

Major Review

*Current address: Department of Molecular Biosciences, Northwestern University, 2205 Tech Drive, Evanston, IL 60208-3500, USA.

Cite this article: Sinnige T, Stroobants K, Dobson CM, Vendruscolo M (2020). Biophysical studies of protein misfolding and aggregation in *in vivo* models of Alzheimer's and Parkinson's diseases. *Quarterly Reviews of Biophysics* **53**, e10, 1–43. <https://doi.org/10.1017/S0033583520000025>

Received: 28 February 2020

Revised: 19 April 2020

Accepted: 20 April 2020

Key words:

Alzheimer's disease; biophysical methods; *in vivo* models; Parkinson's disease; protein misfolding

Authors for correspondence:





Tessa Sinnige,

E-mail: tessa.sinnige@northwestern.edu;

Michele Vendruscolo,

E-mail: mv245@cam.ac.uk

Biophysical studies of protein misfolding and aggregation in *in vivo* models of Alzheimer's and Parkinson's diseases

Tessa Sinnige* , Karen Stroobants , Christopher M. Dobson 
and Michele Vendruscolo 

Department of Chemistry, Centre for Misfolding Diseases, University of Cambridge, Lensfield Road, Cambridge CB2 1EW, UK

Abstract

Neurodegenerative disorders, including Alzheimer's (AD) and Parkinson's diseases (PD), are characterised by the formation of aberrant assemblies of misfolded proteins. The discovery of disease-modifying drugs for these disorders is challenging, in part because we still have a limited understanding of their molecular origins. In this review, we discuss how biophysical approaches can help explain the formation of the aberrant conformational states of proteins whose neurotoxic effects underlie these diseases. We discuss in particular models based on the transgenic expression of amyloid- β (A β) and tau in AD, and α -synuclein in PD. Because biophysical methods have enabled an accurate quantification and a detailed understanding of the molecular mechanisms underlying protein misfolding and aggregation *in vitro*, we expect that the further development of these methods to probe directly the corresponding mechanisms *in vivo* will open effective routes for diagnostic and therapeutic interventions.

Table of contents

Introduction	1
Determination of protein aggregation mechanisms using <i>in vitro</i> biophysical methods	2
A β aggregation	3
Tau aggregation	5
α -Synuclein aggregation	6
<i>In vivo</i> models of protein misfolding diseases	7
A β models	7
Tau models	19
Models combining A β and tau	22
α -Synuclein models	24
Identification of biological pathways using <i>in vivo</i> models	26
Biological pathways in Alzheimer's disease models	27
Biological pathways in Parkinson's disease models	30
Methodological challenges	31
Conclusions	32

Introduction

Protein misfolding and aggregation are associated with a wide variety of human diseases (Hardy and Selkoe, 2002; Eisenberg and Jucker, 2012; Jucker and Walker, 2013; Knowles, Vendruscolo and Dobson, 2014; Chiti and Dobson, 2017). These diseases have been linked to a progressive failure of the protein homeostasis system, which controls protein folding, trafficking and degradation (Balch *et al.*, 2008; Kim *et al.*, 2013). The capacity of this system has been shown to decline with age both in model organisms and in humans, which can result in the accumulation of aberrant protein aggregates (Labbadia and Morimoto, 2015). Neurons appear to be particularly vulnerable to such events (Freer *et al.*, 2016; Surmeier *et al.*, 2017; Luna *et al.*, 2018; Fu *et al.*, 2019), and the vast majority of neurodegenerative diseases are accompanied by the formation of protein deposits, including those of amyloid- β (A β) and tau in Alzheimer's disease (AD), and of α -synuclein in Parkinson's disease (PD) (Hardy

© The Author(s), 2020. Published by Cambridge University Press. This is an Open Access article, distributed under the terms of the Creative Commons Attribution licence (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted re-use, distribution, and reproduction in any medium, provided the original work is properly cited.

Table 1. Commonly used biophysical methods for the study of protein misfolding *in vivo* and *in vitro*

<i>Methods and tools to probe protein misfolding and aggregation in vivo and in situ</i>	
Amyloid binding dyes	ThS, Congo red, NIAD-4, X-34, silver stain (tau tangles)
Microscopy	EM, immunohistochemistry, fluorescence microscopy including FLIM and FRAP
Conformation-specific antibodies	A11 (oligomers), MC1 (misfolded tau), TOC1 (tau oligomers), FILA-1 (α -synuclein oligomers), DesAbs (designed antibodies)
Detergent solubility	Sarkosyl, SDS
Size determination	SEC, sucrose gradient, SDS-PAGE, western blot
Visualisation of deposits	CLARITY, 3DISCO, Scale, MRI, PET
Biochemical assays	ELISA, SIMOA, amyloid-seeding assay, PMCA, RT-QuIC
<i>Methods to probe cellular pathways in vivo and in situ</i>	
Genetic screens	RNAi, random mutagenesis
Transcriptomics	RNA microarrays, next-generation sequencing (RNAseq)
Proteomics	TMT, iTRAQ, label-free mass spectrometry
<i>Methods to probe protein aggregation in vitro</i>	
Aggregation kinetics	ThT fluorescence, quartz crystal microbalance, filter trap assay
Secondary structure determination	CD spectroscopy, FTIR spectroscopy
Size/shape determination	DLS, SEC-MALS, smFRET, SAXS, AFM, super-resolution microscopy
3D structure determination	Cryo-EM, solid-state NMR spectroscopy, X-ray diffraction

The application of these methods to study protein aggregation and the associated references are mentioned throughout the main text.

and Selkoe, 2002; Eisenberg and Jucker, 2012; Jucker and Walker, 2013; Knowles *et al.*, 2014; Chiti and Dobson, 2017).

Although mature A β fibrils are in many cases relatively inert, studies in cell culture and animal models have revealed that misfolded oligomers, which are formed during the process of fibril formation, possess cytotoxic properties (Bucciantini *et al.*, 2002; Baglioni *et al.*, 2006; Haass and Selkoe, 2007; Bemporad and Chiti, 2012; Benilova *et al.*, 2012; Chiti and Dobson, 2017). Several mechanisms have been proposed to account for the toxicity of these oligomers, including disruption of lipid membranes, transcriptional deregulation, mitochondrial dysfunction, oxidative stress, proteasome inhibition and disruption of synaptic plasticity (Haass and Selkoe, 2007; Benilova *et al.*, 2012; Roberts and Brown, 2015; De *et al.*, 2019). Some of these mechanisms affect components of the protein homeostasis system, resulting in a collapse of protein quality control and a further increase in protein misfolding and toxicity (Labbadia and Morimoto, 2015). On the whole, however, the molecular mechanisms whereby protein misfolding gives rise to toxicity are not yet understood to the extent needed to enable the development of disease-modifying treatments for the vast majority of these disorders.

In this context, *in vivo* models can both increase our understanding of the mechanisms of disease and provide a platform for drug screening and preclinical development (Dawson *et al.*, 2018; Götz *et al.*, 2018). A variety of model organisms has been exploited to probe protein misfolding diseases, ranging from simple unicellular systems such as yeast, to more complex ones such as rodents and primates. Simple organisms lack specific aspects of human biology, but can be faster and more cost-effective to work with, making them particularly suitable for high-throughput screens, and for the study of highly conserved cellular pathways (Khurana and Lindquist, 2010; Rincon-Limas *et al.*, 2012; Sin *et al.*, 2014). More complex organisms can recapitulate human

disease phenotypes better, and are typically used in preclinical studies to examine efficacy and safety prior to testing in humans (Jucker, 2010; Sasaguri *et al.*, 2017).

In general, *in vivo* models of disease can be studied at different levels. Organismal and cellular phenotypes can be related to the aggregation states of disease-associated proteins at the molecular level by biochemical and biophysical tools (Table 1). Correspondingly, at the system level, disrupted molecular pathways can be comprehensively monitored by genomics, transcriptomics and proteomics techniques (Barabási *et al.*, 2011). With these approaches, the association of a perturbation (e.g. the expression of a mutant gene) with a phenotype (i.e. an aspect of the disease) is translated into knowledge of specific biochemical processes associated with the disease under investigation. Finally, a comparison with *in vitro* studies can contribute to deciphering the molecular mechanisms of these processes, and represents a powerful strategy for revealing the molecular origins of a disease, and for identifying possible targets for therapeutic interventions (Knowles *et al.*, 2014; Chiti and Dobson, 2017).

Here, we provide an overview of the analysis of protein misfolding in transgenic models of AD and PD based on the transgenic expression of A β or tau, and of α -synuclein, respectively. We furthermore discuss methodological challenges in developing biophysical tools to probe *in vivo* the aggregation process and the resulting aggregate species, which are required to provide quantitative assessments of cause–effect relationships in disease and of the efficacy of potential therapies.

Determination of protein aggregation mechanisms using *in vitro* biophysical methods

Biophysical studies of protein aggregation *in vitro* enable the identification of possible molecular mechanisms of protein

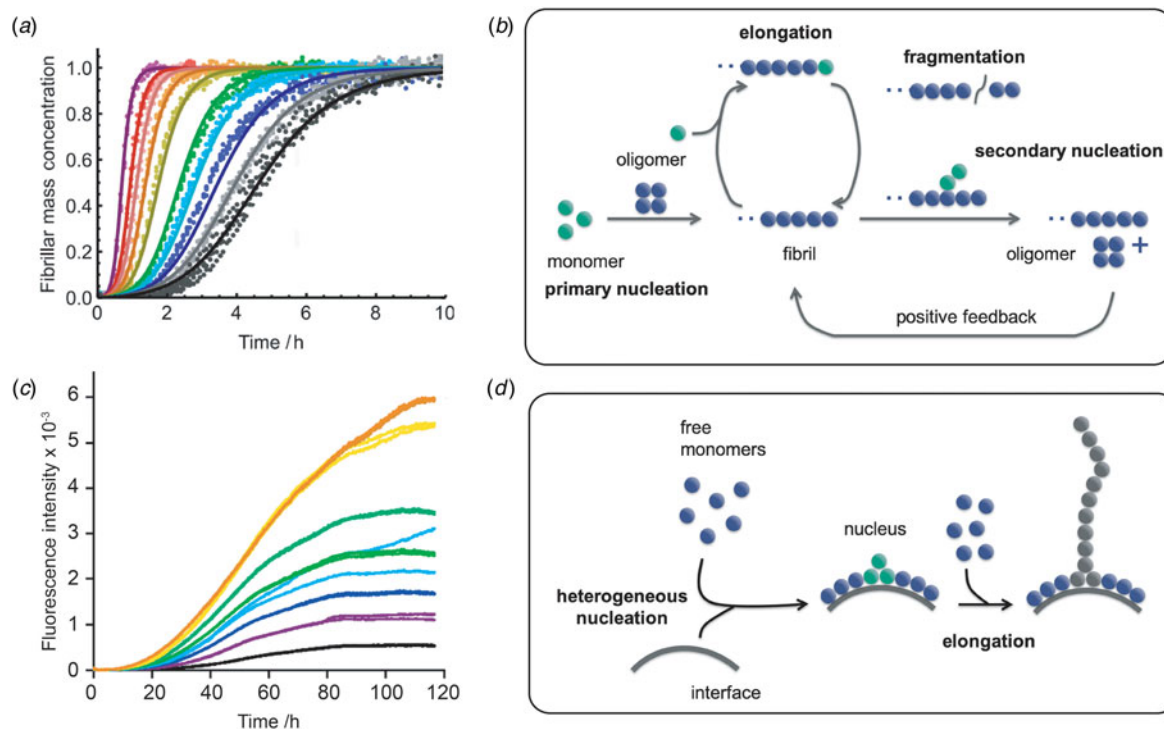


Fig. 1. Mechanisms of amyloid formation *in vitro*. (a) *In vitro* ThT aggregation assay of A β 42 at different starting monomer concentrations. The curves are fitted using a model that includes both primary and secondary nucleation. Figure adapted from Cohen *et al.* (2013b). (b) Schematic representation of the kinetic model of fibril formation including primary and secondary processes. Soluble monomeric forms of proteins undergo primary nucleation to generate oligomeric species that have the potential to convert into fibrils. The formation of fibrils is significantly enhanced by secondary processes that enable aggregates to proliferate by the further association of soluble protein molecules. The latter processes include fragmentation, which generates new fibril ends at which growth occurs, and surface catalysed nucleation, in which fibril surface functions as a template for the generation of new oligomeric and fibrillar species. Figure based on Cohen *et al.* (2013b). (c) ThT aggregation assay of α -synuclein in the presence of different concentrations of dimyristoylphosphatidylserine (DMPS) vesicles (black, 60 μ M; purple, 120 μ M; dark blue, 180 μ M; light blue, 240 μ M; dark green, 300 μ M; light green, 450 μ M; yellow, 600 μ M; orange, 1200 μ M). Figure adapted from Galvagnion *et al.* (2015). (d) Schematic representation of the dominating mechanism of amyloid formation by α -synuclein in the presence of small unilamellar lipid vesicles, which present an interface for nucleation to occur. Figure based on Galvagnion *et al.* (2015).

aggregation *in vivo* and the corresponding neurodegenerative processes (Knowles *et al.*, 2014; Chiti and Dobson, 2017). This approach is powerful not only because *in vitro* systems are amenable to a wider range of technologies compared to *in vivo* systems (Table 1) but also because experimental conditions can be carefully controlled, and the effects of specific perturbations can be examined, e.g. with the introduction of cellular components whose relevance has been revealed by *in vivo* studies.

In particular, the kinetics of the protein aggregation process can be measured quantitatively *in vitro*, leading to a detailed mechanistic understanding of the process itself. Protein aggregation over time is typically monitored by measuring the fluorescence of the amyloid-sensitive dye thioflavin T (ThT). The analysis of the concentration dependence of the time courses of the ThT signal leads to a description of the microscopic steps in the aggregation process and the associated rate constants (Fig. 1a) (Knowles *et al.*, 2009; Cohen *et al.*, 2012; Meisl *et al.*, 2016a; Michaels *et al.*, 2017). Aggregation reactions that start from purely monomeric species require primary nucleation events, which involve the assembly of some monomers into disordered oligomers, which can then undergo a structural conversion into forms that have the capacity to grow by elongation through the addition of more monomers (Michaels *et al.*, 2020). When fibrils are formed, secondary processes play a significant role in a variety of systems, and in some cases can dominate the overall aggregation behaviour (Cohen *et al.*, 2013b; Meisl *et al.*, 2014).

These processes are dependent on the concentration of the existing amyloid aggregates, and include fibril fragmentation as well as secondary nucleation of monomers on fibril surfaces (Fig. 1b). Secondary processes represent a feed-forward loop and cause an exponential proliferation of fibrils (Cohen *et al.*, 2013b; Meisl *et al.*, 2014). It is of great importance to understand the relative contributions of the microscopic steps underlying the aggregation process in the context of disease, especially given that potentially toxic oligomers may arise from both primary and secondary nucleation.

We present here recent insights obtained from *in vitro* experiments on the aggregation mechanisms of A β , tau and α -synuclein, in particular those that can be correlated directly with observations made in *in vivo* models.

A β aggregation

The aggregation of A β into extracellular amyloid plaques is a hallmark of both the familial (early-onset) and sporadic (late-onset) forms of AD. The involvement of amyloid precursor protein (APP) and its sequential processing into A β by β -secretase and γ -secretase (Fig. 2a) is well established through the amyloid hypothesis (Selkoe and Hardy, 2016). In particular, mutations in APP or in the presenilin (PSEN) subunit of γ -secretase are directly linked to familial forms of the disease, and triplication of chromosome 21, on which the APP gene is located, is

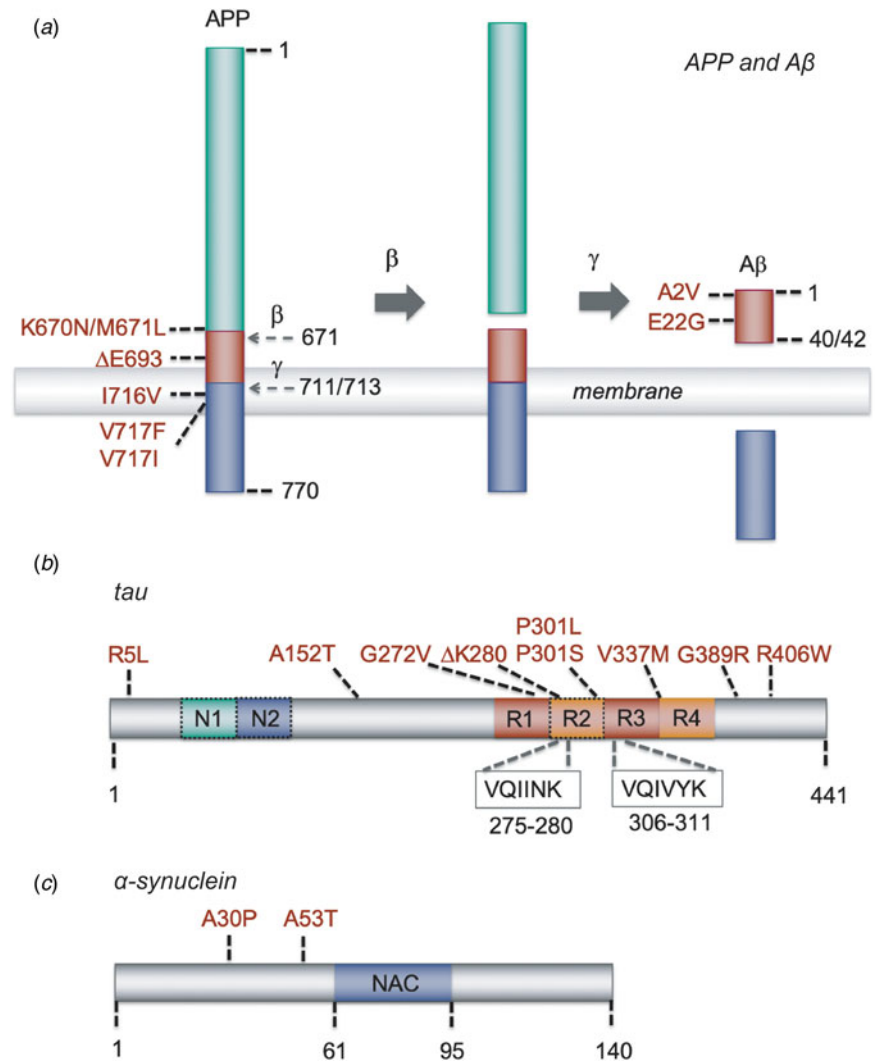


Fig. 2. Schematic representation of A β , tau and α -synuclein, and of the mutations that have been used in the models described in this paper. (a) APP and its sequential processing into the A β peptide by β -secretase and γ -secretase. Cleavage sites are indicated by the residue number at the N-terminal side of the cleavage; mutations are indicated with the residue numbers of full-length APP and those of the A β _{40/42} cleavage product. Other APP processing pathways, such as α -secretase processing, and other A β forms, such as A β ₃₉ or A β ₄₃, are omitted for clarity. (b) The *MAPT* gene encoding tau undergoes splicing to yield six possible isoforms. The full-length 2N4R tau isoform of 441 residues are shown; 1N and 0N isoforms lack the second or both N-terminal inserts, respectively, and 3R isoforms lack the second microtubule repeat (dashed boxes). The hexapeptide stretches VQIINK and VQIVYK, which have the propensity to form β -strands and are essential for the aggregation of tau, are located at the microtubule binding repeats. (c) Schematic representation of α -synuclein, containing three major regions: an N-terminal amphipathic domain with α -helical propensity where several familial mutations are located (residues 1–61), a central hydrophobic part (residues 61–95) known as the NAC region that is essential for amyloid formation, and an acidic C-terminal region (residues 95–140).

associated with the development of AD in patients with Down syndrome (Karran and De Strooper, 2016; Selkoe and Hardy, 2016). The relationship between A β aggregation and the onset and progression of AD has been debated extensively (Herrup, 2015; Karran and De Strooper, 2016; Selkoe and Hardy, 2016), with studies suggesting that amyloid deposition may precede the onset of symptoms (Bateman *et al.*, 2012; Jack *et al.*, 2013; Villemagne *et al.*, 2013), and supporting a relationship between A β oligomers and cellular toxicity (Benilova *et al.*, 2012; Chiti and Dobson, 2017).

