 3'-phosphoadenosine 5'-phosphosulfate homeostasis in

rats with deficient dietary intake of sulfur. *Drug Metab Dispos* **22**, 725–730.

- 50. Hu JF, Zhao XH, Parpia B, *et al.* (1993) Dietary intakes and urinary excretion of calcium and acids: a cross-sectional study of women in China. *Am J Clin Nutr* **58**, 398–406.
- 51. Whiting SJ & Draper HH (1980) The role of sulfate in the calciuria of high protein diets in adult rats. *J Nutr* **110**, 212–222.
- 52. Thorpe M, Mojtahedi MC, Chapman-Novakofski K, et al. (2008) A positive association of lumbar spine bone mineral density with dietary protein is suppressed by a negative association with protein sulfur. J Nutr 138, 80–85.
- Jefferson LS & Kimball SR (2001) Amino acid regulation of gene expression. J Nutr 131, 2460S–2466S.
- Kilberg MS, Pan YX, Chen H, *et al.* (2005) Nutritional control of gene expression: how mammalian cells respond to amino acid limitation. *Annu Rev Nutr* 25, 59–85.
- 55. Jousse C, Averous J, Bruhat A, *et al.* (2004) Amino acids as regulators of gene expression: molecular mechanisms. *Biochem Biophys Res Commun* **313**, 447–452.
- Shigemitsu K, Tsujishita Y, Miyake H, et al. (1999) Structural requirement of leucine for activation of p70 S6 kinase. FEBS Lett 447, 303–306.
- Stubbs AK, Wheelhouse NM, Lomax MA, *et al.* (2002) Nutrient-hormone interaction in the ovine liver: methionine supply selectively modulates growth hormone-induced IGF-I gene expression. *J Endocrinol* 174, 335–341.
- Tesseraud S, Bigot K & Taouis M (2003) Amino acid availability regulates S6K1 and protein synthesis in avian insulininsensitive QM7 myoblasts. *FEBS Lett* 540, 176–180.
- Kimball SR & Jefferson LS (2005) Role of amino acids in the translational control of protein synthesis in mammals. *Semin Cell Dev Biol* 16, 21–27.
- Wek RC, Jiang HY & Anthony TG (2006) Coping with stress: eIF2 kinases and translational control. *Biochem Soc Trans* 34, 7–11.
- 61. Tremblay F, Lavigne C, Jacques H, *et al.* (2007) Role of dietary proteins and amino acids in the pathogenesis of insulin resistance. *Annu Rev Nutr* **27**, 293–310.
- Averous J, Bruhat A, Jousse C, *et al.* (2004) Induction of CHOP expression by amino acid limitation requires both ATF4 expression and ATF2 phosphorylation. *J Biol Chem* 279, 5288–5297.
- 63. Jousse C, Bruhat A, Ferrara M, *et al.* (2000) Evidence for multiple signaling pathways in the regulation of gene expression by amino acids in human cell lines. *J Nutr* **130**, 1555–1560.
- Bruhat A, Jousse C, Wang XZ, et al. (1997) Amino acid limitation induces expression of CHOP, a CCAAT/enhancer binding protein-related gene, at both transcriptional and post-transcriptional levels. J Biol Chem 272, 17588–17593.
- Lee JI, Dominy JE Jr, Sikalidis AK, *et al.* (2008) HepG2/C3A cells respond to cysteine deprivation by induction of the amino acid deprivation/integrated stress response pathway. *Physiol Genomics* 33, 218–229.
- 66. Roybal CN, Yang S, Sun CW, *et al.* (2004) Homocysteine increases the expression of vascular endothelial growth factor by a mechanism involving endoplasmic reticulum stress and transcription factor ATF4. *J Biol Chem* 279, 14844–14852.
- Tremblay F & Marette A (2001) Amino acid and insulin signaling via the mTOR/p70 S6 kinase pathway. A negative feedback mechanism leading to insulin resistance in skeletal muscle cells. J Biol Chem 276, 38052–38060.
- Hinault C, Van Obberghen E & Mothe-Satney I (2006) Role of amino acids in insulin signaling in adipocytes and their potential to decrease insulin resistance of adipose tissue. *J Nutr Biochem* 17, 374–378.
- Tesseraud S, Métayer-Coustard S, Boussaid S, et al. (2007) Insulin and amino acid availability regulate atrogin-1 in avian QT6 cells. Biochem Biophys Res Commun 357, 181–186.