Several A β variants of different lengths and with different post-translational modifications exist in the human brain, with the two most common forms being of 40 (A β ₄₀) and 42 (A β ₄₂) residues (Kummer and Heneka, 2014). Studying *in vitro* the aggregation of these different forms of A β , in particular with respect to oligomer formation, has led to the establishment of correlations between aggregation mechanisms and cytotoxic processes, in particular by revealing the importance of secondary nucleation in the proliferation of the aggregates (Cohen *et al.*, 2013b; Meisl *et al.*, 2014) (Fig. 1a and b). A β ₄₂ aggregates faster than A β ₄₀ *in vitro*, with the relative contributions of the microscopic aggregation steps being different for the two isoforms, as in particular the rate of primary nucleation is enhanced for A β ₄₂ (Meisl *et al.*, 2014). From *in vitro* studies, it has furthermore become clear that the

two forms can affect the behaviour of each other, although the details of such interplay still remain unclear (Pauwels *et al.*, 2012; Yu-Jen and Yun-Ru, 2014; Cukalevski *et al.*, 2015; Tran *et al.*, 2017).

Familial mutations linked to early-onset AD include those in the PSENs that affect the cleavage of A β , leading to increased levels of A β ₄₂ in particular. However, some mutations also occur within the A β sequence itself, and these have been found to cause increased aggregation propensities (see <http://www.alzforum.org/mutations>). A study expressing a series of A β ₄₂ mutants in *Drosophila* has found a remarkable correlation between the *in vitro* aggregation rates of the variants and measures of cytotoxicity, namely reduced locomotor function and lifespan, as described in the Section 'A β models' (Luheshi *et al.*, 2007). Using *in vitro* assays, further mechanistic insights have been obtained to probe which of the microscopic steps in the aggregation process are affected for the A β ₄₂ variants. For example, for the A2V A β ₄₂ mutant associated with early-onset AD, it has been shown that secondary nucleation is significantly enhanced compared to that observed for the wild-type form, potentially generating a larger flux of toxic oligomers (Meisl *et al.*, 2016b). In addition, mutations of residues E22 and D23, which are surface-exposed, have been found to lead to enhanced secondary nucleation, whereas the elongation rates are close to that of the wild-type peptide

(Yang *et al.*, 2018). Although the aggregation process may be affected by extrinsic factors *in vivo*, these findings suggest that the surface properties of A β fibrils play an important role in the generation of toxicity.

Given the cytotoxicity of the oligomeric forms of A β (Bucciantini *et al.*, 2002; Baglioni *et al.*, 2006; Haass and Selkoe, 2007; Bemporad and Chiti, 2012; Benilova *et al.*, 2012; Chiti and Dobson, 2017), the structural features of these assemblies are under close investigation. Characterisation of a particular form of such oligomers by solid-state NMR spectroscopy (Table 1) has provided evidence for parallel β -sheet content (Parthasarathy *et al.*, 2015). Other structural studies on samples prepared *in vitro* have provided evidence for the existence of more loosely packed A β 42 oligomers (Ahmed *et al.*, 2010), as well as antiparallel β -sheet oligomers (Cerf *et al.*, 2009; Stroud *et al.*, 2012; Huang *et al.*, 2015). A structural study of A β 40 oligomers stabilised by zinc ions has also revealed a compresence of both parallel and antiparallel β -sheet conformations (Mannini *et al.*, 2018). Mature A β 42 fibrils, on the other hand, have only ever been found to adopt parallel β -sheet conformations as evidenced by recent structure determination using solid-state NMR spectroscopy (Xiao *et al.*, 2015; Colvin *et al.*, 2016; Wälti *et al.*, 2016) and cryo-electron microscopy (cryo-EM, Table 1) (Gremer *et al.*, 2017). A conformational mapping of A β 42 oligomers using a panel of single-domain antibodies rationally designed to scan the whole A β 42 sequence (Aprile *et al.*, 2017) has revealed that early oligomers produced during the aggregation reaction are structurally different from those produced at later stages, and correspondingly are toxic through different mechanisms (De *et al.*, 2019).

Lipid membranes are likely to play a central role in the aggregation process of A β *in vivo*, given that the peptide is generated from the cleavage of APP within lipid bilayers. As such, aggregation is likely to occur in the vicinity of these membranes, whether intracellularly within endosomes or lysosomes, or extracellularly near the plasma membrane. Depending on the types of lipids and the fluidity of the membranes, studies of synthetic liposomes in *in vitro* aggregation reactions of A β have led to a wide range of different results. Lipids in a gel phase have been found to have a stronger inhibitory effect on A β aggregation kinetics than those in liquid-crystalline bilayers (Hellstrand *et al.*, 2010). Furthermore, a relationship may exist between the levels of cholesterol and A β deposition, as the major risk factor for late-onset AD, the presence of a mutation in the *ApoE* gene, is involved in cholesterol metabolism (Mahley, 1988). Incorporation of cholesterol in lipid vesicles in aggregation assays has revealed that cholesterol does indeed promote A β 42 aggregation (Habchi *et al.*, 2018). This effect could be due to the fact that cholesterol modifies the fluidity of the membrane, although a direct molecular interaction between cholesterol and A β 42 has also been reported (Di Scala *et al.*, 2014). Study of the effects of the lipid membrane composition and fluidity in more detail will be important as these properties may be altered during ageing and neurodegeneration, e.g. as a consequence of lipid oxidation, which not only modifies the lipid types but also reduces membrane fluidity (Borst *et al.*, 2000).

Tau aggregation

Intracellular neurofibrillary tangles composed of hyperphosphorylated tau are a hallmark of AD, along with the presence of extracellular amyloid plaques. Tau deposits are observed in a family of neurodegenerative disorders collectively known as

tauopathies, which in addition to AD include Pick's disease, corticobasal degeneration, progressive supranuclear palsy, argyrophilic grain disease and frontotemporal dementia (Lee *et al.*, 2001). In the human brain, the *MAPT* gene encoding tau undergoes alternative splicing leading to six possible isoforms, depending on the number of N-terminal insertions (0N, 1N or 2N) and microtubule binding repeats (3R or 4R) (Fig. 2b) (Goedert *et al.*, 1989). Mutations in *MAPT* are associated with the development of several tauopathies, although specific mutations have not been reported for AD patients.

Tau was originally described as a protein that binds and stabilises microtubules (Weingarten *et al.*, 1975). It is intrinsically disordered in its monomeric form, and has several phosphorylation sites which can be modified to regulate microtubule stability, with hyperphosphorylation being associated with disease (Wang and Mandelkow, 2016). The classical histopathological staging of the disease (Braak and Braak, 1991), as well as recent advances in positron emission tomography (PET) imaging have revealed that tau deposition is more closely related to the symptoms of AD than is the presence of amyloid plaques (Ossenkoppe *et al.*, 2016; Tosun *et al.*, 2017; Xia *et al.*, 2017). However, the mechanisms by which tau causes cellular toxicity are not fully understood, and the relationship with A β accumulation is still under debate (Spires-Jones and Hyman, 2014).

The C-terminal region of tau contains the microtubule binding repeats, which include the short hexapeptide stretches VQIINK and VQIVYK that have a high propensity to form β -strands and are essential for the aggregation of tau (Wang and Mandelkow, 2016). Tau is highly hydrophilic and intrinsically disordered, although it has been shown to adopt preferentially a compact conformation in which the N- and C-terminal regions are in close proximity (Jeganathan *et al.*, 2006). The N-terminal part of tau is known as the projection domain, extending away from the interaction site when tau is bound to microtubules, and away from the fibril surface in the case of fibrillar tau. Given that truncated versions of tau, or constructs comprising only the C-terminal repeat region, have increased aggregation propensity, the N-terminal region is thought to exert a protective effect (Wang and Mandelkow, 2016). Under typical *in vitro* conditions, however, tau does not aggregate spontaneously, but to form fibrils requires the addition of catalysts, as for example polyanionic molecules such as heparin. The negative charges may be required to overcome the repulsion between tau molecules, each of which carries a net positive charge in the C-terminal repeat region, although heparin also induces the aggregation of phosphorylated tau with a net negative charge by an unknown mechanism (Wang and Mandelkow, 2016).

In vitro studies have been instrumental in complementing *in vivo* work to shed light on the role of familial mutations, splicing and phosphorylation in tau aggregation and toxicity. A key finding has been that disease-associated mutations reduce the affinity of tau for microtubules, which may increase the risk of aggregation *in vivo* as the same region of the protein is involved in both processes (Hong *et al.*, 1998). The six tau isoforms have been shown to vary in their aggregation propensity, with the 4R isoforms being more aggregation-prone than the 3R isoforms (Zhong *et al.*, 2012). How this finding relates to pathological conditions is unclear, however, as in AD a mixture of 3R and 4R isoforms is found in tau tangles, whereas in other tauopathies such as Pick's disease and argyrophilic grain disease either the 3R or 4R tau species dominates. A potential explanation could again be provided by post-translational modifications such as phosphorylation.

Out of the 85 theoretical phosphorylation sites in the tau sequence, about 45 have been observed in tau derived from AD brains (Hanger *et al.*, 2009). *In vitro* assays have led to the identification of the potential kinases and phosphatases associated with various sites, and for example GSK-3 has been found to be capable of targeting many sites (Hanger *et al.*, 2009), consistent with its important role in tau toxicity as observed in a variety of animal models as outlined in the Section ‘Tau models’. In light of the complex patterns of phosphorylation, it is not surprising that conflicting results have been obtained on whether hyperphosphorylation of tau promotes or inhibits its aggregation. Hyperphosphorylated tau extracted from AD brains is able to form fibrils *in vitro*, whereas dephosphorylation using alkaline phosphatase inhibits aggregation into fibrils (del Alonso *et al.*, 2001). Conversely, pseudo-phosphorylated constructs comprising the tau repeat region show much slower aggregation *in vitro*, although they may result in the formation of a larger number of oligomers (Kumar *et al.*, 2014).

Directly and quantitatively observing the formation of oligomeric species has been challenging due to their heterogeneous and transient nature, but an elegant single-molecule Förster resonance energy transfer (smFRET, Table 1) assay has yielded important insights into the generation of tau oligomers (Shammas *et al.*, 2015). In this assay, relatively small oligomers were detected early in the aggregation process and these were found to subsequently convert into fibrils. The familial mutants Δ K280 and P301L were not only observed to aggregate faster, but also to form a larger population of oligomers compared to the wild-type protein, the oligomers being of the same apparent size. An anti-aggregation mutant with two proline substitutions that is less toxic in mouse models (Eckermann *et al.*, 2007; Mocanu *et al.*, 2008) was also found to form oligomers, however. Thus, the levels of oligomeric species cannot be directly correlated with toxicity, presumably because the mutations lead to oligomers with distinct structural characteristics (Shammas *et al.*, 2015). In a follow-up study, different types of oligomers could be distinguished by varying the buffer conditions in the smFRET system (Kjaergaard *et al.*, 2018). One type was found to be sensitive to the ionic strength of the buffer, and to be in rapid exchange with tau monomers, whereas the other type was found to be more stable and presumably unable to convert to fibrils (Kjaergaard *et al.*, 2018). Both species may be toxic, but it is likely that they operate *via* different mechanisms.

Recently, liquid–liquid phase separation has attracted attention as a biophysical phenomenon that may act as a precursor to the aggregation of several disease-related proteins (Shin and Brangwynne, 2017; Alberti and Dormann, 2019). Also, the repeat region of tau has been found to condense into liquid droplets *in vitro*, depending on parameters such as the protein concentration, pH value and temperature (Ambadipudi *et al.*, 2017). Phase separation has been observed to induce reversibly the formation of β -strands as detected by circular dichroism (CD, Table 1) spectroscopy. The absence of ThT fluorescence suggests that the phase-separated state does not consist of amyloid fibrils, however, although the addition of heparin to the droplets was found to induce amyloid-like aggregation. NMR spectroscopy (Table 1) has furthermore shown that the conformation of tau is not drastically altered in the phase-separated state, whereas paramagnetic relaxation experiments and ^{13}C -detected experiments have provided evidence for involvement of the repeat regions and the hexapeptide motifs (Ambadipudi *et al.*, 2017, 2019).

Full-length, phosphorylated tau has also been found to phase separate under physiological conditions and to form droplets both *in*

vitro and in neuronal cells in culture (Wegmann *et al.*, 2018). The process has been suggested to be mediated by weak hydrophobic interactions between the hexapeptide repeats, as the liquid droplets were observed to be sensitive to dissociation by urea and by 1,6-hexanediol. The N-terminal region of tau by itself was also observed to phase separate, but in this case driven by electrostatic interactions that can be disrupted by salt. The importance of electrostatics is further underscored by the effect of tau phosphorylation, which was shown to be required for efficient droplet formation. Finally, tau liquid droplets were found to convert rapidly into gels, and after prolonged incubation into non-spherical thioflavin S (ThS) positive aggregates, confirming that tau phase separation can be on pathway to aggregate formation (Wegmann *et al.*, 2018). Whether phase separation drives tau aggregation in human disease remains to be determined, and it will be exciting to study this phenomenon in *in vivo* models, e.g. by using fluorescence lifetime imaging microscopy (FLIM, Table 1) in *Caenorhabditis elegans* (Laine *et al.*, 2019) (see Section ‘ α -Synuclein models’).

α -Synuclein aggregation

The 140-residue protein α -synuclein (Fig. 2c) is the major component of the insoluble intracellular deposits known as Lewy bodies and Lewy neurites that are characteristically found in the brains of PD patients, as well as in those suffering from dementia with Lewy bodies (Spillantini *et al.*, 1998c) and multiple system atrophy (Spillantini *et al.*, 1998a), a set of diseases collectively termed synucleinopathies. PD is associated with the specific loss of dopaminergic neurons in the substantia nigra, which results in impaired motor function, as well as a variety of other deficits including behavioural changes, sleep problems and a diminished sense of smell. Gene duplications and triplications and several point mutations in the SNCA gene, encoding α -synuclein, in hereditary forms of PD have supported a causative role for its aggregation in the disease process (Kalinderi *et al.*, 2016), although the molecular mechanisms by which this pathology occurs are still poorly understood.

Many of the biophysical properties of α -synuclein that have been established through *in vitro* studies have been shown to be valuable in interpreting *in vivo* models. *In vitro* aggregation assays using recombinant α -synuclein have shown that its spontaneous aggregation is negligible at neutral pH, although when preformed seeds are added the reaction can proceed rapidly (Buell *et al.*, 2014). At slightly acidic pH, secondary processes in which existing fibrils catalyse the formation of new ones on their surfaces start to dominate (Buell *et al.*, 2014). Such conditions may be relevant in cellular compartments such as lysosomes and synaptic vesicles. Furthermore, the presence of small unilamellar vesicles in the aggregation assay dramatically increases primary nucleation as α -synuclein binds to the lipid bilayers, which initiates the aggregation process (Fig. 1c and d) (Galvagnion *et al.*, 2015, 2016). In agreement with the propensity of α -synuclein to interact with lipid membranes, genetic screens have systematically identified proteins involved in vesicle trafficking to be related to α -synuclein-mediated toxicity, as described in the Section ‘Biological pathways in Parkinson’s disease models’. These findings do not seem to arise simply as a result of the expression of α -synuclein in non-neuronal cell types such as yeast or *C. elegans* body wall muscle cells, as many of the proteins identified in the screens have been confirmed in neuronal models and in neurons from higher organisms (Cooper *et al.*, 2006; Gitler *et al.*, 2008; Hamamichi *et al.*, 2008).

Although the presence of mature Lewy body pathology does not appear to be essential for toxicity to be observed in the vast majority of *in vivo* models as discussed in the Section ‘ α -Synuclein models’, the process of protein aggregation is likely to play a causative role in synucleinopathies, given that the more aggregation prone α -synuclein variants such as A53T (Conway *et al.*, 2000) or the C-terminal truncation fragment (Crowther *et al.*, 1998) are typically associated with increased toxicity. Data on the A30P mutant are, however, more difficult to interpret, as its aggregation kinetics measured *in vitro* are very similar to those of the wild-type protein (Flagmeier *et al.*, 2016), yet it does not readily form inclusions in yeast (Outeiro and Lindquist, 2003) or in *C. elegans* body wall muscle cells (Perni *et al.*, submitted), although A30P aggregates were demonstrated to form in fly (Feany and Bender, 2000) and rodent (Klein *et al.*, 2002; Lo Bianco *et al.*, 2002; Lauwers *et al.*, 2007; da Silveira *et al.*, 2009) models. These differences may be related to the reduced affinity for lipid bilayers of this latter variant (Bodner *et al.*, 2010). It will be interesting to study these mechanisms in more detail and also to carry out studies *in vivo* of the more recently identified familial PD mutants E46K, H50Q and G51D, which appear to be less aggregation prone *in vitro* (Flagmeier *et al.*, 2016), to shed more light on the aggregation process with respect to toxicity.

Oligomers have been suggested to play a key role in the toxicity of α -synuclein, although their molecular features *in vivo* have remained elusive. Stable α -synuclein oligomers can be generated *in vitro* and are toxic to neuronal cells in culture, causing increased levels of reactive oxygen species (ROS) (Cremades *et al.*, 2012; Chen *et al.*, 2015). Detailed structural examination of stable toxic compared to non-toxic α -synuclein oligomers has revealed that toxicity is linked to strong binding to membrane surfaces and to insertion of a well-structured core into the lipid bilayer, whereas the more disordered species do not enter the bilayer and are not toxic (Fusco *et al.*, 2017). This proposed mechanism of toxicity is interesting in the context of the interference with vesicle trafficking and lipid metabolism that was found in the *in vivo* models of α -synuclein as described in the Section ‘Biological pathways in Parkinson’s disease models’, highlighting the power of connecting the observations made in cellular and animal models with detailed biophysical characterisation *in vitro* to understand the mechanisms behind protein misfolding diseases.

In vivo models of protein misfolding diseases

A common way to create an *in vivo* model of a protein misfolding disease is through the transgenic expression of a human gene associated with the disease in a non-human organism. The gene may typically encode for the protein that misfolds and aggregates, or for a protein that regulates this process. These transgenic models are often based on mutant genes known from familial cases of the disease in order to obtain a strong phenotype with high penetrance. Overexpression is usually employed to obtain high levels of the disease-associated protein, which facilitates the detection of a response in the organism.

Commonly used methods of probing the process of protein misfolding and aggregation in transgenic models include staining with amyloid-binding dyes or conformation-specific antibodies, ultra-structural examination by EM, and biochemical assays based on the solubility, the size or the shape of the misfolded species (Table 1). In parallel, the phenotype of the model can be assessed both at the organismal and the cellular levels, with the goal of correlating the conformational state of the disease-associated protein

to neurotoxicity. In this section, we discuss this approach in the context of yeast, worm, fly and rodent models based on the transgenic expression of A β , tau or α -synuclein (Fig. 3).

A β models

Yeast models

Yeast, in particular *Saccharomyces cerevisiae*, is one of the simplest and most-studied eukaryotic organisms available for the study of cellular pathologies (Verduyck *et al.*, 2016; Tenreiro *et al.*, 2017; Seynnaeve *et al.*, 2018; Rencus-Lazar *et al.*, 2019). This organism has a short generation time, and an arsenal of research tools is available for its study (Smith and Snyder, 2006). *S. cerevisiae* was the first eukaryote to have its complete genome sequenced (Goffeau *et al.*, 1996), and we now know that at least 60% of the *S. cerevisiae* genes have significant homology, or share conserved domains, with human genes (Khurana and Lindquist, 2010). However, as not all homologues have conserved functions, and not all human disease-related proteins have a yeast homologue, an often-used strategy to create a disease model is to express the human disease-related gene in yeast.

Yeast lacks known homologues of APP and β - and γ -secretases, yet transgenic variants of this organism have been successfully employed to investigate APP processing (Zhang *et al.*, 1994, 1997; Lüthi *et al.*, 2003) and to screen for inhibitors of β -secretase (Middendorp *et al.*, 2004) and A β -mediated toxicity (Bharadwaj *et al.*, 2012; Amen and Kaganovich, 2016; Seynnaeve *et al.*, 2018). It has proven challenging, however, to generate yeast models to observe the full A β aggregation process and its associated toxicity (Verduyck *et al.*, 2016; Seynnaeve *et al.*, 2018). Expression of A β 42 in fusion with green fluorescent protein (GFP-A β 42) in this organism was found to result in small punctate patches in newly transformed cells, which become larger as the cells age, and a modest reduction of about 5% in cellular growth was observed (Caine *et al.*, 2007). Cells expressing an A β 42 fusion protein with the MRF domain of the yeast prion Sup35p have been reported as a model system for A β 42 oligomerisation, where a decrease in activity of the reporter was shown to be associated with the formation of sodium dodecyl sulphate (SDS)-stable oligomers (Bagriantsev and Liebman, 2006).

To model the generation of A β in the secretory pathway, experiments have been carried out using the peptide fused to an endoplasmic reticulum (ER)-targeting sequence (Treichel *et al.*, 2011). This model was shown to lead to the accumulation of A β in the secretory pathway, accompanied by growth defects, which were more severe for A β 42 than for A β 40. Consistent with these observations, A β 42 was shown to form more oligomers than A β 40. These observations were confirmed in an independent study in which the expression of A β 42 with the ER-targeting sequence was furthermore shown to lead to impaired mitochondrial function and proteasomal activity (Chen and Petranovic, 2015). Further to these results, expression of an MF α -A β 42-GFP fusion construct that also enters the secretory pathway and causes toxicity was found to be exacerbated for the familial, more aggregation-prone E22G Arctic mutant (D’Angelo *et al.*, 2013), although the aggregation state of the fusion proteins was not examined. An overview of the various yeast models used in the study of A β aggregation is presented in Table 2.

Worm models

The nematode worm *C. elegans* has been used extensively to investigate the molecular processes underlying the cytotoxicity

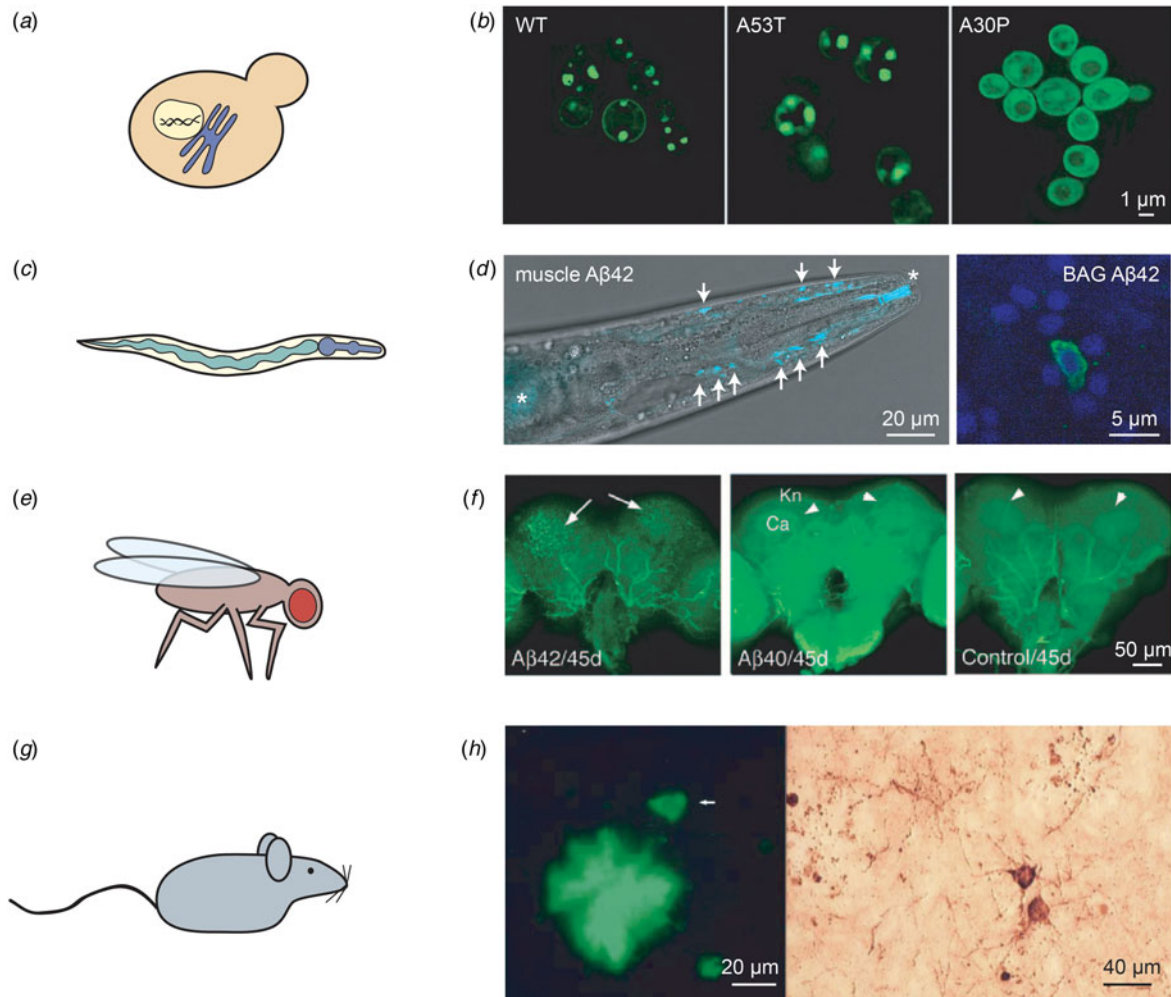


Fig. 3. Examples of *in vivo* models of protein misfolding for the study of AD and PD. (a) The budding yeast *S. cerevisiae* is the most widely used eukaryotic model organism, featuring a conserved cellular organisation, including the nucleus and the secretory system. (b) The expression of two copies of human α -synuclein (either in the wild-type or the familial A53T and A30P forms) in *S. cerevisiae* leads to the formation of cytoplasmic puncta for the wild-type and A53T variants, but not for A30P; reproduced from Outeiro and Lindquist (2003) with permission from *Science*. (c) The nematode worm *C. elegans* is a multicellular organism that consists of several tissues, such as a digestive system comprised of the pharynx and the intestine. (d) Left: *C. elegans* expressing A β 42 in the body wall muscle cells display deposits that can be stained with the amyloid-binding dye X-34. The head region of an A β 42-expressing animal is shown, with deposits indicated by arrows; asterisks mark unspecific staining of the mouth and intestine. Right: Close-up of *C. elegans* expressing A β 42 in the two BAG neurons, stained with an anti-A β antibody (green) and DAPI (blue). Adapted from Sinnige *et al.* (2019). (e) The fruit fly *D. melanogaster* has a more complex brain than *C. elegans*, and possesses a compound eye that is often used as a read-out for toxicity. (f) The expression of A β 42, but not of A β 40, leads to the formation of ThS-positive deposits. The arrows indicate deposits; Kn: Kenyon cell layer; Ca: calyx. Reproduced from Iijima *et al.* (2004); copyright 2004, National Academy of Sciences. (g) Rodent models are typically used for pre-clinical studies, but also provide a more complex system for fundamental studies of human disease mechanisms. (h) Overexpression in mice of human APP and tau with familial mutations leads to the formation of A β deposits positive for ThS staining (left) and structures resembling tau tangles as visualised by Bielschowsky silver staining (right). The arrow in the left panel indicates a neuron. Reproduced from Ribé *et al.* (2005) with permission from Elsevier.

of A β aggregation (Link, 2006; Alexander *et al.*, 2014), and for the discovery of compounds capable of inhibiting such processes (Habchi *et al.*, 2016; Perni *et al.*, 2018). *C. elegans* has the advantage over yeast that it is multicellular, yet has a relatively simple body plan with 959 cells for the adult hermaphrodite, including a nervous system comprised of 302 neurons. The complete genome of *C. elegans* was sequenced in 1998 (The *C. elegans* Sequencing Consortium, 1998), and follow-up studies showed that 38% of the *C. elegans* genes have known human homologues (Shaye and Greenwald, 2011). The nematode typically lives up to 2–3 weeks under laboratory conditions, a convenient experimental timescale, and it has been shown to exhibit age-associated phenotypes, such as deterioration of muscle structure, as observed in higher organisms including humans (Herndon *et al.*, 2002).

Although *C. elegans* contains the APP homologue *apl-1*, this gene lacks a sequence region homologous to human A β . Moreover, as *C. elegans* lacks the β -secretase activity required to process full-length APP, transgenic models have been largely based on the overexpression of the human A β 42 sequence (Table 3). In the first reported model, A β 42 was placed under control of the *unc-54* promoter for expression in the body wall muscle cells (Link, 1995). For efficient expression, a signal sequence upstream of A β was found to be required, and the peptide was shown to be properly processed, yet A β deposits were found in the cytoplasm of the muscle cells rather than in the secretory system or in the extracellular space (Link *et al.*, 2001). However, co-immunoprecipitation has revealed that A β interacts with the ER-homologue of the Hsp70 chaperone in this model,

Table 2. List of yeast models of protein misfolding in AD and PD

Misfolded protein	Construct expressed	Organismal phenotype	Protein aggregation behaviour	Pathways (proteins) involved	Primary references
<i>Alzheimer's disease</i>					
A β	A β 42-Sup35p MRF	Not determined	SDS-stable oligomers	Heat shock response (Hsp104)	Bagriantsev and Liebman (2006)
	A β 42-GFP and GFP-A β 42	Reduced growth rate	Punctate patches; co-localisation with lipids	Heat shock response	Caine <i>et al.</i> (2007)
	A β 42 with an ER targeting sequence	Reduced growth rate	Oligomers not resistant to boiling or SDS	Endocytosis (yeast PICALM)	Treusch <i>et al.</i> (2011)
	A β 42 with an ER targeting sequence	Reduced growth rate	Oligomers	Respiration, proteasomal degradation	Chen and Petranovic (2015)
	MFC α -A β 42-GFP WT and E22G	Reduced viability and growth rate	Accumulation in the secretory pathway	Membrane trafficking (yeast PICALM)	D'Angelo <i>et al.</i> (2013)
Tau	Tau 2N3R and 2N4R	No change in growth rate	Hyperphosphorylated and misfolded tau; small sarkosyl-insoluble fraction	Phosphorylation (yeast GSK-3 β and CDK5)	Vandebroek <i>et al.</i> (2005)
	Tau 2N4R WT, P301L, G272V, N279K, Δ K280, V337M, R406W, S409E, S409A	No change in growth rate	Phosphorylation of S409 increases sarkosyl-insoluble aggregates and higher order oligomers	Oxidative stress; mitochondrial dysfunction	Vanhelmont <i>et al.</i> (2010)
<i>Parkinson's disease</i>					
α -Synuclein (α syn)	α syn-GFP WT, A30P, A53T	Reduced growth rate at higher expression levels; less severe for A30P	Inclusions for WT and A53T co-localising with ubiquitin; A30P diffusely distributed	Proteasomal degradation; lipid metabolism; ER-to-Golgi vesicle trafficking	Outeiro and Lindquist (2003)
	α syn-GFP WT, A30P, A53T	No reduction in growth rate or viability	WT and A53T form inclusions at the plasma membrane; A53T inclusions are ThS-positive; A30P diffuse	Lipid synthesis; proteasome; autophagy; endocytosis; oxidative stress	Zabrocki <i>et al.</i> (2005)
	GFP/GST- α syn WT, A30P, A53T	Reduced viability for overexpression of WT and A53T but not A30P	Inclusions for WT and A53T but not for A30P; GST-fusions of WT and A30P form soluble oligomers	Heat shock response; proteasomal degradation	Dixon <i>et al.</i> (2005)
	GFP- α syn WT, A30P, A53T	Reduced growth rate for all three strains	Inclusions for WT and A53T but not for A30P	Heat shock response; oxidative stress; apoptosis	Flower <i>et al.</i> (2005)

suggesting that it enters the secretory pathway, but is then rerouted to the cytoplasm for degradation (Fonte *et al.*, 2002). Examination using amyloid-specific dyes and immuno-EM has shown that a subset of the deposits is fibrillar, whereas a dimeric A β construct forms deposits but without fibrillar structures (Fay *et al.*, 1998; Link *et al.*, 2001). From a set of mutants designed to interfere with β -strand formation, the L17P and M35C variants also fail to produce deposits that stain with amyloid-sensitive dye ThS (Fay *et al.*, 1998).

Subsequently, it was found that in the original model A β is processed to residues 3-42 (McCull *et al.*, 2009), a fragment that is a major component of human amyloid plaques with the N-terminal residue modified to a pyroglutamate (Harigaya *et al.*, 2000). A model has therefore been created with two extra residues in between the signal sequence and that of A β , resulting in expression of the 1-42 peptide (McCull *et al.*, 2012). In both strains, amyloidogenic protein aggregation (Fig. 3d, left panel) results in paralysis of the worms within several days of adulthood, yet the strain expressing A β (1-42) is more temperature sensitive and shows increased paralysis at higher temperatures, perhaps

reflecting differences in the aggregation behaviour of the two isoforms (McCull *et al.*, 2012).

A model in which A β (3-42) is expressed in neurons features oligomeric species recognised by the NU-4 antibody, and displays a phenotype with impaired chemotaxis (Wu *et al.*, 2006) and defects in associative learning (Dosanjh *et al.*, 2010). Interestingly, these worms are defective in sensing benzaldehyde, but not diacetyl, suggesting that the vulnerability of the associated sensory pathways towards A β toxicity differs. In a more recent model expressing the corrected A β (1-42) variant throughout the nervous system, the protein was found to accumulate in the insoluble fraction during ageing, whereas neuromuscular defects and behavioural abnormalities were observed earlier on (Fong *et al.*, 2016). Expressing A β specifically in glutamatergic neurons results in visible neurodegeneration of the glutamatergic neurons in the tail of the worm (Treusch *et al.*, 2011). However, a model designed in our own laboratory that selectively expresses low levels of A β 42 in the two BAG sensory neurons, which are also glutamatergic, shows moderate behavioural defects in a CO₂ sensing assay without overt signs of neurodegeneration, perhaps because

Table 3. List of worm models of protein misfolding in AD and PD

Misfolded protein	Protein expressed	Tissue	Organismal/cellular phenotype	Protein aggregation behaviour	Pathways (genes) involved	Primary reference	
<i>Alzheimer's disease</i>							
A β	A β 3-42 with signal peptide	Body wall muscle cells	Paralysis phenotype	ThT and X-34 positive deposits, fibrils in EM	Molecular chaperones (ER Hsp70, sHSPs)	Link (1995)	
	A β 1-42 with signal peptide	Body wall muscle cells	Paralysis phenotype	ThT and X-34 positive deposits	Molecular chaperones (Hsp70, Hsp90, TRiC/CCT and others); lysosomal pathways (subunit of vacuolar ATPase); protein synthesis (ribosomal subunits)	McColl <i>et al.</i> (2012)	
	A β 3-42 with signal peptide	Neurons	Chemotaxis and learning impairments	Oligomeric species recognised by NU-4 antibody		Wu <i>et al.</i> (2006)	
	A β 1-42 with signal peptide	Neurons	Neuromuscular defects, reduced food sensing	Insoluble protein observed at old age	Metabolism, mitochondrial function	Fong <i>et al.</i> (2016)	
	A β 42 with signal peptide	Glutamatergic neurons	Neurodegeneration of the tail glutamatergic neurons	ND		Treusch <i>et al.</i> (2011)	
	A β 1-42 with signal peptide	BAG neurons	Subtle modulation of behavioural response to CO ₂	No ThT or X-34 staining observed	Calcium signalling	Sinnige <i>et al.</i> (2019)	
Tau	Tau 1N4R WT, P301L and V337M	Neurons	Degeneration of axons, uncoordinated phenotype (stronger for mutants)	Phosphorylation, accumulation of insoluble tau (more/faster for mutants)	Protein phosphorylation; chaperones; proteasomal degradation	Kraemer <i>et al.</i> (2003)	
	Tau 3R and 4R WT, 4R P301L and 4R R406W	Mechanosensory neurons	WT slightly impaired touch response; mutants strong age-related decline in touch response; neurodegeneration; loss of microtubules	Accumulation in cell bodies and neurites; no signs of aggregation	Kinases (GSK3); chaperones (Hsp70)	Miyasaka <i>et al.</i> (2005)	
	Tau 3R WT, PHP (phosphomimic)	Neurons	Partial uncoordinated phenotype and reduced thrashing; PHP more gaps in axons	Protein in detergent-soluble but not insoluble fraction		Brandt <i>et al.</i> (2009)	
	tau 1N4R V337 + tau F3 Δ 280 fragment	Neurons	Worsened locomotory phenotype and neuronal abnormalities compared to tau V337M alone; perturbed mitochondrial transport	ThS-positive inclusions		Fatouros <i>et al.</i> (2012)	
Amyloid- β + tau	A β 42 + tau F3 Δ 280 fragment	Neurons	Shortened lifespan and worse defects in egg laying and chemotaxis compared to single transgenic lines; dopaminergic neurodegeneration	More Congo red positive deposits compared to single-transgenic lines	Protein phosphorylation; ageing	Wang <i>et al.</i> (2018)	
<i>Parkinson's disease</i>							
α -Synuclein (α syn)	α syn-YFP	Body wall muscle cells	Motility defect	Cytoplasmic inclusions, largely mobile until old age	Vesicle trafficking; lipid metabolism	van Ham <i>et al.</i> (2008)	
	α syn-GFP	Body wall muscle cells	Not determined	Cytoplasmic inclusions	PD-related pathways (pink, parkin); endolysosomal pathways; autophagy	Hamamichi <i>et al.</i> (2008)	
	α syn WT, A53T	Neurons	Dopaminergic neuronal loss; motor deficit	Rare inclusions		Lakso <i>et al.</i> (2003)	
	α syn WT	Dopaminergic neurons	Neurodegeneration	No inclusions observed	Molecular chaperones (torsinA)	Cao <i>et al.</i> (2005)	
	α syn WT, A53T, A30P	Dopaminergic neurons	Loss of dendrites and behavioural defects in A53T and A30P	Occasional clump/dot like distribution		Kuwahara <i>et al.</i> (2006)	
	α syn WT, A53T, A30P	Neuronal	No motility or development phenotype	Not determined		Vesicle trafficking	Kuwahara <i>et al.</i> (2008)
	α syn WT-YFP, designed proline mutants	Muscle cells	Not determined		WT forms elongated structures; triple proline mutant remains diffuse		Karpinar <i>et al.</i> (2009)

these neurons are more resistant to toxic protein aggregation than others (Sinnige *et al.*, 2019). Notably, in this model A β 42 appears to be localised mainly in the cell body (Fig. 3d, right panel) and does not visibly form aggregates, supporting this notion.

Fly models

Drosophila melanogaster, the fruit fly, is a common model organism for neurodegenerative diseases (Rincon-Limas *et al.*, 2012; Prüßing *et al.*, 2013) (Table 4), whose main advantage over *C. elegans* is its more sophisticated nervous system. The central nervous system of the fly comprises glia as well as neurons, and contains specialised sub-structures such as a compound eye and mushroom bodies for olfactory learning and memory, both of which are often employed in disease models. Limitations for the modelling of neurodegenerative disorders, on the other hand, are the absence of an adaptive immune system, and a lack of blood vessels.

Similar to *C. elegans*, *Drosophila* lacks a sequence region corresponding to human A β in its APP homologue. A β -secretase homologue does exist (Carimine-Simmen *et al.*, 2009), but has low levels of activity (Fossgreen *et al.*, 1998; Greeve *et al.*, 2004). γ -Secretase activity has also been demonstrated by expression in *Drosophila* of a construct comprising the A β region, the transmembrane region and the C-terminal segment of human APP, leading to the generation of a 4 kDa A β fragment that may correspond to either A β 40 or A β 42 (Fossgreen *et al.*, 1998). A transgenic model for A β overproduction has been designed in which human APP is expressed together with human β -secretase in the eye, resulting in the generation of a 4 kDa A β peptide and progressive degeneration of the retina and neuropil regions (Greeve *et al.*, 2004). ThS-positive inclusions with a star-like appearance have been found in older flies, yet neurodegeneration precedes the formation of these amyloid deposits. Co-expression of human APP and β -secretase with *Drosophila* PSEN containing mutations corresponding to those in familial AD was found to reduce the onset of amyloid deposition remarkably, from 68 days to 37 days, and to cause more severe neurodegeneration (Greeve *et al.*, 2004).