- Mihm S, Ennen J, Pessara U, *et al.* (1991) Inhibition of HIV-1 replication and NF-κB activity by cysteine and cysteine derivatives. *AIDS* 5, 497–503.
- 71. Shibanuma M, Kuroki T & Nose K (1994) Inhibition by *N*-acetyl-L-cysteine of interleukin-6 mRNA induction and activation of NF κ B by tumor necrosis factor α in a mouse fibroblastic cell line, Balb/3T3. *FEBS Lett* **353**, 62–66.
- Paterson RL, Galley HF & Webster NR (2003) The effect of N-acetylcysteine on nuclear factor-κB activation, interleukin-6, interleukin-8, and intercellular adhesion molecule-1 expression in patients with sepsis. Crit Care Med 31, 2574–2578.
- Lee KS, Kim SR, Park HS, *et al.* (2007) A novel thiol compound, N-acetylcysteine amide, attenuates allergic airway disease by regulating activation of NF-κB and hypoxia-inducible factor-1α. *Exp Mol Med* **39**, 756–768.
- Barnes PJ & Karin M (1997) Nuclear factor-κB: a pivotal transcription factor in chronic inflammatory diseases. *N Engl J Med* 336, 1066–1071.
- 75. Chen F, Castranova V, Shi X, *et al.* (1999) New insights into the role of nuclear factor-κB, a ubiquitous transcription factor in the initiation of diseases. *Clin Chem* **45**, 7–17.
- Grimble RF (2006) The effects of sulfur amino acid intake on immune function in humans. J Nutr 136, Suppl. 6, 1660S–1665S.
- Oda H (2006) Functions of sulfur-containing amino acids in lipid metabolism. J Nutr 136, Suppl. 6, 1666S–1669S.
- Troen AM, Lutgens E, Smith DE, et al. (2003) The atherogenic effect of excess methionine intake. Proc Natl Acad Sci U S A 100, 15089–15094.
- 79. Gerhard GT & Duell PB (1999) Homocysteine and atherosclerosis. *Curr Opin Lipidol* **10**, 417–428.
- Zhou J, Werstuck GH, Lhoták S, *et al.* (2008) Hyperhomocysteinemia induced by methionine supplementation does not independently cause atherosclerosis in C57BL/6J mice. *FASEB J* 22, 2569–2578.
- Liu C, Wang Q, Guo H, *et al.* (2008) Plasma S-adenosylhomocysteine is a better biomarker of atherosclerosis than homocysteine in apolipoprotein E-deficient mice fed high dietary methionine. J Nutr 138, 311–315.
- Jamison RL, Hartigan P, Kaufman JS, *et al.* (2007) Effect of homocysteine lowering on mortality and vascular disease in advanced chronic kidney disease and end-stage renal disease: a randomized controlled trial. *JAMA* 298, 1163–1170.
- Albert CM, Cook NR, Gaziano JM, *et al.* (2008) Effect of folic acid and B vitamins on risk of cardiovascular events and total mortality among women at high risk for cardiovascular disease: a randomized trial. *JAMA* 299, 2027–2036.
- Egger G, Liang G, Aparicio A, *et al.* (2004) Epigenetics in human disease and prospects for epigenetic therapy. *Nature* 429, 457–463.
- Ulrey CL, Liu L, Andrews LG, et al. (2005) The impact of metabolism on DNA methylation. Hum Mol Genet 14, Spec. no. 1 R139–R147.
- Grønbaek K, Hother C & Jones PA (2007) Epigenetic changes in cancer. *APMIS* 115, 1039–1059.
- Waterland RA & Michels KB (2007) Epigenetic epidemiology of the developmental origins hypothesis. *Annu Rev Nutr* 27, 363–388.
- Nafee TM, Farrell WE, Carroll WD, *et al.* (2008) Epigenetic control of fetal gene expression. *BJOG* 115, 158–168.
- 89. Waterland RA (2006) Assessing the effects of high methionine intake on DNA methylation. *J Nutr* **136**, Suppl. 6, 1706S–1710S.
- Brosnan JT, da Silva R & Brosnan ME (2007) Amino acids and the regulation of methyl balance in humans. *Curr Opin Clin Nutr Metab Care* 10, 52–57.
- Williams KT & Schalinske KL (2007) New insights into the regulation of methyl group and homocysteine metabolism. J Nutr 137, 311–314.

- Dunlevy LP, Burren KA, Chitty LS, *et al.* (2006) Excess methionine suppresses the methylation cycle and inhibits neural tube closure in mouse embryos. *FEBS Lett* 580, 2803–2807.
- Reed MC, Nijhout HF, Sparks R, et al. (2004) A mathematical model of the methionine cycle. J Theor Biol 226, 33–43.
- Reed MC, Nijhout HF, Neuhouser ML, *et al.* (2006) A mathematical model gives insights into nutritional and genetic aspects of folate-mediated one-carbon metabolism. *J Nutr* 136, 2653–2661.
- Pogribny IP, Ross SA, Wise C, et al. (2006) Irreversible global DNA hypomethylation as a key step in hepatocarcinogenesis induced by dietary methyl deficiency. *Mutat Res* 593, 80–87.
- Pogribny IP, Karpf AR, James SR, *et al.* (2008) Epigenetic alterations in the brains of Fisher 344 rats induced by longterm administration of folate/methyl-deficient diet. *Brain Res* 1237, 25–34.

- 97. Lu X, Freund JN, Muller M, *et al.* (2008) Differential regulation of CDX1 and CDX2 gene expression by deficiency in methyl group donors. *Biochimie* **90**, 697–704.
- Rees WD, Wilson FA & Maloney CA (2006) Sulfur amino acid metabolism in pregnancy: the impact of methionine in the maternal diet. *J Nutr* 136, Suppl. 6, 1701S–1705S.
- 99. Waterland RA, Dolinoy DC, Lin JR, *et al.* (2006) Maternal methyl supplements increase offspring DNA methylation at Axin Fused. *Genesis* **44**, 401–406.
- Rees WD (2002) Manipulating the sulfur amino acid content of the early diet and its implications for long-term health. *Proc Nutr Soc* 61, 71–77.
- Sinclair KD, Allegrucci C, Singh R, *et al.* (2007) DNA methylation, insulin resistance, and blood pressure in offspring determined by maternal periconceptional B vitamin and methionine status. *Proc Natl Acad Sci U S A* 104, 19351–19356.

https://doi.org/10.1017/S0007114508159025 Published online by Cambridge University Press