Expression of human APP and β -secretase throughout the *Drosophila* nervous system was found to result in A β deposition in the cortical layer, as shown by antibody staining, and memory defects in young flies, followed by extensive neurodegeneration in older flies (Sarantseva *et al.*, 2009). Using a similar approach, a model has been generated that displays a more severe phenotype, including abnormal wing morphology at the time of eclosion and strong neuroanatomical changes, suggesting a developmental defect. In this model, punctate A β deposits, which stain with the amyloid-binding dye X-34, are present in relatively young flies (Chakraborty *et al.*, 2011). The reason for the discrepancy between these two models has been suggested to lie in the different caloric intake of the flies, which may affect γ -secretase processing (Chakraborty *et al.*, 2011). Raising the flies at lower temperatures was found to result in a milder phenotype, with A β deposits accumulating during ageing, accompanied by impaired climbing and memory functions and visible neurodegeneration (Mhatre *et al.*, 2014).

To avoid the complications associated with APP processing, another approach has been exploited in which A β 40 and A β 42 are directly expressed following the pre-proenkephalin signal peptide, which was shown to be cleaved correctly (Finelli *et al.*, 2004). Flies expressing A β 42 have a rough eye phenotype and the peptide accumulates in the insoluble protein fraction, whereas flies

expressing the less aggregation-prone A β 40 appear normal (Finelli *et al.*, 2004). The lines that were created have varying expression levels of A β 42 in the eye, depending on the integration site of the construct and the number of copies. Aggregation and toxicity were found to be proportional to the expression levels, with low A β 42 levels showing small deposits and modest neurodegeneration, and higher levels resulting in larger deposits and more severe morphological defects (Finelli *et al.*, 2004).

Expression of A β 42 throughout the nervous system leads to locomotory and other behavioural deficits in relatively young flies, in addition to a dramatically reduced lifespan (Finelli *et al.*, 2004). Further characterisation of this model has revealed that A β 42 forms deposits in various regions of the brain, a subset in the region of the Kenyon cell bodies being ThS positive (Fig. 3f) (Iijima *et al.*, 2004). However, fibrils could not be identified by using EM, suggesting that A β 42 forms diffuse, relatively unstructured deposits rather than mature amyloid plaques in this model. Similar experiments with A β 40 have shown that it accumulates in the same brain areas, but deposits are not visible. Although both A β 40 and A β 42 expression cause age-associated learning defects, only A β 42 results in locomotory defects and neurodegeneration at later ages. This observation suggests that visible protein aggregation is required in this model to observe neurodegeneration, but not neuronal dysfunction, which could perhaps be mediated by oligomers (Iijima *et al.*, 2004).

Using the signal peptide of the *Drosophila necrotic* gene, lines have been generated expressing A β 40 and A β 42, and also A β 42 with the Arctic mutation E22G in the *Drosophila* nervous system (Crowther *et al.*, 2005). All A β variants were found initially to accumulate in intracellular deposits, which react with an antibody specific to oligomeric A β (Crowther *et al.*, 2005). In older flies, A β 42 shows extracellular deposits accompanied by vacuolar structures, which are exacerbated in the E22G mutant, whereas they do not occur in A β 40 flies. The extracellular deposits do not stain with Congo red, indicating that they do not consist of mature amyloid fibrils, but might resemble diffuse plaques as in the previously described fly model (Iijima *et al.*, 2004). The A β 42 flies have a rough eye phenotype and locomotory defects, which occur before the appearance of the extracellular deposits, and the phenotype is exacerbated by the Arctic mutation. Furthermore, the A β 42 flies have a shorter lifespan than controls, whereas the survival of the A β 40 flies is indistinguishable from controls. The phenotype of the A β 42 flies can be partially restored by treatment with the amyloid-specific dye Congo red, suggesting that toxicity is directly caused by A β 42 aggregation.

In a subsequent study, a panel of A β 42 mutants with different aggregation propensities has been expressed in the *Drosophila* nervous system. The computationally predicted aggregation rates were found to correlate strongly with lifespan and locomotory function, especially when the propensity to form protofibrillar (oligomeric) aggregates was considered (Luheshi *et al.*, 2007). Similarly, whereas expression of A β 40 itself is relatively harmless to the flies, point mutations that increase the formation of prefibrillar A β 40 species *in vitro* result in a toxic phenotype when expressed in *Drosophila* (Brorsson *et al.*, 2010). Furthermore, an engineered affibody has been shown to restore the phenotype of A β 42 flies, by binding to the monomer and targeting it for degradation (Luheshi *et al.*, 2010). On the other hand, the KW1 antibody, which is specific to A β 40 oligomers, induces aggregation into toxic protofibrillar species, shortening the lifespan of the flies (Wacker *et al.*, 2014). Further evidence for the role of intermediate species in toxicity has been provided by the expression of

Table 4. List of fruit fly models of protein misfolding in AD and PD

Misfolded protein	Construct expressed	Tissue	Organismal/cellular phenotype	Protein aggregation behaviour	Pathways (genes) involved	Primary reference
<i>Alzheimer's disease</i>						
A β	APP + β -secretase	Eye	Neurodegeneration of retina and neuropil of the optical lobe	ThS-positive deposits in retina and neuropil with a star-like appearance in EM		Greeve <i>et al.</i> (2004)
	APP WT, K670N/M671L + β -secretase	Neurons	Age-associated memory defects and neurodegeneration	SDS stable A β oligomers; A β deposition in cortical layer		Sarantseva <i>et al.</i> (2009)
	APP + β -secretase	Neurons	Impaired climbing and memory; age-related neurodegeneration; developmental defects when raised at higher temperature	A β deposition; punctate X-34 positive deposits when raised at higher temperature		Chakraborty <i>et al.</i> (2011), Mhatre <i>et al.</i> (2014)
	A β 42 ^a	Eye, neurons	Rough eye phenotype; locomotory and learning defects; shortened lifespan; neurodegeneration	ThS-positive deposits; A β mostly insoluble	Secretory pathways; cholesterol homeostasis; regulation of chromatin structure and function; immune response (Toll)	Finelli <i>et al.</i> (2004), Iijima <i>et al.</i> (2004)
	A β 42 WT and E22G	Neurons	Rough eye phenotype; neurodegeneration; locomotor defects; shortened lifespan	Intracellular A β accumulation; diffuse extracellular deposits positive for oligomer antibody but not for Congo red; E22G forms SDS-resistant tetramers		Crowther <i>et al.</i> (2005)
	16 variants of A β 42	Neurons	Locomotor defects and shortened lifespan	Not determined experimentally; good correlation between severity of phenotype and predicted aggregation propensity		Luheshi <i>et al.</i> (2007)
	Tandem A β 40, tandem A β 42	Eye, neurons	Tandem A β 42 causes rough eye phenotype and reduced lifespan; tandem A β 40 no effect	Both form SDS-insoluble aggregates; tandem A β 42 forms soluble oligomeric species		Speretta <i>et al.</i> (2012)
	A β 42 WT, E22G, L17P	Neurons	Learning and locomotory defects; shortened lifespan	Intracellular ThS-positive deposits; oligomers for WT and E22G		Iijima and Iijima-ando (2008)
	A β 42 WT, E22G	Giant fibre neurons	Locomotory defects; shortened lifespan; failed synaptic transmission; depletion of presynaptic mitochondria and synaptic vesicles	Accumulation in the cell bodies and axons	Synaptic transmission	Zhao <i>et al.</i> (2010)
Tau	Tau WT, R406W	(Cholinergic) neurons	Shortened lifespan; age-related neurodegeneration	Misfolded and phosphorylated tau; no fibrils found	Protein phosphorylation; cell adhesion	Wittmann <i>et al.</i> (2001)
	Tau 2N4R WT + GSK3- β	Eye	Neurodegeneration starting from larval stage	Flame-shaped aggregates; PHFs and SFs in EM; tangles positive for phosphorylated tau	Autophagy and lysosomal pathways; RNA processing; chromatin regulation; molecular chaperones; cytoskeleton	Jackson <i>et al.</i> (2002)

	Tau 0N4R WT	Glia	Reduced lifespan; increased apoptosis	Glial inclusions of phosphorylated, ThS and Bielschowsky silver positive tau; fibrils in sarkosyl-insoluble fractions in aged flies		Colodner and Feany (2010)
	Tau 0N4R WT, R5L, S320F, S352L, G389R, R406W	Neurons, glia	Various types of cellular stress	Inclusions of phosphorylated tau	Apoptosis; oxidative stress (JNK signalling); autophagy; UPR	Bardai <i>et al.</i> (2018)
Amyloid- β + tau	A β 42 + tau 1N4R WT, R406W	(Motor) neurons	Exacerbated abnormal axon morphology, locomotory defects and shortened lifespan compared to single transgenics	Stronger tau phosphorylation; aggregation not examined		Folwell <i>et al.</i> (2010)
<i>Parkinson's disease</i>						
α -Synuclein (α syn)	α syn WT, A30P, A53T	Neurons	Loss of dopaminergic neurons; climbing impairment	Lewy body-like round inclusions with radiating fibrils, thread and grain-like structures (EM)		Feany and Bender (2000)
	α syn WT, A30P, A53T	Dopaminergic neurons	Loss of dopaminergic neurons	Lewy body-like and Lewy neurite-like staining; α syn mostly soluble in high salt/detergent buffer	Heat shock response (Hsp70)	Auluck <i>et al.</i> (2002)
	α syn WT	Neurons	Widespread neurodegeneration; reduced lifespan; locomotory defects	Ubiquitinated α syn deposits	Actin cytoskeleton; apoptosis	Ordonez <i>et al.</i> (2018)
	α syn WT, Δ 71-82, 1-120	Neurons	No neurodegeneration for Δ 71-82, slightly increased for 1-120	Δ 71-82 no inclusions, more PK sensitive protein and less oligomers than WT; 1-120 more inclusions and increased PK resistance compared to WT		Periquet <i>et al.</i> (2007)
	α syn S129A, S129D	Neurons	S129A suppresses dopaminergic neuronal loss; S129D enhances retinal degeneration	S129A more PK resistance, larger and more inclusions, more protein in sarkosyl-insoluble fraction; S129D similar to WT		Chen and Feany (2005)
	α syn WT, A30P, A53T, A56P and triple P mutant	Neurons	Triple P mutant reduces lifespan and motility most strongly; degeneration of dopaminergic neurons	Triple P mutant is predicted to increase the formation of oligomers	A53T: mitochondrial proteostasis (TRAP1)	Karpinar <i>et al.</i> (2009)

^aAll A β 40/42 expressed using signal peptide.

tandem constructs of A β 40 and A β 42, in which two monomers are connected with a linker to increase the effective concentrations and aggregation rates. Expression of both tandem constructs leads to the formation of deposits in the fly brain, but only the tandem A β 42 flies show a dramatic reduction in lifespan relative to those expressing the monomeric peptide, an effect that is associated with an increase in SDS-soluble species (Speretta *et al.*, 2012).

A β 42, the E22G Arctic mutant, as well as a designed mutant with lower aggregation propensity have furthermore been expressed in the *Drosophila* nervous system using the pre-proenkephalin signal peptide (Iijima *et al.*, 2008). In this study, the A β variants were found to be localised intracellularly, residing in the ER, Golgi and lysosomes, but in contrast to the model by Crowther and co-workers, no oligomer-specific antibody staining was observed. None of the lines was found to show Congo red staining, demonstrating that mature fibrils are not noticeably formed either. The Arctic mutant shows an increased population of dimers and trimers by western blot, a larger number of ThS-positive deposits in the cell bodies accompanied by neuronal loss, a shortened lifespan and stronger locomotory defects than the model expressing wild-type A β 42. The designed mutant, however, forms more ThS-positive deposits in the neurites leading to neuropil loss, and has a longer lifespan, yet shows learning defects similar to the Arctic mutant. These discrepancies have been suggested to arise from a differential intracellular distribution of the variants, in addition to their different aggregation propensities, highlighting the importance of the biological environment in mediating cellular toxicity (Iijima *et al.*, 2008).

Further studies on flies expressing A β variants in the giant fibre neurons have demonstrated that intracellular A β , which accumulates in cell bodies and axons, leads to impairments in synaptic function, accompanied by reduced numbers of mitochondria and synaptic vesicles (Zhao *et al.*, 2010). Again, the severity of the phenotype was found to be correlated with expression levels and to be stronger for the Arctic mutant. In addition, it has been shown that levels of the synaptic protein bruchpilot and presynaptic calcium channels decline with age in flies expressing the Arctic mutant, compared to controls (Huang *et al.*, 2013). This *Drosophila* model expressing A β in a subset of neurons has thus proved to be suitable to study perturbations at the single-neuron level, and has provided strong evidence for a connection between A β accumulation and synaptic dysfunction even in this relatively simple model organism.

Rodent models

One of first-transgenic mouse models of AD shown to develop amyloid plaques resembling those in the human disease relied on the overexpression of the V717F mutant of human APP, which is associated with early-onset AD, driven by the platelet-derived growth factor- β (PDGF- β) promoter (Games *et al.*, 1995). These mice, referred to as PDAPP from the combination of PDGF- β and APP, show ThS-positive extracellular plaques accompanied by dystrophic neurites and reactive astrocytes. The plaques vary in morphology from round to diffuse. The pathology is age-related, with plaques first appearing between 6 and 9 months of age, and increasing in number as the mice age further. This model displays synaptic loss correlated with increased plaque deposition in older mice (Dodart *et al.*, 2000) as well as learning defects (Chen *et al.*, 2000), but without overt neuronal loss (Irizarry *et al.*, 1997).

A wide variety of models based on mutant APP overexpression has been subsequently generated, which similarly display amyloid plaques and are associated with synaptic dysfunction and gliosis (reviewed in Elder *et al.* (2010); Hall and Roberson (2012); Drummond and Wisniewski (2017); see <http://www.alzforum.org> for a full list of available rodent models and Table 5 for those discussed here). These models differ from one another in various ways including the ratio of A β 40 to A β 42, the morphology of the plaques, the age at which they appear and the brain regions that are affected. A widely used model is the Tg2576 mouse, in which human APP is overexpressed with the Swedish double mutation K670N/M671L using the prion protein (PrP) promoter that drives strong expression in neurons (Hsiao *et al.*, 1996). In this model, dense ThS-positive deposits are found in older mice (11–13 months), a subset of which displays Congo red birefringence. Dystrophic neurites have also been found in this case, as well as reactive astrocytes and microglia. At the age where plaques are apparent, the mice show behavioural impairment in the Morris water maze test. However, there is no direct correlation between the quantity of plaques and cognitive defects, as the behavioural phenotype remains stable between 6 and 14 months, while the plaque load continues to increase. Rather, the levels of a 56 kDa soluble A β species, referred to as A β *56, identified in extracts from Tg2576 mice by SDS-PAGE and size exclusion chromatography (SEC) (Table 1) correlate with memory impairment (Lesné *et al.*, 2006). Furthermore, this species affects long-term memory when purified and infused into young rats, supporting the hypothesis that A β oligomers are responsible for neurotoxicity in AD.

A complication of APP-based transgenic mouse models is that its processing gives rise to several variants of the A β peptide, as well as other intracellular and extracellular fragments of APP (Fig. 2a), which may all contribute to the observed phenotype. To address specifically the role of A β accumulation, a different approach has been taken in expressing constructs comprising the BRI protein fused to either A β 40 or A β 42, which are released by the furin cleavage site directly preceding the A β sequence (McGowan *et al.*, 2005). Expression of the A β 40 construct leads to high levels of free peptide, which however remains soluble, so that no amyloid plaques are observed. In contrast, BRI-A β 42 mice show amyloid pathology starting at 3 months of age, including the presence of diffuse and compact ThS-positive plaques, as well as deposits in blood vessels in the brain, termed cerebral amyloid angiopathy. However, although dystrophic neurites and reactive astrogliosis have been observed surrounding the plaques, this study reported that the mice have no behavioural abnormalities, suggesting that A β 42 deposition by itself is not sufficient to trigger neurotoxicity, at least up to the age investigated (McGowan *et al.*, 2005). It is interesting to compare these findings to those from the *Drosophila* and *C. elegans* models discussed earlier, where A β fusions do lead to behavioural phenotypes. These observations can perhaps be attributed to toxicity arising from intracellular rather than extracellular A β in these worm and fly models. Intracellular A β may also be present in APP-based mouse models (LaFerla *et al.*, 2007), but not in those developed using the BRI fusion approach.

Mutations in the PSEN genes *PSEN1* and *PSEN2* associated with early onset forms of AD have been employed to increase the levels of the more aggregation-prone A β 42 in mouse models. Expression of familial mutant, but not wild-type, *PSEN1* has been shown to increase specifically the levels of A β 42 over A β 40 in mice (Borchelt *et al.*, 1996; Duff *et al.*, 1996; Citron *et al.*,

Table 5. List of rodent models of protein misfolding in AD and PD

Misfolded protein	Name of the model	Construct expressed	Organismal/cellular phenotype	Protein aggregation behaviour	Pathways (genes) involved	Primary reference
<i>Alzheimer's disease</i>						
A β	<i>Mouse</i>					
	PDAPP	APP V717F	Dystrophic neurites and reactive astrocytes; synaptic loss and learning defects	ThS-positive extracellular plaques starting from 6–9 months		Games <i>et al.</i> (1995)
	Tg2576	APP K670N/M671L	Dystrophic neurites and reactive astrocytes and microglia; impairment in Morris water maze	ThS-positive plaques at 11–13 months, some of which are Congo red birefringent; soluble species A β *56	Immune response (progranulin); intracellular trafficking (Klc-1)	Hsiao <i>et al.</i> (1996)
	BRI-A β 40/42	BRI-A β 40/42	Dystrophic neurites and reactive astrocytes around A β 42 plaques, but no behavioural abnormalities	ThS-positive plaques for A β 42; deposits in blood vessels; no aggregation for A β 40	Immune response (progranulin); A β and lipid metabolism (ApoE)	McGowan <i>et al.</i> (2005)
	PS2APP	APP K670N/M671L + PSEN2 N141I	Dystrophic neurites and reactive astrocytes and microglia; mild deficiencies in synaptic plasticity and in Morris water maze	Congophilic A β plaques at 9 months		Richards <i>et al.</i> (2003)
	APP ^{SL} PS1KI	APP K670N/M671L/V717I + PSEN1 M233T/L235P	Neuronal loss in hippocampus; impairment in working memory and motor tasks	ThS-positive plaques and intracellular A β deposits		Casas <i>et al.</i> (2004)
	APPPS1	APP K670N/M671L + PSEN1 L166P	Dystrophic neurites and reactive glia; cognitive impairment at 8 months	Congophilic A β plaques starting from 6 weeks	Immune response (progranulin); mitochondrial function	Radde <i>et al.</i> (2006)
	5xFAD	APP K670N/M671L/I716V/V717 + PSEN1 M146L/L286V	Neuronal degeneration and loss; deficits in Y-maze test at 4–5 months	Small and numerous ThS-positive plaques from 2 months; intracellular A β puncta from 1.5 months	Immune response; cell differentiation; neurogenesis; histone modification; transcription; regulation of calcium binding; synaptic plasticity	Oakley <i>et al.</i> (2006)
	APP ^{NL-F} and APP ^{NL-G-F}	APP K670N/M671L/I716V (/E693E) knock-in	Neuroinflammation, synaptic alterations, memory impairment in Y-maze test	A β plaques first observed at 6 months for NL-F mice, and at 2 months for NL-G-F		Saito <i>et al.</i> (2014)
	<i>Rat</i>					
McGill-R-Thy1-APP	APP K670N/M671L/V717I	Degenerating glutamatergic and cholinergic neurons surrounding plaques; learning and memory deficits at 3 months; limited neuronal loss at 18 months	ThS-positive plaques; intracellular A β accumulation; soluble trimeric A β species	Stress response; synaptic function; mitochondrial function; amino acid metabolism	Leon <i>et al.</i> (2010)	
TgF344-AD	APP K670N/M671L + PSEN1 Δ exon9	Inflammation and phagocytosis of neuronal debris; significant neuronal loss; deficits in behavioural assays	Widespread ThS-positive plaques at 16 months; intracellular A β 42; soluble oligomeric A β species; Gallyas silver tau aggregates		Cohen <i>et al.</i> (2013a)	
Tau	<i>Mouse</i>					
	htau	Genomic human tau in mouse tau knock-out background	Memory impairment; significant neuronal loss between 8 and 18 months	Phosphorylated and misfolded tau; fibrillar tau in somatodendritic compartments resembling pre-tangle state; PHFs in sarkosyl-insoluble fraction		Andorfer <i>et al.</i> (2003)
	Tau4RTg2652	Tau 1N4R WT	Abnormal axon morphologies and dystrophic neurites; slight motor impairment; memory deficit in Barnes maze	Phosphorylated and misfolded tau; low levels of oligomers; no tangles		Wheeler <i>et al.</i> (2015)

(Continued)

Table 5. (Continued.)

Misfolded protein	Name of the model	Construct expressed	Organismal/cellular phenotype	Protein aggregation behaviour	Pathways (genes) involved	Primary reference
	JNPL3	Tau 0N4R P301L	Motor and behavioural deficits	Tau tangles in brain and spinal cord that are phosphorylated and ThS, Congo red and Gallyas silver positive; fibrils in EM; sarkosyl-insoluble material in aged mice	Anti-apoptotic factors; microtubule-dependent intracellular trafficking; immune response	Lewis <i>et al.</i> (2000)
	pR5	Tau 2N4R P301L	Reactive astrocytes; apoptotic neurons	ThS and Gallyas silver positive tangles; granular accumulations of phosphorylated and misfolded tau; short fibrils in sarkosyl-insoluble fraction	Advanced glycated end products (glyoxalase I); mitochondrial dysfunction and ROS; synaptic function	Götz <i>et al.</i> (2001a)
	rTg4510	Tau 0N4R P301L	Atrophy in cortex and hippocampus; impairments in Morris water maze test; locomotory deficits	Misfolded and hyperphosphorylated tau; tangles; sarkosyl insoluble material; fibrils observed by using EM	Insolubility of kinases, enzymes, chaperones	Ramsden <i>et al.</i> (2005)
	PS19	Tau 1N4R P301S	Activated microglia and astrocytes; neuronal inflammation; neurodegeneration	Phosphorylated and insoluble tau; ThS, Congo red and Gallyas silver tangles, globular and thread-like structures	Cytoskeleton (GTPase-regulatory proteins)	Yoshiyama <i>et al.</i> (2007)
	TauΔK280	Tau 2N4R ΔK280	Synaptic loss	Misfolded and phosphorylated tau; partially insoluble tau; tangles at old age		Eckermann <i>et al.</i> (2007)
	TauRDΔK280	Tau 0N4R ΔK280	Astroglia; loss of synapses and neurons	Hyperphosphorylated and insoluble tau; Gallyas silver positive tangles; fibrils in sarkosyl-insoluble fraction		Mocanu <i>et al.</i> (2008)
	htau40AT	Tau 2N4R A152T	Reactive astrocytes and microglia; loss of synapses and neurons; neuronal hyperexcitability; memory deficits	Hyperphosphorylated, misfolded tau; Gallyas silver and ThS-positive tangles; sarkosyl-insoluble tau	Autophagy and lysosomal pathways	Decker <i>et al.</i> (2016), Sydow <i>et al.</i> (2016)
	hTau-A152T	Tau 1N4R A152T	Reactive astrocytes; behavioural deficits; neuronal hyperexcitability; age-related neuronal loss	Misfolded, soluble tau		Maeda <i>et al.</i> (2016)
	<i>Rat</i>					
	3R tau151-391	Tau 3R residues 151–391	Shortened lifespan; no neuronal loss	Phosphorylated, misfolded and sarkosyl-insoluble tau; ThS and Congo red positive tangles		Filipcik <i>et al.</i> (2012)
Amyloid-β + tau	<i>Mouse</i>					
	TAPP	APP K670N/M671L + tau 0N4R P301L	Inflammation and dystrophic neurites surrounding Aβ plaques as in Tg2576; motor symptoms as in JNPL3	Increased tau tangles compared to JNPL3; no change in Aβ deposition		Lewis <i>et al.</i> (2001)
	3xTg-AD	APP K670N/M671L + tau 0N4R P301L + PSEN1 M146V	Synaptic dysfunction	Intracellular Aβ followed by plaques; misfolded and hyperphosphorylated tau; ThS and Gallyas silver positive tau tangles	Oxidative phosphorylation; ion transport; apoptosis; mitochondrial protein synthesis	Oddo <i>et al.</i> (2003)
	PDAPP/PS19	APP V717F + tau 1N4R P301S	Stronger motor phenotype and reduced lifespan compared to PS19	Spread of hyperphosphorylated tau in brain regions following Braak staging; accelerated formation of ThS-positive tau tangles compared to PS19		Hurtado <i>et al.</i> (2010)
	5xFAD/PS19	APP K670N/M671L/I716V/V717I + PSEN1 M146L/L286V + tau 1N4R P301S	Increased inflammation, loss of synapses and neuronal death compared to PS19 or 5xFAD	Increased hyperphosphorylated and misfolded tau compared to PS19		Saul <i>et al.</i> (2013)

TauPS2APP	APP K670N/M671L + PSEN2 N141I + tau 2N4R P301L	Mild cognitive impairment; no observable neuronal damage or loss	Accumulation of SDS-stable A β species as in PS2APP; phosphorylated tau spreading with age; Gallyas silver positive tau tangles; PHF-like fibrils in EM	Oxidative phosphorylation	Rhein <i>et al.</i> (2009); Grueninger <i>et al.</i> (2010)	
PLB1 triple	APP K670N/M671L/V717I + PSEN1 A246E + tau P301L/R406W	Impaired synaptic plasticity; cognitive defects; altered glucose metabolism; sleep disturbances	ThS and Congo red positive A β plaques; phosphorylated tau starting at the same age of 6 months		Platt <i>et al.</i> (2011)	
APP ^{swe} /tau ^{VLW}	APP K670N/M671L + tau 2N4R G272V/P301L/R406W	More neuronal loss than single-transgenic lines	Increased tau phosphorylation and accumulation in sarkosyl-insoluble fraction compared to tau ^{VLW} ; increased plaque load compared to Tg2576		Pérez <i>et al.</i> (2005); Ribé <i>et al.</i> (2005)	
Tau/PS1/APP	APP K670N/M671L/V717I + PSEN1 M146L + tau 0N3R WT	Dystrophic neurites; inflammation	Phosphorylated tau surrounding A β plaques but no tangles		Boutajangout <i>et al.</i> (2004)	
APP/PS1/rTg21221	APP K670N/M671L + PSEN1 Δ exon9 + tau 0N4R WT	More dystrophic neurites compared to APP/PS1 but no increased loss of synapses or neurons	Increased area of ThS-positive amyloid deposits; misfolded and phosphorylated tau but no tangles		Jackson <i>et al.</i> (2016)	
APPOSK-Tg/tau264	APP Δ 693 + tau 3R/4R WT	Accelerated synaptic loss, memory impairment and neuronal loss compared to single-transgenic lines	Intracellular and oligomeric A β ; phosphorylated tau; Gallyas positive tau tangles; fibrillar tau in EM		Umeda <i>et al.</i> (2014)	
App ^{NL-(G-)F} /MAPT dKI	APP K670N/M671L/I716V (/E693E) + MAPT knock-in	No effect on inflammation or memory compared to App ^{NL-G-F} model	Accelerated phosphorylation of tau compared to MAPT knock-in only, but no tangles		Saito <i>et al.</i> (2019)	
APP/PS1 x rTauEC	APP K670N/M671L + PSEN1 Δ exon9 + tau 0N4R P301L	Increased neurodegeneration and neuronal loss	Increased spread of misfolded tau from the entorhinal cortex to distal brain regions; more amyloid plaques than in APP/PS1		Pooler <i>et al.</i> (2015)	
<i>Parkinson's disease</i>						
α -Synuclein (α syn)	<i>Mouse</i>					
	Line D	α syn WT	Loss of dopaminergic neuron terminals in striatum; motor defect	Nuclear, ER-associated and cytoplasmic granular inclusions	Masliah <i>et al.</i> (2000)	
	Line 61	α syn WT	Loss of dopamine; motor symptoms; olfactory deficits	PK-resistant inclusions	Membrane signalling (Gnal, Pde10a, Itpr1); extracellular matrix remodelling and inflammation (Hspg2, Lama5 and Lamb2)	Rockenstein <i>et al.</i> (2002)
	M83	α syn A53T	Motor defects; no loss of dopaminergic neurons	Fibrillar and ThS-positive protein		Giasson <i>et al.</i> (2002)
	hm ² α -SYN	α syn A30P + A53T	Locomotor defects; loss of dopaminergic neurons	No cytoplasmic inclusions identified	Mitochondrial function and oxidative stress; ubiquitin-proteasome system; ER stress	Richfield <i>et al.</i> (2002)
	Syn130m	α syn WT 1-130	Specific loss of dopaminergic neurons in <i>substantia nigra</i> , presumably during development	No LB-like structures identified by different staining methods or western blot		Wakamatsu <i>et al.</i> (2008)
	SNCA-OVX	Genomic α syn WT	Locomotor defects; loss of dopaminergic neurons; reduced dopamine release; increased clustering of vesicles at dopaminergic synapses	High-molecular weight species but no PK-resistant protein		Janezic <i>et al.</i> (2013)

(Continued)

Table 5. (Continued.)

Misfolded protein	Name of the model	Construct expressed	Organismal/cellular phenotype	Protein aggregation behaviour	Pathways (genes) involved	Primary reference
	A53T transgenic	syn A53T	Locomotor defects; loss of dopaminergic neurons; reduced dopamine release; fragmentation of Golgi apparatus	Inclusions found at axons and in vacuolar structures in dopaminergic neurons	Lysosomal and autophagic pathways	Lin <i>et al.</i> (2012)
	3K	Genomic <i>syn</i> E35K + E46K + E61K	Progressive locomotor defects; dopaminergic neurodegeneration; partially restored with L-DOPA	Granular inclusions of PK-resistant <i>syn</i> with S129 phosphorylation and C-terminal truncation; co-localisation with vesicles at synaptic terminals; lipid-rich accumulation		Nuber <i>et al.</i> (2018)
	<i>Rat</i>					
	BAC transgenic	Genomic <i>syn</i> WT	Changes in novelty-seeking and avoidance behaviour; reduced sense of smell; locomotor defects; dopaminergic neurodegeneration	Accumulation of insoluble and PK-resistant protein; C-terminally truncated <i>syn</i> in insoluble fraction		Nuber <i>et al.</i> (2013)

1997), and crossing these lines with APP mutant mice has led to accelerated A β deposition (Borchelt *et al.*, 1997; Holcomb *et al.*, 1998). Several PSAPP mouse models based on different combinations of APP and PSEN1/2 mutants exist, and all show similar pathologies to that of the APP-transgenic mice, but at an earlier age and accompanied by a stronger behavioural phenotype. They also suffer from greater neuronal loss, although not at levels comparable to the widespread degeneration in human AD (McGowan *et al.*, 2006; Elder *et al.*, 2010; Hall and Roberson, 2012). For example, the PS2APP mouse model, a cross of the APP K670N/M671L mutant and PSEN2 with the N141I mutation, displays congophilic amyloid plaques at 9 months of age, which are surrounded by dystrophic neurites and activated microglia and astrocytes (Richards *et al.*, 2003). However, deficiencies in synaptic plasticity and behavioural tests such as the Morris water maze are relatively mild (Richards *et al.*, 2003).

In the APP^{SL}PS1KI mouse model, which expresses APP with V717I and K670N/M671L mutations, and PSEN1 with M233T and L235P mutations, large numbers of plaques are already apparent at 6 months of age, and about 50% neuronal loss in the CA1/2 layer of the hippocampus has been observed at 10 months of age (Casas *et al.*, 2004). The loss of neurons appears to correlate better with the presence of intracellular A β aggregates that can be stained with ThS than with extracellular plaques, which is interesting given evidence for the presence of intracellular A β in human AD (LaFerla *et al.*, 2007). Coinciding with this pathology, the mice become increasingly impaired in working memory and motor tasks from 6 months of age (Wirths *et al.*, 2008). Apart from the fact that this model produces A β 42 as the dominant A β species, another interesting feature in relation to human AD is the progressive N-terminal truncation and modification of A β 42 into species that have been detected in patient material (Casas *et al.*, 2004).

A model with very extensive amyloid deposition has been created using the L166P mutant form of PSEN1, which has led to a disease onset as young as 24 years of age in humans, crossed with a mouse strain expressing APP K670N/M671L (Radde *et al.*, 2006). In the resulting model, congophilic amyloid plaques appear in the neocortex at 6 weeks, whereas deposits in different regions of the hippocampus occur between 2 and 5 months. As in the other mouse models, the plaques are surrounded by dystrophic neurites and glia, but cognitive impairment is only evident at 8 months (Radde *et al.*, 2006).

The approach of combining APP and PSEN mutants has been further extended through the generation of a mouse strain with a total of five familial AD mutations, the 5xFAD mouse, which has become one of the most frequently used models in AD research (Oakley *et al.*, 2006). Constructs of APP with K670N/M671L, I716V and V717I mutations, and PSEN1 with M146L/L286V, were co-injected to create this model, resulting in the rapid accumulation of A β 42 and ThS-positive plaques from 2 months of age. The plaques are smaller but more numerous than in the Tg2576 mouse, consistent with an increase in nucleation events due to the higher concentration of A β 42. In this model, intracellular A β aggregation also appears to play a role, as indicated by small ThS-positive puncta in pyramidal neurons. It has been speculated that extracellular plaques could arise from these aggregates upon cell death, as indeed degenerating neurons with intracellular deposits are sometimes found in the vicinity of plaques. These regions of co-localisation are correlated with synaptic degeneration and significant loss of neurons. At the behavioural level, there is no phenotype at 2 months in these mice, but at 4–5

months, memory defects are apparent in the Y-maze test (Oakley *et al.*, 2006).

Mouse models based on the overexpression of human genes produce artificially high protein levels, including other APP fragments in addition to A β , altogether complicating the interpretation of the phenotype. Second-generation mouse models remedy some of these issues by making use of knock-in strategies to produce human A β in the context of endogenous mouse APP (Sasaguri *et al.*, 2017). For example, the A β sequence of mouse APP was humanised, and Swedish (K670M/M671L) and Iberian (I716F) mutations were introduced in order to generate high levels of preferentially A β 42 (Saito *et al.*, 2014). The resulting APP^{NL-F} mice display progressive A β pathology and inflammation, accompanied by memory impairment in the Y-maze test. Adding the Arctic mutation E693G has resulted in the APP^{NL-G-F} model that displays earlier A β deposition and a stronger inflammatory response.

Another strategy to avoid overexpression consists in triggering an amyloid cascade response by intracerebral infusion of A β seed aggregates produced either *in vitro* or derived from cell systems, other animal models or AD patients (Jucker and Walker, 2018). These non-transgenic mouse models are particularly useful in the study of the mechanisms of propagation of the aggregates across cells and brain tissues, but the infusion process tends to generate inflammatory reactions that complicate the interpretation of the results.

Given that rats are evolutionarily more closely related to humans than mice, and display more complex patterns of behaviour, rat models have certain advantages for recapitulating and testing the pathogenic mechanisms behind AD. Although early rat models transgenic for APP did not display amyloid plaques, probably due to insufficient levels of expression (reviewed in Do Carmo and Cuellar, 2013), the McGill-R-Thy1-APP model has been successful in generating many aspects of AD pathology with just a single transgene of APP combining Swedish (K670N/M671L) and Indiana (V717F) mutations (Leon *et al.*, 2010). These rats accumulate intracellular A β from just 1 week after birth, and the first amyloid deposits show at 6 months in a particular brain region, the subiculum. Investigations in older animals have specifically identified degenerating glutamatergic and cholinergic neurons surrounding compact, ThS-positive plaques, showing the vulnerability of these types of neurons. Furthermore, the levels of soluble trimeric A β species are greatly elevated in homozygous rats, and were found to correlate with learning and memory deficits at 3 months of age (Leon *et al.*, 2010). This observation points towards toxicity of soluble A β species, rather than mature amyloid plaques. This model thus recapitulates many features of human AD, although a subsequent study has shown that neuronal loss is in fact very limited, being largely restricted to the subiculum at 18 months of age (Hegglund *et al.*, 2015).

Another useful rat model, TgF344-AD, has been developed by expressing K670N/M671L mutant APP and PSEN1 with a deletion of exon 9 (Cohen *et al.*, 2013a). At 16 months of age, the rats display widespread deposition of ThS-positive amyloid plaques in the cingulate cortex and hippocampus, as well as the striatum and cerebellum, all of which are regions affected by amyloid deposition in AD patients. The model also features intraneuronal accumulation of A β 42, and soluble oligomeric A β species have been found which correlate with significant neuronal loss; in addition the model displays inflammation and phagocytosis of neuronal debris. At the behavioural level, the rats show defects in various assays, and these progress with age following the

development of the observed pathology. In contrast to most APP/PSEN-based mouse models, this rat model displays tau pathology consisting of Gallyas silver-stained tangles both in vicinity of the amyloid plaques, and also in regions devoid of plaques, as observed in human AD (Braak and Braak, 1991). The presence of tau tangles is critical to study the interplay between A β and tau pathology, which is still poorly understood, as outlined in the next sections.

Tau models

Yeast models

Yeast models have been employed to compare the *in vivo* aggregation propensities of various tau isoforms and disease-associated mutations, as well as the role of specific phosphorylation sites (Table 2). Upon expression of the human tau isoforms 2N3R and 2N4R in yeast, a small fraction of both protein constructs accumulates in the sarkosyl-insoluble aggregate fraction, with the 2N4R isoform being more extensively phosphorylated than the 2N3R isoform (Vandebroek *et al.*, 2005). Comparison of a set of six familial point mutants in the 2N4R isoform showed that all become hyperphosphorylated, and accumulate more strongly in the sarkosyl-insoluble fraction than the wild-type protein (Vanhelmont *et al.*, 2010). Purified hyperphosphorylated, soluble wild-type tau quickly forms structured filaments *in vitro*, and is able to seed tau isolated from transgenic yeast cells (Vandebroek *et al.*, 2005). The P301L tau variant found in FTDP-17 patients, purified from yeast, has been shown to aggregate *in vitro* when presented with a microtubule surface as a substrate, in contrast to wild-type tau-4R (Vandebroek *et al.*, 2006). Although yeast models have thus confirmed key aspects of tau phosphorylation and aggregation, no toxicity has been observed in any of the reported expression systems as judged from the growth rates of the cells (Vandebroek *et al.*, 2005; De Vos *et al.*, 2011).

Worm models

The first *C. elegans* models of tauopathy have been based on the overexpression of a construct comprising human tau 1N4R in nerve cells (Kraemer *et al.*, 2003). In addition to that expressing the wild-type protein, lines expressing the mutants P301L and V337M, which are associated with frontotemporal dementia and parkinsonism linked to chromosome 17 (FTDP-17), have been generated, all resulting in an uncoordinated phenotype with reduced thrashing rates and shortened lifespan. Tau becomes phosphorylated at the majority of known sites in these models, and it accumulates in the detergent-soluble fraction as the worms age, as well as in the detergent-insoluble fraction in the case of the mutant lines. In addition to presynaptic defects in cholinergic transmission, degeneration of GABAergic axons has been observed. However, tau was found to be diffusely localised in the axons, and only the V337M variant showed clear deposits in electron micrographs. This mutant line contains the highest level of insoluble tau, and displays the strongest phenotype, but the results obtained on wild-type and P301L tau suggest that the formation of deposits is not required for neurodegeneration (Kraemer *et al.*, 2003).

A study in which the 3R and 4R isoforms of wild-type tau, and the 4R isoforms of P301L and R406W were expressed specifically in the six mechanosensory neurons of *C. elegans* has largely corroborated these findings (Miyasaka *et al.*, 2005). The touch response is only mildly affected in worms expressing wild-type tau, but declines strongly with age in the mutant strains, both

of which show corresponding signs of neurodegeneration. Ultrastructural examination by using EM has shown accumulation of tau in the cell bodies and neuronal processes with concomitant loss of microtubules, yet no evidence for fibrillar aggregates was found.

Expression of different tau variants with increased or decreased aggregation propensities has, however, highlighted the relevance of toxic protein aggregates in the *C. elegans* models. Co-expression of V337M tau with a short aggregation-prone tau fragment, F3ΔK280, leads to the formation of ThS-positive inclusions, and increased accumulation of full-length tau in the detergent-soluble and detergent-insoluble fractions during ageing compared to the strain expressing V337M tau alone (Fatouros *et al.*, 2012). Concomitantly, the locomotory phenotype is more severe, the neurons display more morphological abnormalities, and the transport of mitochondria is perturbed. The introduction of mutations predicted to prevent the formation of β -strands in the F3ΔK280 fragment has the opposite effect, and in addition compounds known to inhibit tau aggregation have been shown to reduce the accumulation of insoluble tau and partially restore the locomotory phenotype of the strain expressing the aggregation-prone fragment (Fatouros *et al.*, 2012).

Fly models

The first observation of tau toxicity in flies was serendipitous, when a reporter construct consisting of *Drosophila* tau tagged with GFP was expressed in sensory neurons with the aim to study axonal development. Morphological defects were observed, providing the first demonstration of tau toxicity in the fly, and similar results were obtained upon the expression of human tau (Williams *et al.*, 2000). Since then, *Drosophila* has been used extensively to study tau aggregation and toxicity, as well as the effects of phosphorylation by different kinases (see Sun and Chen (2014) for a more complete overview, and Table 4 for the models here discussed). As in yeast and *C. elegans*, the effects of FTDP-17 mutations have also been examined, for example in models expressing human wild-type and R406W tau forms throughout the nervous system (Wittmann *et al.*, 2001). Although both of these strains display a reduction in lifespan, the phenotype is stronger for the R406W mutant, and severe neurodegeneration is evident from vacuolar structures in the cortex. Both the wild-type and R406W tau forms are recognised by antibodies specific to pathogenic phosphorylation and misfolded conformations of tau, but evidence for fibrillar structures has not been found in investigations by EM.

Co-expression of the *Drosophila* homologue of the kinase GSK-3 β with full-length tau in the fly eye does result in the characteristic flame-shaped aggregates resembling those found in human tauopathies (Jackson *et al.*, 2002). The antibody AT100, which is specific for phosphorylated tau in tangles, recognises tau only when co-expressed with GSK-3 β in the fly. Co-expression of the kinase results in fibrillar aggregates with the dimensions of human paired helical filaments (PHFs) and straight filaments (SFs) as observed by EM, and exacerbates neurodegeneration compared to the expression of tau alone (Jackson *et al.*, 2002). In agreement with the toxic effects of phosphorylation by GSK-3 β , inhibition of the kinase restores the locomotory phenotype in a model expressing human wild-type tau 0N3R in motor neurons (Mudher *et al.*, 2004). Another kinase, PAR-1, has been shown to phosphorylate tau upstream of GSK-3 β , and its overexpression exacerbates the rough eye phenotype of the R406W model, but in the absence of visible tau aggregates (Nishimura *et al.*, 2004).

In certain tauopathies, fibrillary tangles occur in glia in addition to neurons (Lee *et al.*, 2001). A *Drosophila* model in which human 0N4R tau is expressed specifically in adult glia shows a reduction in lifespan and the presence of apoptotic signals in glia and, by non-cell autonomous mechanisms, in cholinergic neurons (Colodner and Feany, 2010). In aged flies, tau is found in sarkosyl-insoluble fractions, which contain straight fibrils as shown by EM studies. Immunohistochemistry has confirmed that the glia contain inclusions consisting of phosphorylated tau, which are furthermore positive for ThS and Bielschowsky silver staining, and contain fibrillar structures in EM. Expression of tau in both glia and neurons leads to increased toxicity compared to that in neurons alone, as measured by the number of apoptotic cells, indicating the importance of glial tau in these neurodegenerative disorders.

In a recent study, a comparison of a panel of *Drosophila* models expressing wild-type tau and five FTDP-17-linked mutations at similar expression levels has further strengthened the correlation between tau phosphorylation and neurotoxicity (Bardai *et al.*, 2018). Mutants that display the strongest aggregation, indicated by the number of inclusions *in vivo* and ThS fluorescence of the corresponding recombinant proteins *in vitro*, show the least caspase activation and induction of stress signalling, demonstrating that mature aggregates do not cause significant toxicity. The mutants generating the highest toxicity, on the other hand, have the highest phosphorylation levels and show increased F-actin accumulation, suggesting a correlation between actin stabilisation and toxicity (Bardai *et al.*, 2018).

Rodent models

As in the case of APP-based mouse models, initial rodent models overexpressing human wild-type tau constructs have not displayed clear behavioural phenotypes, nor produced significant numbers of tau tangles, which is why most models have been generated using tau with mutations found in FTDP-17 (reviewed in Hutton *et al.*, 2001; Lee *et al.*, 2005). An overview of about 30 current transgenic mouse models of tau is available at <http://www.alzforum.org>; we highlight here those that are commonly used, or that have led to particularly interesting insights (Table 5).

One of the few successful models based on wild-type tau is the htau model, a cross of a strain expressing human tau, including all regulatory regions and leading to all splicing variants, with a knock-out strain for endogenous mouse tau (Andorfer *et al.*, 2003). Expression of the human construct by itself does not lead to clear tau pathology (Duff *et al.*, 2000), suggesting that the interplay between mouse (which only occurs as 4R) and human isoforms is important. In the htau model, tau was observed to be correctly localised in the axons in 6-week-old animals, but tau staining was found to shift towards the cell bodies as the animals age, accompanied by increased phosphorylation at particular sites. Beyond 9 months, tau aggregation was inferred from immunohistochemistry and immuno-EM, and the sarkosyl-insoluble fraction was found to contain PHFs with similar dimensions to the ones found in AD brains (Andorfer *et al.*, 2003). The mice suffer significant neuronal loss at old age, between 8 and 18 months, but the majority of dying neurons does not appear to contain fibrillar aggregates (Andorfer *et al.*, 2005). However, memory impairments and synaptic defects occur at 12 months, but not at 4 months, indicating that tau pathology does need to develop to a certain stage to cause damage (Polydoro *et al.*, 2009).

A more recent model based on wild-type tau, Tau4RTg2652, has been generated to overexpress high levels of 1N4R wild-type tau, driven by the Thy1.2 promoter, resulting in approximately 12-fold overexpression relative to endogenous mouse tau (Wheeler *et al.*, 2015). In this model, misfolded and phosphorylated tau is detectable by various antibodies in the forebrain, amygdala, hippocampus and brainstem even at 3 months, but tangles are not observed by ThS and Gallyas staining even at 24 months of age. The levels of oligomeric tau as recognised by the TOC1 antibody are relatively low, but abnormal axon morphologies and dystrophic neurites have been nevertheless identified throughout the brain. The mice show a slight motor phenotype, but the most obvious behavioural phenotype is an impairment in spatial memory as assessed in the Barnes maze test. Altogether, these data suggest that pre-tangle tau, consisting of altered conformations and phosphorylation states, but not oligomers or fibrils, could be sufficient to drive neuronal dysfunction. Both the tau pathology and the behavioural phenotype remain stable from 3 months onwards, which may make this model suitable for rapid studies of the early stages of sporadic tauopathies, including AD.

Most other existing tau models are based on tau mutations linked to FTDP-17, which are not directly related to AD, but provide strong phenotypes, which are useful for fundamental studies of tau pathology, as well as for preclinical trials targeted at halting tau toxicity. The JNPL3 mouse, overexpressing tau 0N4R with the P301L mutation from the PrP promoter, shows impaired motor function and abundant tau tangles in several regions of the brain as well as the spinal cord; the latter are, however, typically not associated with AD (Lewis *et al.*, 2000). The tau accumulations are recognised by a range of conformational and phospho-specific antibodies, and are positive for Congo red, ThS and other staining agents such as Gallyas silver. Different morphologies have been found, including the flame-shaped tangles characteristic of AD, but also aggregates resembling Pick bodies and small, dense inclusions similar to those found in FTDP-17. EM studies have confirmed the presence of fibrils, and sarkosyl-insoluble protein has been found in extracts of brain and spinal cord from aged JNPL3 mice, but not from control lines expressing wild-type human tau.

In another mouse model, known as pR5, the P301L mutant is expressed in the context of the longest tau isoform 2N4R under the control of the Thy1.2 promoter (Götz *et al.*, 2001a). In this model, tau also accumulates in various brain regions, and forms tangles stained by ThS and Gallyas silver in e.g. the spinal cord and layers 5 and 6 of the cortex. Reactive astrocytes and apoptotic neurons have been found in the same regions. Other neurons, including pyramidal cells in the hippocampus, show granular tau accumulation that is recognised by a subset of phospho- and conformation-specific antibodies. The sarkosyl-insoluble fraction of brain extracts contains fibrils as observed by using EM, but they are shorter and narrower than fibrils obtained from AD patient material. This is in line with the observation that P301L fibrils from FTDP-17 patients have a different morphology from that of V337M FTDP-17 fibrils or wild-type AD fibrils (Spillantini *et al.*, 1998b), suggesting that the point mutations can significantly affect the fibril structure. Furthermore, the strong overexpression in mouse models may give rise to shorter fibrils than those observed in AD, because the likelihood of primary nucleation is higher.

The rTg4510 model has also been designed to express P301L tau (0N4R), in this case using an elegant expression system that

can be inhibited with doxycycline (Ramsden *et al.*, 2005; SantaCruz *et al.*, 2005). The Ca²⁺/calmodulin kinase II (CaMKII) promoter has been used to drive expression specifically in the forebrain, where the first tau tangles appear at 2.5 months of age, leading up to extensive tau pathology and increasing atrophy in the cortex and hippocampus from 5 months onwards (Ramsden *et al.*, 2005). Misfolded tau conformations and phosphorylation also develop with age, as indicated using various antibodies, and the main tau species shifts from the soluble 55 kDa form at 2.5 months to the sarkosyl-insoluble, extensively phosphorylated 64 kDa species at 8.5 months. At the ultrastructural level, SFs have been observed using EM on fixed tissue samples from 10-month-old animals, which occasionally adopt a typical herring bone pattern which has also been observed in deposits in Pick's disease. At the behavioural level, spatial memory in the Morris water maze test is impaired from 4 months of age, and locomotory deficits have been observed in 10-month old mice (Ramsden *et al.*, 2005).

The possibility of lowering tau expression in this model has led to interesting insights into the relationship between tau pathology and toxicity (SantaCruz *et al.*, 2005). Suppressing tau expression at 2.5 months causes the pathology to be halted. In contrast, when doxycycline treatment is given at 4 months or later, tau phosphorylation and accumulation in the insoluble fraction proceed, and numbers of tau tangles grow similarly as in untreated animals. Given the significant reduction in tau mRNA, this suggests that the soluble species that were already present can develop into pathological tau. However, brain atrophy and memory defects were found to be largely restored if the doxycycline treatment was sufficiently long, demonstrating that the presence of tau tangles is not directly correlated with the disruption of cognitive function (SantaCruz *et al.*, 2005).

A model expressing tau 1N4R with the P301S mutation, PS19, shows a strong motor phenotype which presents difficulties for behavioural studies, yet it has been a useful model because of its robust tau pathology and evident neurodegeneration (Yoshiyama *et al.*, 2007). Phosphorylated tau has been shown to accumulate rapidly, and increased levels of insoluble tau have been observed during ageing. At 6 months of age, flame-shaped tau tangles and globular aggregates, as well as thread-like structures, can be visualised in the hippocampus, amygdala and spinal cord by Gallyas silver staining, and by staining with ThS and Congo red. Activated microglia and astrocytes, as well as neuronal inflammatory markers, develop from 3 months onwards. Tau pathology and inflammation precede large scale neurodegeneration, which is observed from 9 months onwards in the hippocampus and other brain regions. However, hippocampal synapses are already lost by 3 months of age, preceding the appearance of tau tangles. As such, the results obtained from this model add to the notion that neurodegeneration is initiated before mature protein aggregates are formed.

The ΔK280 mutation, which is known from a FTDP-17 case, is directly correlated with the aggregation propensity of tau, being located in a segment of the protein that forms a β-strand in the fibril structure. Mouse models based on this pro-aggregation mutant have been generated in parallel with an anti-aggregation mutant consisting of ΔK280 combined with two proline mutations (I277P and I308P), which strongly inhibit β-strand formation (Eckermann *et al.*, 2007; Mocanu *et al.*, 2008). These mutants have been expressed in the context of full-length human tau 2N4R (Eckermann *et al.*, 2007), and in a short construct comprising only the repeat domain of tau (Mocanu *et al.*,

2008) in two respective studies, leading to very similar results. The pro-aggregation mutant adopts a misfolded conformation that is recognised by the MC1 antibody, becomes phosphorylated at several sites and is partially found in the sarkosyl-insoluble fraction, whereas the anti-aggregation mutant does not display these features. Furthermore, the pro-aggregation mutant has been shown to induce co-aggregation of endogenous mouse tau, which co-localises with the exogenous human tau in the same filaments extracted from the mouse brain. In both studies, quantification from electron micrographs has shown that the pro-aggregation mutant causes more synaptic loss than the anti-aggregation mutant. The strong correlation between *in vitro* and *in vivo* aggregation propensities is furthermore underscored by the fact that the truncated construct leads to Gallyas-positive tau tangles much earlier than full-length tau, in line with a protective effect of the flanking regions on tau aggregation observed *in vitro*. These models also make use of the regulatable expression system, and switching off tau expression has been shown largely to reverse tau pathology (Eckermann *et al.*, 2007; Mocanu *et al.*, 2008), in contrast to the earlier study which reported that tau tangles continue to develop after reducing transgene expression, although memory function recovered (SantaCruz *et al.*, 2005). However, phosphorylation and aggregation of endogenous mouse tau persist in the pro-aggregation model, possibly seeded by the pathological human tau expressed prior to switching off the transgene (Mocanu *et al.*, 2008).

More recently, mouse models have been generated expressing tau with the A152T mutation, which has been found to increase the risk of FTD as well as AD. In one model, the mutation has been introduced in the context of full length tau 2N4R (Decker *et al.*, 2016; Sydow *et al.*, 2016), whereas another model that was simultaneously reported is based on the expression of tau 1N4R (Maeda *et al.*, 2016). Both models display reactive microglia and astrocytes and neuronal hyperexcitability, with deficits in behavioural assays and neuronal loss occurring at later age. The model expressing A152T tau 2N4R was shown to develop ThS and Gallyas silver positive tau tangles starting from 3 months of age (Sydow *et al.*, 2016), but in the model expressing the 1N4R isoform, the misfolded tau was found to remain largely soluble (Maeda *et al.*, 2016). Thus, soluble A152T tau oligomers may be responsible for the overlapping phenotypes of these two models.

Rat-transgenic models have been used less extensively than mouse to study human tauopathies. A rat model expressing full-length tau with the P301L mutation displays tau phosphorylation and accumulation in the sarkosyl-insoluble fraction, but no tangles, nor behavioural abnormalities (Korhonen *et al.*, 2011). Models based on truncated 4R and 3R isoforms (residues 151–391), which are thought to play a role in AD, do display full tau pathology including the presence of phosphorylated, misfolded and sarkosyl-insoluble tau, and ThS and Congo red positive tangles, but without overt neuronal loss (Zilka *et al.*, 2006; Koson *et al.*, 2008; Filipcik *et al.*, 2012).

Models combining A β and tau

Worm models

The link between A β and tau pathologies in AD has remained incompletely understood, in part because animal models have not been particularly successful in recapitulating both features of the disease. Crossing *C. elegans* strains expressing A β 42 and an aggregation-prone tau mutant in neurons was shown to lead to an exacerbation of the toxic phenotype in terms of lifespan,

viability of offspring, chemotaxis and other behavioural assays (Wang *et al.*, 2018). Conversely, crossing an anti-aggregation tau variant and an A β 42 strain has not led to a significantly different phenotype compared to A β 42 alone. The total aggregation load as observed from Congo red staining was found to be much increased in the double-transgenic line with the pro-aggregation tau mutant, and signs of neurodegeneration and neuronal loss were observed, but a mechanistic link between A β 42 and tau toxicity has not been revealed from these studies.

Fly models

Co-expression of A β 42 has been shown to exacerbate the phenotype of wild-type human tau-transgenic flies, manifesting in disrupted axonal transport, abnormal axonal morphology, impaired motor function and shortened lifespan (Folwell *et al.*, 2010). This effect has been suggested to be mediated by increased tau phosphorylation in the presence of A β 42, but the responsible kinases have so far not been identified. In an inducible fly model expressing A β 42 with the Arctic mutation (E22G), the *Drosophila* kinase GSK-3 β is activated and A β 42-mediated toxicity is reduced when endogenous fly tau is removed (Sofola *et al.*, 2010). However, differences in the phosphorylation of *Drosophila* tau in the presence or absence of Arctic A β 42 are below detection limits. Thus, the link between A β 42 and tau phosphorylation remains unclear from the *Drosophila* studies.

Rodent models

APP/PSEN-based rodent models have largely failed to induce significant tau pathology, although tau knock-out models have demonstrated that the presence of tau is required for A β -related toxicity to occur fully (Roberson *et al.*, 2007; Leroy *et al.*, 2012). To induce both A β and tau pathology, however, it has been necessary to develop double- or triple-transgenic models overproducing combinations of human APP, PSEN1/2 and tau, often with mutations that do not occur simultaneously in AD patients. Nevertheless, these models have proved to be useful for understanding the interactions between A β and tau pathology in AD.

The first-transgenic mouse model combining APP with tau has been the TAPP mouse, resulting from a cross of Tg2576, which expresses APP containing the Swedish mutation (K670N/M671L) with the JNPL3 mouse, which expresses the P301L tau variant (Lewis *et al.*, 2001). The number of tau tangles is highly increased in the TAPP mouse compared to the original JNPL3 mouse, and tangles are found in brain regions that are unaffected in the JNPL3 model, including the hippocampus, amygdala and entorhinal cortex. Other regions that are affected by tau pathology in the JNPL3 mouse, such as the hindbrain and spinal cord, do not show an increase. A β deposition is, however, not changed compared to the Tg2576 model, in line with the amyloid cascade hypothesis, according to which A β aggregation occurs upstream of tau pathology (Selkoe and Hardy, 2016). Despite the clear link, tau tangles are not identified in close vicinity of amyloid plaques, indicating that other mechanisms, perhaps mediated by intracellular A β , may play a role. These results have been confirmed by an independent study in which A β 42 fibrils were injected into the pR5 mouse model which also expresses tau P301L, resulting in a 5-fold increase in tau tangles (Götz *et al.*, 2001a, 2001b). In this experiment, the main increase in tau tangles was found in neurons in the amygdala, which are distant but projecting towards the injection site, raising the possibility that retrograde transport of A β 42 can promote tau tangle formation.

A triple-transgenic mouse model, the 3xTg-AD mouse, has been created by co-injecting constructs of the Swedish APP variant (K670N/M671L) and the P301L tau variant into embryos carrying the M146V PSEN1 variant (Oddo *et al.*, 2003). This model has been used extensively in AD research because the temporal and spatial appearance of amyloid plaques and tau tangles resembles that observed in the human disease. Antibody staining for A β is first observed intracellularly at 3–4 months of age, mostly in the neocortex and subsequently in the hippocampus. Plaques develop progressively in the same regions between 6 and 12 months. Tau pathology is initiated later and follows a different spatial pattern, first accumulating in pyramidal neurons in the hippocampus, followed by cortical regions. Aggregated tau can be recognised by conformation-specific and phospho-specific antibodies, as well as Gallyas silver and ThS staining. Although this sequence of events does not prove causality between A β and tau aggregation, both proteins have been found to co-localise within the same neurons, suggesting a role for intracellular A β in triggering tau pathology. Synaptic defects are observed in the hippocampus at 6 months in this model, which correlates with the presence of intracellular A β .

Models based on the PS19 line, expressing the P301S tau variant, have largely confirmed these findings. Crossing this line with the PDAPP mouse or the 5xFAD line has resulted in exacerbated tau pathology, whereas A β deposition is not significantly affected (Hurtado *et al.*, 2010; Saul *et al.*, 2013). The 5xFAD/PS19 model furthermore shows increased astrogliosis compared to the original transgenic lines, as well as increased loss of synapses and neuronal death (Saul *et al.*, 2013). The PDAPP/PS19 mice have been used for detailed analysis of the spatial progression of tau pathology, which matches to some extent the Braak staging of AD (Hurtado *et al.*, 2010). However, a limitation of the PS19 model and crosses thereof is their strong motor phenotype, which interferes with commonly used behavioural assays.

Another triple-transgenic model, TauPS2APP, has resulted from crossing the PS2APP line expressing the K670N/M671L APP variant and the N141I PSEN2, with the pR5 tau P301L strain (Grüninger *et al.*, 2010). In contrast to the models described above, amyloid plaques and accumulations of phosphorylated tau develop in parallel in this model, and in the same regions of the hippocampus. Gallyas silver staining and immuno-EM confirmed that the deposits of tau are of fibrillar nature, and that structures resembling the PHFs observed in AD are present within dendrites. Phosphorylation of S422 has been shown by means of highly specific antibodies to be significantly increased in the triple-transgenic line, compared to the original tau model. However, the triple-transgenic mice do not show signs of increased neuronal loss compared to wild-type controls, and perform similarly to the PS2APP strain in behavioural assays.

PLB1 double mice have been generated by targeted knock-in of the APP K670N/M671L/V717I and tau P301L/R406W mutants (Platt *et al.*, 2011), resulting in more moderate expression levels than in models generated by pronuclear injection, a process that leads to integration of multiple copies of the transgene. This line has been subsequently crossed with a PSEN1 A246E strain (Borchelt *et al.*, 1997) to generate the PLB1 triple mouse model (Platt *et al.*, 2011). Despite the low levels of A β in the triple strain, intracellular staining as well as a small number of extracellular ThS and Congo red positive plaques appear at 6 months of age in the hippocampus and cortex, and the number of plaques increases during ageing. Phosphorylated tau has been identified in the same brain regions from 6 months of age, but no evidence

for mature tau tangles was found. Synaptic plasticity, together with social recognition and spatial memory, start to decline at the same age. Furthermore, glucose metabolism and sleep behaviour are altered in the PLB1 triple mice, as found in patients suffering from AD. Altogether, despite some discrepancies such as the relatively fast accumulation of phosphorylated tau, this less aggressive model recapitulates many features of the early phases of human AD.

Although most studies using double- and triple-transgenic mice have shown a unidirectional relationship between APP/A β expression and increased tau pathology, there is an example in which amyloid deposition is increased by mutant tau expression. In that work, Tg2576 expressing APP with the K670N/M671L Swedish mutation was crossed with a mutant tau line, tau^{VLW}, containing G272V, P301L and R406W mutations (Fig. 3h) (Pérez *et al.*, 2005; Ribé *et al.*, 2005). Not only does the expression of APP^{Swe} lead to increased tau phosphorylation and accumulation in the sarkosyl-insoluble fraction (Pérez *et al.*, 2005), but expression of the tau mutant also increases the numbers of amyloid plaques by up to 5-fold at 16 months of age (Ribé *et al.*, 2005). The double-transgenic animals also suffer from more pronounced neuronal loss than either APP^{Swe} or tau^{VLW} littermates (Ribé *et al.*, 2005).

The models discussed so far have been based on tau mutations found in FTDP-17 patients, or combinations thereof, and they typically display tau pathology in the single-transgenic lines, albeit increased by expression of A β . In a model combining wild-type human tau 0N3R with APP K670N/M671L/V717I and PSEN1 M146L, tau tangles are absent (Boutajangout *et al.*, 2004). However, in addition to APP-stained and ubiquitin-stained dystrophic neurites, tau-positive dystrophic neurites are also found surrounding amyloid plaques in these triple-transgenic mice. In these neurites, tau is recognised by a subset of phospho-specific and conformation-specific antibodies, but is not fibrillar, as indicated from a lack of Gallyas or Congo red staining. Similar observations have been reported for the TAPP and 3xTg-AD mice, although in those models tau tangles do occur in more distant regions from the A β plaques (Lewis *et al.*, 2001; Oddo *et al.*, 2003). Similarly, in a more recent model resulting from a cross of WT human tau 4R with APP^{Swe} and PSEN1 Δ exon9, no evidence was found for the formation of tau tangles, although the mice were only examined up to 10 months of age (Jackson *et al.*, 2016). This model, however, displays increased plaque counts and dystrophic neurites compared to the APP/PSEN1 mice, although no increase in neuronal loss or astrogliosis was observed.

In contrast, another model based on human wild-type tau does feature the development of tangles. This model is the result of a cross between animals expressing APP Δ E693 (the Osaka mutant) and a tau construct comprising 3R and 4R isoforms (Umeda *et al.*, 2014). APP Δ E693 is not associated with amyloid plaque formation, but generates high levels of A β oligomers. These oligomers, detected by the 11A1 conformational antibody, appear earlier in double-transgenic mice than in the original APP line, being evident intracellularly in the regions of the hippocampus and cerebral cortex at 6 months of age. Furthermore, hyperphosphorylated tau, recognised by AT8 and PHF1 antibodies, is formed more rapidly in the CA3 region of the hippocampus and in the cortex in double-transgenic animals, appearing between 6 and 12 months. Tau tangles are present in the same regions as detected by Gallyas silver staining and EM from 18 months of age onwards. Loss of synapses and neuronal death in

the CA3 region are also increased in the double-transgenic line, as are the memory deficits observed in the Morris water maze test. Altogether, whereas other models have failed to produce the full pathology of wild-type tau isoforms, the combination of tau with APP Δ E693 has had better success in doing so. This finding demonstrates that the presence of A β plaques is not a prerequisite for tau pathology and neuronal dysfunction to occur, but A β oligomers may be responsible for triggering this cascade instead.

Taking advantage of the knock-in strategy described in the Section 'A β models' for the APP-based models, a recent study reports on the humanisation of the mouse *MAPT* gene (Saito *et al.*, 2019). Although the subcellular distribution of human tau appears normal in this model, crossing it with the APP knock-in model, APP^{NL-G-F}, results in enhanced tau phosphorylation at several sites. Tau tangles were not identified, yet these more physiological models may prove useful for further studies into the relationship between A β and tau pathology.

Finally, it is worth noting that the spreading of tau pathology by means of neuronal connectivity has received much attention in recent years (Goedert *et al.*, 2017; Jucker and Walker, 2018). In a model expressing the human P301L tau variant exclusively in layer II of the entorhinal cortex, it has been shown that this protein is present in synaptically connected neurons that do not express it, and that it seeds the aggregation of endogenous mouse tau (de Calignon *et al.*, 2012). Crossing this line with the model that expresses APP^{swc} and PSEN1 Δ exon9 has led to an increased spread of misfolded tau, and to increased neuronal loss in the entorhinal cortex where tau is expressed (Pooler *et al.*, 2015). In addition, at 16 months of age, the triple-transgenic animals display more and larger A β deposits in the entorhinal and somatosensory cortices, and more dystrophic neurites surrounding the plaques.

In summary, although it has proved to be difficult to recapitulate the complex pathology and phenotype of human AD in rodent models, there has been significant progress towards this objective as a result of combining in a variety of ways transgenes for APP, PSENs and tau.

α -Synuclein models

Yeast models

The first yeast model for PD, based on the expression of human α -synuclein in *S. cerevisiae*, was reported in 2003 (Fig. 3b) (Outeiro and Lindquist, 2003). Expression of a single copy of GFP-tagged human α -synuclein resulted in the localisation of the protein at lipid membranes, whereas expression of two copies caused it to form cytoplasmic inclusions containing lipid molecules, as shown by staining with the lipophilic dye Nile red, in addition to protein. Furthermore, a dose-dependent cytotoxicity was observed, consistent with a toxic gain of function associated with α -synuclein expression. Two familial α -synuclein mutants A30P and A53T were also investigated in this study. While the A53T mutant was found to display very similar behaviour to wild-type α -synuclein, the A30P mutant was shown to maintain a cytoplasmic distribution at single copy level, rather than accumulating at the lipid membrane, and to exhibit toxicity only when expressed from a high-copy plasmid (Outeiro and Lindquist, 2003). Subsequently, inclusions of A53T α -synuclein, either GFP-tagged or unlabelled, were shown to react with ThS, indicating an amyloid-like structure, whereas staining was not observed in cells expressing the A30P mutant (Zabrocki *et al.*, 2005). These *in vivo* models (Table 2) have confirmed the importance of the

interactions of α -synuclein with lipid membranes, and have provided insights into the correlation between inclusion formation, amyloid-like aggregation and cytotoxicity.

Worm models

The tendency of α -synuclein to accumulate into cytoplasmic inclusions was further corroborated in *C. elegans* models (Table 3). A model overexpressing yellow-fluorescent protein (YFP)-tagged α -synuclein in the body wall muscle cells displays inclusions from day 1 of adulthood, and these increase in number as the worms age (van Ham *et al.*, 2008). Interestingly, fluorescence recovery after photobleaching (FRAP) measurements (Table 1) have indicated that α -synuclein remains mobile within the inclusions, and only in old worms (from day 11 onwards) immobile inclusions, indicative of aggregated proteins, occur (van Ham *et al.*, 2008). These results were confirmed recently by FLIM (Table 1), in which reduction of the fluorescence lifetime of the YFP tag, indicative of fibril formation by α -synuclein, is only observed after day 10 (Laine *et al.*, 2019). However, motility defects are observed from day 4 onwards (van Ham *et al.*, 2008), suggesting that inclusion formation generates toxicity prior to the formation of detectable amyloid-like aggregates.

C. elegans has a particular advantage for PD research in that it contains eight dopaminergic neurons that can easily be tracked. Models overexpressing wild-type and mutant α -synuclein in dopaminergic neurons have been generated in several independent studies (Lakso *et al.*, 2003; Cao *et al.*, 2005; Kuwahara *et al.*, 2006). Although inclusions have only been found in a small fraction of neurons (Lakso *et al.*, 2003; Kuwahara *et al.*, 2006), or not at all (Cao *et al.*, 2005), all studies report a loss of neuronal integrity. Expression of A53T and A30P, but not wild-type, α -synuclein was found to be associated with a loss of dendrites, in agreement with behavioural defects in food sensing, which depends on dopaminergic neurons (Kuwahara *et al.*, 2006). This phenotype has been observed in 3-day old animals (Kuwahara *et al.*, 2006), and in 4-day old animals in another study (Lakso *et al.*, 2003), suggesting that neurotoxicity occurs relatively early in the lifespan of the animals. Furthermore, these studies suggest that α -synuclein neurotoxicity is not correlated with inclusion formation, but may be caused by the presence of smaller aggregates or soluble oligomeric species of α -synuclein.

Fly models

Expression of wild-type, A53T and A30P α -synuclein in the nervous system of *Drosophila* was found to lead to the formation of age-related Lewy body-like inclusions, correlated with the loss of dopaminergic neurons and impaired climbing function (Feany and Bender, 2000). In another model expressing the same α -synuclein variants specifically in the dopaminergic neurons, neuronal loss was also observed which could be restored by overexpression of the molecular chaperone Hsp70 (Auluck *et al.*, 2002). Some controversy remains, however, about the extent of neuronal loss in fly models of PD, and other groups who used different methods have reported that the neurons might have been dysfunctional instead of lost (Pesah *et al.*, 2005; Botella, Bayersdorfer and Schneuwly, 2008). The inclusions in the original model were examined by EM to reveal mostly round aggregates consisting of a rather homogeneous core, surrounded by radiating filaments (Feany and Bender, 2000). The model also displays diffuse aggregates and neuritic threads. Perhaps surprisingly, no differences between the phenotype of the wild-type and mutant α -synuclein lines have been found in this study (Feany and

Bender, 2000). The severity of the phenotype may correlate better with the α -synuclein expression levels, given that a more recent model achieving higher expression levels in the fly brain using the Q binary expression system shows exacerbated locomotory defects and a shortened lifespan, accompanied by α -synuclein inclusions that are mostly ubiquitin-positive, and vacuolar neurodegeneration (Ordonez *et al.*, 2018).

The relationship between α -synuclein aggregation and neurotoxicity has been further addressed by models expressing truncated or modified forms of the protein. A construct lacking part of the non-amyloid component (NAC) domain, which is essential for α -synuclein aggregation (Fig. 2c), does not induce either inclusion formation or degeneration of dopaminergic neurons, whereas a more aggregation-prone C-terminal truncation mutant shows increased deposition of proteins resistant to proteinase K (PK), as well as higher toxicity, supporting a direct correlation between α -synuclein aggregation and toxicity (Periquet *et al.*, 2007). Notably, analysis by western blot has indicated that the levels of soluble oligomers are decreased in the NAC deletion flies, and increased in the C-terminal mutant line, leaving the question open as to whether the PK-resistant species or the soluble oligomers are responsible for toxicity (Periquet *et al.*, 2007).

Other studies have shown that the presence of large inclusions does not correlate with toxicity, and may even be protective. When phosphorylation of S129 is blocked by the mutation of serine to alanine, more inclusions are formed, correlating with more PK-resistant protein, but a reduced loss of dopaminergic neurons (Chen and Feany, 2005). How this observation relates to PD in humans is unclear, as S129-phosphorylated α -synuclein is found in Lewy bodies (Fujiwara *et al.*, 2002). However, a subsequent study has shown that S129 phosphorylation increases oligomer formation, explaining the exacerbated phenotype in the fly model, whereas phosphorylation at Y215 is protective, as blocking this modification results in increased oligomer levels and toxicity (Chen *et al.*, 2009).

The role of α -synuclein oligomers has been further addressed by designing proline mutants that prevent β -strand formation in α -synuclein (Karpinar *et al.*, 2009). These mutations have been found to inhibit fibril formation *in vitro*, yet increase the generation of oligomeric species. When expressed in *C. elegans* vulval muscles, wild-type α -synuclein-YFP forms elongated structures, whereas a triple proline mutant shows a diffuse localisation consistent with the absence of large aggregates (Karpinar *et al.*, 2009). However, the triple proline mutant shows an increased toxicity in both worm and fly models when expressed in dopaminergic neurons, supporting a role for α -synuclein oligomers in neurotoxicity (Karpinar *et al.*, 2009).

Rodent and primate models

As described above in the context of AD models, it has been challenging to develop a rodent model that recapitulates the main characteristic features of PD, including Lewy body pathology, specific loss of dopaminergic neurons in the substantia nigra, and loss of motor function. A complication arises from the fact that mice and rats express their endogenous forms of α -synuclein, which differ from the human form at seven residues, one of the differences being the A53T substitution, which in humans is associated with familial PD. Here, we discuss rodent models based on the transgenic expression of human α -synuclein, focusing on its misfolding and aggregation and the associated phenotypes (Table 5). For a more complete description, see Fernagut and Chesselet (2004); Dehay and Fernagut (2016) and Visanji *et al.*

(2016), and for a discussion on models designed to study the spreading of α -synuclein pathology from cell-to-cell in a prion-like fashion, which is thought to play a role in synucleinopathies, we refer to Dehay *et al.* (2016).

The first-transgenic mouse model expressing human wild-type α -synuclein, which used the PDGF- β promoter, showed accumulation of the protein in several brain regions, including the neocortex, hippocampus and substantia nigra (Masliah *et al.*, 2000). However, these inclusions did not resemble Lewy bodies, appearing in the nucleus as well as the cytoplasm, being associated with the ER, having a less well-defined shape, and consisting of granular rather than fibrillar material. Use of the Thy1 promoter led to a different spatial expression pattern of wild-type α -synuclein (Rockenstein *et al.*, 2002), and extensive characterisation of this model, termed Line 61, has revealed PK-resistant inclusions, phosphorylation of S129, reduced dopamine levels, and progressive PD-like symptoms including olfactory deficits and locomotory impairments, but no substantial neuronal loss (Chesselet *et al.*, 2012). A model expressing A53T α -synuclein under control of the strong PrP promoter showed insoluble, fibrillar protein and inclusions that stain for ThS, in contrast to the wild-type protein (Giasson *et al.*, 2002). At 8 months of age, the A53T mice start to develop motor defects, whereas wild-type α -synuclein-transgenic mice are unaffected. However, the A53T phenotype is not associated with a loss of dopaminergic neurons in the substantia nigra in this model (Giasson *et al.*, 2002).

Conversely, other transgenic models led to degeneration of dopaminergic neurons in the substantia nigra but without the presence of observable inclusions, as exemplified by models expressing a combined A30P + A53T mutant (Richfield *et al.*, 2002; Thiruchelvam *et al.*, 2004), and a C-terminally truncated construct (Wakamatsu *et al.*, 2008), respectively. In a model expressing wild-type human α -synuclein from a bacterial artificial chromosome (BAC) construct that includes the native human regulatory sequences, defects in dopaminergic neurotransmission are accompanied by the presence of high-molecular weight soluble species, although PK-resistant aggregates have not been identified, suggesting that toxicity may arise from intermediate species along the aggregation pathway (Janezic *et al.*, 2013). A mouse model with inducible A53T α -synuclein expression similarly shows high-molecular weight species on western blots in the absence of large inclusions, which correlate with perturbations in the Golgi apparatus in midbrain dopaminergic neurons, as well as changes in lysosomal and autophagic markers (Lin *et al.*, 2012).

Recently, a mouse model was created following the hypothesis that native α -synuclein can occur as folded tetramers, such that a shift to the monomeric conformation puts the protein at risk of aggregation (Bartels *et al.*, 2011; Dettmer *et al.*, 2015). Overexpression of a triple mutant thought to disrupt the tetramers, referred to as 3K because of the three E-to-K substitutions, gives rise to a strong motor phenotype, selective degeneration of dopaminergic neurons in the substantia nigra, and abnormal accumulation of α -synuclein co-localising with vesicles (Nuber *et al.*, 2018). When compared at similar expression levels, the phenotype of the 3K mutant mice is stronger than that of a model expressing a BAC construct with the single E46K mutant that is found in some cases of familial PD. In addition, 3K is present in C-terminally truncated and S129 phosphorylated forms, and in older mice fibrillar material is present in the α -synuclein/vesicle assemblies (Nuber *et al.*, 2018), reminiscent of recent investigations showing that Lewy bodies contain vesicular structures (Shahmoradian *et al.*, 2019; Mahul-Mellier *et al.*, 2020).

Applying the BAC approach to create transgenic rats has led to a model that also recapitulates these and other features of human PD (Nuber *et al.*, 2013). Full-length as well as C-terminally truncated insoluble α -synuclein accumulate as the rats age, following a similar development to that observed in the Braak stages of PD (Braak *et al.*, 2003). PK-resistant inclusions are found in several brain regions including the substantia nigra, and the presence of aggregated protein has been further confirmed by dot blots using the FILA-1 antibody which recognises α -synuclein oligomers and fibrils but not monomers (Nuber *et al.*, 2013). Electron micrographs have shown inclusions containing lysosomes and lipid droplets, as well as accumulated vesicles at synaptic termini. In addition to the accumulation of α -synuclein, the phenotype of this rat model also recapitulates certain aspects of the course of PD in humans, with early changes in avoidance and novelty-seeking behaviour and defects in the sense of smell, followed by later motor symptoms, which correlate with the loss of dopaminergic neurons (Nuber *et al.*, 2013).

The injection of α -synuclein constructs in lentiviral or recombinant adeno-associated viral (rAAV) vectors has been used as an alternative approach to generate high local concentrations of the protein in the substantia nigra and to induce rapid neurodegeneration specifically in this region. The majority of viral vector rat and mouse models displays α -synuclein accumulation in the cytoplasm as well as in dystrophic neurites, yet does not feature Lewy body-like pathology (Kirik *et al.*, 2002; Lo Bianco *et al.*, 2002; Chung *et al.*, 2009; Dehay and Fernagut, 2016). However, PK-resistant α -synuclein and small ThS-positive inclusions have been observed in a rat rAAV model (da Silveira *et al.*, 2009). α -Synuclein accumulates close to the nucleus, the ER and Golgi in this model, and some fibrillar structures were observed in electron micrographs to co-localise with lysosomes and other vesicles. Interestingly, the number of ThS-positive inclusions was significantly increased when using the S129A α -synuclein mutation, which cannot be phosphorylated (da Silveira *et al.*, 2009), an observation analogous to the results obtained in the *Drosophila* model (Chen and Feany, 2005). On the other hand, the phosphomimetic S129D mutation displays fewer, but larger ThS-positive aggregates, whereas no difference in toxicity was found between the different phosphorylation states (da Silveira *et al.*, 2009). However, another study using rat rAAV models reports that the non-phosphorylatable S129A mutant causes more dopaminergic neurodegeneration, whereas S129D forms more inclusions, although the latter have not been examined ultrastructurally (Gorbatyuk *et al.*, 2008). The precise role of S129 phosphorylation in α -synuclein aggregation and toxicity thus remains to be determined (Sato *et al.*, 2013).

Viral vector models have provided a unique means to assess the effect of varying α -synuclein concentrations on its aggregation and the time course of neurodegeneration following injection. The appearance of ThS-positive α -synuclein A53T inclusions is dose-dependent and time-dependent in rats, and correlates with dopaminergic dysfunction (Van der Perren *et al.*, 2015). Western blots have confirmed that insoluble and phosphorylated α -synuclein accumulates over time, and that high-molecular weight oligomeric α -synuclein species are apparent at later time points after injection (Van der Perren *et al.*, 2015). Using the same type of viral vector, a mouse model has also been established that displays Lewy body-like pathology and degeneration of dopaminergic neurons, phosphorylation of S129 and impairments in motor behaviour (Oliveras-Salva *et al.*, 2013). Both wild-type and A53T α -synuclein lead to similar levels of protein accumulation and loss of dopaminergic neurons using this approach. Another

type of optimised rAAV vector has also been employed successfully in mice to express A53T α -synuclein locally leading to a robust phenotype, including the loss of dopaminergic neurons and reduced dopamine levels, accompanied by the formation of PK-resistant α -synuclein inclusions in cell bodies and dystrophic neurites, resembling PD pathology (Ip *et al.*, 2017).

The success with rAAV vectors in rodents has led to the development of non-human primate models using the same technology, resulting in a marmoset model that features α -synuclein accumulation in the cytoplasm and neurites, as well as partial dopaminergic neurodegeneration and motor symptoms (Kirik *et al.*, 2003). In a subsequent study, the monkeys were followed over a longer period of up to a year, leading to the discovery of ubiquitin-positive inclusions, which also occur in human PD (Eslamboli *et al.*, 2007). Furthermore, animals injected with the A53T α -synuclein construct show stronger neurodegeneration compared to the wild-type protein and a worsened phenotype at later time points, potentially making this a useful model to test therapeutic strategies.

Identification of biological pathways using *in vivo* models

Several current disease models are successful in recapitulating key aspects of protein misfolding and aggregation, and exhibit toxic phenotypes, making them useful to interrogate disease mechanisms by identifying the perturbed cellular pathways. Excellent tools are available for commonly used model organisms to probe these questions at the system level by means of genomic, transcriptomic and proteomic studies.

Simple model organisms, including yeast, worms and flies, are amenable to large screens to identify genetic modifiers of protein aggregation and the associated toxicity. In addition, transcriptomics and proteomics can be applied to any organism of which the RNA or protein content can be extracted, to inform on the dysregulation of cellular pathways upon expression of the disease-related gene. Improvements in sequencing technologies have recently increased dramatically the scope of transcriptomics studies, which were previously limited to microarray analysis on a predefined set of genes, whereas genome-wide expression levels can now be determined routinely with high sensitivity and few artefacts using next-generation sequencing methods such as RNAseq (Wang *et al.*, 2009; Breschi *et al.*, 2017). Similarly, developments in mass spectrometry and methods for data analysis have greatly enhanced the possibilities of performing comparative and quantitative proteomics to identify changes in protein abundance as well as post-translational modifications (Bantscheff *et al.*, 2012; Feng *et al.*, 2017; Schubert *et al.*, 2017). Mass spectrometers have not only improved in terms of sensitivity and resolution, but also the opportunities for quantitative analysis of animal and human samples have expanded significantly by chemical labelling approaches such as tandem mass tags (TMTs) and isobaric tags for absolute and relative quantification (iTRAQ). Alternatively, label-free approaches based on counting the number of peptide-to-spectrum matches for each protein have gained popularity. In addition, dedicated proteomic workflows exist that inform on protein–protein interactions and on protein conformations, although these methods are currently largely restricted to unicellular organisms and cultured cells (Minde *et al.*, 2017).

In this section, we highlight the key pathways and genes that have been identified using these various technologies in relation to protein misfolding in the organismal models of AD and PD (Tables 2–5).

Biological pathways in Alzheimer's disease models

Yeast models

Of the *in vivo* models described here, yeast is perhaps the most highly amenable to systematic analysis at the levels of the genome, transcriptome and proteome. Extensive toolkits are available, and profiling of genetic interactions, mRNA expression levels, protein levels, metabolite levels, biological effects of small molecules and phenotypic responses can be performed on a large scale fairly readily (Khurana and Lindquist, 2010). Although yeast does not have a nervous system, or the secretase activity required for APP processing, it has been used to identify conserved-genetic factors involved in A β toxicity (Treusch *et al.*, 2011). The most potent suppressors of such toxicity were found to be genes involved in endocytosis, including the yeast homologue of *PICALM*, which is a known risk factor for AD as identified by genome-wide association studies (GWAS). A number of these hits have been confirmed in a *C. elegans* model expressing A β 42 in glutamatergic neurons (Treusch *et al.*, 2011). Microarray analysis revealed the induction of ER stress and down-regulation of mitochondrial processes upon expression of A β 42 targeted to the secretory pathway (Chen *et al.*, 2017).

Worm models

The main power of *C. elegans* in genetic screens is the ease with which RNA interference (RNAi) can be performed, simply by feeding the worms with bacteria expressing the relevant double-stranded RNA (dsRNA). Libraries targeting 86% of all the annotated *C. elegans* genes are available and can be routinely screened if an appropriate read-out is available (Sin *et al.*, 2014). Performing RNAi by feeding is very efficient for certain tissues, such as intestinal and muscle cells, but works less well in neurons. With respect to A β toxicity, *C. elegans* has been used for an RNAi screen specifically targeted at a conserved set of molecular chaperones (Brehme *et al.*, 2014). Out of 219 molecular chaperone genes tested, knockdown of 18 led to increased paralysis rates in animals expressing A β in the muscle cells. The majority of these genes are also protective in worms expressing polyglutamine, the aggregation of which is linked to Huntington's disease and a variety of other conditions, indicating that their effects may be related to a general improvement in protein homeostasis capacity (Brehme *et al.*, 2014). Another RNAi screen in the *C. elegans* model expressing A β in the muscle cells was targeted at genes with known human homologues, leading to the identification of various hits, including subunits of a vacuolar ATPase and of the ribosome, and a component of the TRiC/CCT chaperone (Khabirova *et al.*, 2014). Mitochondrial function was furthermore shown to be impaired in a model expressing A β 42 in the neurons (Fong *et al.*, 2016), and metabolomics and transcriptomics subsequently demonstrated that early changes in tricarboxylic acid cycle metabolism were underlying this phenotype (Teo *et al.*, 2019).

The link between A β accumulation and the action of molecular chaperones was further supported by mass spectrometry, in a study where two Hsp70 proteins and three small heat shock proteins (sHSPs) were found to co-localise with A β deposits in the body wall muscle cells of *C. elegans* (Fonte *et al.*, 2002). In a study on an inducible *C. elegans* model expressing A β in body wall muscle cells, gene expression changes assessed by RNA microarrays also include sHSPs and Hsp70, as well as mitochondrial unfolded protein response (UPR) genes and proteasomal factors. An aggregation-prone GFP variant was employed as a

control in this study, enabling the identification of the genes that are differentially expressed specifically as a consequence of A β -mediated toxicity, rather than resulting from protein aggregation in general (Hassan *et al.*, 2015). The changes in gene expression resemble those induced by a pore-forming toxin, Cry5B, suggesting that A β may exert its toxic effects in this model by affecting membrane integrity.

In a genome-wide screen in *C. elegans* expressing mutant tau in neurons, molecular chaperones have been identified along with proteases, kinases and phosphatases, and a number of other enzymes, signalling proteins and DNA/RNA regulating factors (Kraemer *et al.*, 2006). The identification of kinases and phosphatases is consistent with the known importance of tau phosphorylation in AD. However, RNAi knockdown of the *C. elegans* homologues of GSK3- β and MARK worsens the phenotype of the tau model, suggesting that the action of these kinases is beneficial, in contrast to their role in tau pathology in humans (Hanger *et al.*, 2009).

Fly models

Drosophila-genetic screens are often based on screening for suppressors or enhancers of the rough eye phenotype caused by the expression of proteins that give rise to toxicity. Fly genetics have provided excellent tools for genome-wide studies, e.g. using randomly inserted transposable elements containing an upstream activation sequence that drives gene expression upon activation by the GAL4 protein, which can be expressed in a tissue-specific manner. Depending on the location and direction of insertion of the transposable element in the context of a gene, expression can either be enhanced or disrupted. Using this system, a screen has been performed on flies expressing A β 42 in the eye, leading to the identification of 23 modifier genes of which several have been confirmed by mutagenesis (Cao *et al.*, 2008). Consistent with the role of these pathways in human disease, genes operating in the secretory pathway and involved in cholesterol metabolism were found, as well as genes regulating chromatin structure and function (Cao *et al.*, 2008). In a complementary approach using a loss-of-function library, the gene *Toll* has been identified, which bears homology to the mammalian interleukin-1 receptor. Deletion of this gene or mutations in downstream-signalling components ameliorate the rough eye phenotype and shorten the lifespan of the A β 42 flies, suggesting an important role of immune signalling in A β 42-mediated toxicity (Tan *et al.*, 2008).

A microarray transcriptomics study on flies expressing A β 42 throughout the nervous system revealed changes in pathways related to oxidative stress and the immune response (Favrin *et al.*, 2013). Although these pathways are also altered during the ageing of control flies, several genes are affected specifically by the expression of A β 42, such as the superoxide dismutase gene *Sod3* which is upregulated ~8-fold compared to controls. Further changes in the A β 42 flies include the upregulation of a gene involved in ER-to-Golgi transport, and the downregulation of a putative ER molecular chaperone, consistent with the involvement of the secretory pathway in A β 42 toxicity (Favrin *et al.*, 2013).

Drosophila tau models have yielded a set of pathways related to toxicity that are different from those in the A β models. Genetic screens using libraries of transposable element insertions led to the identification of phosphatases and kinases as modifiers of human tau toxicity in the fly eye (Shulman and Feany, 2003; Ambegaokar and Jackson, 2011), as well as diverse pathways including autophagy and lysosomal clearance, RNA processing,

chromatin regulation, molecular chaperones and the cytoskeleton (Ambegaokar and Jackson, 2011). An RNAi screen on a tau R406W model also highlighted the role of the cytoskeleton, in particular dynein/dynactin-mediated retrograde axonal transport, in addition to the activity of kinases and phosphatases, and protein metabolism (Butzlaff *et al.*, 2015). Although the role of kinases and phosphatases seems obvious, given the relationship between the phosphorylation state of tau and its propensity to aggregate and cause toxicity, the genetic interactions were shown to be rather complex (Ambegaokar and Jackson, 2011). The phosphorylation state of tau, as probed with the AT8 antibody, does not correlate with either enhancers or suppressors of aggregation. Furthermore, the reduced expression of the GSK3- β homologue is a potent suppressor not only of tau toxicity but also of that induced by A β 42 and TDP-43, suggesting that the network of kinases and phosphatases may play a role in common mechanisms underlying neurodegeneration (Ambegaokar and Jackson, 2011).

The *Drosophila* model based on tau V337M has been used to validate hits from GWAS performed on human AD patients. In one study, the relatively small number of participants did not produce statistically significant results, yet 6 out of 19 *Drosophila* homologues of candidate genes that were tested were shown to be modifiers of tau toxicity in the fly eye, including the glucose transporter *SLC2A14* (Shulman *et al.*, 2011). In a follow-up study of a larger number of GWAS-identified loci, 9 out of 84 genes were confirmed to be modifiers of toxicity including two genes involved in cell adhesion, *CD2AP* and *FERMT2* (Shulman *et al.*, 2014). Thus, this screening strategy appears to have a high success rate compared to that of unbiased screens, and can validate GWAS results below the threshold for statistical significance. It will be interesting to observe whether or not *Drosophila* models based on A β 42 can confirm the roles of additional associated genes. A limitation is, however, presented by human genes which do not have *Drosophila* homologues, such as those involved in lipid metabolism (*APOE* and *CLU*) and immunity (*CRI*, *CD33* and *MS4A*) that are strongly associated with the risk of late-onset AD (Shulman *et al.*, 2014).

Rodent models

Genetic screening for modifiers of protein misfolding is not commonly undertaken in rodents, but analysis of gene expression levels and of protein abundance have been applied extensively to mouse models of AD to shed light on the cellular pathways that are disturbed in response to protein aggregation. Especially in recent years, methods of analysing gene expression using novel sequencing technologies, as well as methods in mass spectrometry to detect protein levels and modifications, have greatly expanded these possibilities.

A meta-analysis of the transcriptomes of aged and diseased humans *versus* mice has revealed that changes during ageing are relatively well-conserved between the two species, for example consistently showing upregulation of inflammatory factors (Hargis and Blalock, 2017). On the other hand, the gene expression profiles of mouse models of neurodegenerative diseases such as AD compared to healthy littermates display limited overlap with those of human patients (Burns *et al.*, 2015; Hargis and Blalock, 2017), showing, for example, less downregulation of genes associated with mitochondrial function, protein degradation, RNA processing and DNA repair (Burns *et al.*, 2015). Although the differences in gene expression between the animal models and human patients represent a major obstacle for the

screening of drug candidates in preclinical trials, the models do allow perturbations in response to the misfolding of transgenically expressed proteins to be deciphered. Complementary insights into the dysregulation of cellular pathways can be obtained from the quantification of protein abundance, which does not always correlate directly with gene expression levels (Liu *et al.*, 2016), especially under conditions where protein quality control and turnover are perturbed. In the case of AD, transcriptomics and proteomics studies have together pointed towards a consistent set of dysregulated pathways in various rodent models, including the immune response, mitochondrial function and synaptic transmission.

The activation of the immune response is generally thought to be a direct reaction to the deposition of A β plaques. In a comparative microarray analysis of brain tissue from Tg2576 and APPPS1 mice, the strongest change in gene expression was found to be a significant upregulation of the inflammatory factor progranulin, which was confirmed to be abundant in microglia and neurons in the vicinity of dense A β plaques (Pereson *et al.*, 2009). BRI-A β 42 mice similarly display progranulin immunoreactivity surrounding A β plaques, suggesting a direct effect of extracellular plaque formation in triggering this immune response (Pereson *et al.*, 2009).

Another study has attempted to separate events resulting directly from A β plaque formation from those caused by other sources of toxicity by comparing the gene expression pattern of two models, the 5xFAD model which has an aggressive phenotype with early onset A β plaque formation, and a model expressing an N-terminal A β truncated form (A β 4-42) that displays neuronal loss in the absence of A β plaques (Bouter *et al.*, 2014). Despite the different pathologies, both models suffer from age-related memory and learning defects of similar magnitudes. In that study, deep sequencing revealed that more than half of the differentially expressed genes in aged mice expressing A β 4-42 with respect to controls are similarly affected in the 5xFAD model, and that these are associated with in a broad range of biological functions including cell differentiation, neurogenesis, histone modification and transcription. Conversely, the majority of genes differentially expressed in the 5xFAD model compared to control mice are related to the immune response. These genes are not altered in the A β 4-42 mice, suggesting that these perturbations are a direct consequence of reactivity to A β plaques in the 5xFAD model. Astrocytes are, however, activated in both models, as indicated by the upregulation of *Gfap*, encoding the typical marker glial fibrillary acidic protein (Bouter *et al.*, 2014).

Microarray analysis of the gene expression profile of the 5xFAD mouse model also showed changes in genes involved in the immune response, pointing towards activation of the complement system and of interferon- γ regulated targets, concomitant with the appearance of pathological features including intracellular A β 42 accumulation, A β plaque deposition and cognitive symptoms (Landel *et al.*, 2014). A mass spectrometric study specifically focusing on the activation of microglia has shown that the levels of the cytokines IL1 β and IL10 are elevated before widespread plaque deposition occurs (Boza-Serrano *et al.*, 2018). A combined proteomics and genomics study on the 5xFAD mouse has also revealed changes in the expression of genes related to immune function, prior to the onset of A β plaque deposition (Bundy *et al.*, 2019). Other perturbations found in this study include the downregulation of proteins involved in the regulation of calcium binding, which show a trend in the same direction for mRNA and protein levels. A total of 40% of the genes, however,

were found to show an anticorrelation with their corresponding protein levels, underscoring the potential differences between gene expression levels and protein abundance, and the importance of complementing transcriptomics with proteomics.

Given the evidence for a role of mitochondrial dysregulation in AD, a detailed proteomic analysis of mitochondria isolated from 3xTg mice has been performed, leading to the identification of 23 proteins that are either up- or down-regulated compared to non-transgenic mice (Chou *et al.*, 2011). These mitochondrial proteins have a variety of functions, including roles in metabolic processes such as oxidative phosphorylation but also functioning in ion transport, apoptosis and mitochondrial protein synthesis. The changes in protein levels are detectable in mice at 6 months of age, prior to widespread A β plaque deposition and tau tangle formation, thus reflecting a perturbation in mitochondrial homeostasis early in the disease process (Chou *et al.*, 2011). A similar approach on the APPPS1 mouse model has revealed changes in the levels of 60 mitochondrial proteins with roles in the electron transport chain, citric acid cycle, oxidative stress and apoptosis, some of which are dysregulated prior to the onset of A β plaque formation (Völgyi *et al.*, 2017).

An integrated proteomics and transcriptomics study on another APP/PSEN1 model took the approach of first investigating changes at the proteome level in different brain regions, followed by validation of the corresponding genetic pathways at the mRNA level (Kempf *et al.*, 2016). Using mass spectrometry of peptides enriched in phosphorylation and glycosylation, changes in protein levels related to cytoskeleton-based synaptic plasticity were found, and the mRNA levels of upstream regulators such as glutamatergic receptors and neurotrophic factors were shown to be affected accordingly. Using quantitative mass spectrometry on APP, APP/PSEN1 and Bri-A β 42 mice, proteins associated with late-onset AD were found to be altered, most notably ApoE, which was observed to be upregulated in all three models (Savas *et al.*, 2017). Furthermore, proteins involved in excitatory synaptic transmission, in particular trafficking of the AMPA receptor, were found to be dysregulated, as well as mitochondrial proteins.

Several other reports have provided further evidence for the perturbation of synaptic plasticity at the proteome level. One study has specifically focused on the proteomic signature associated with memory defects, revealing an overlapping set of dysregulated membrane proteins involved in neuronal excitability and synaptic plasticity in the 5xFAD model of AD, as well as in aged control mice with memory impairment (Neuner *et al.*, 2017). Changes in the cytoskeleton as well as in synaptic function have been reported for the olfactory bulb of an APP/PSEN1 mouse model, occurring around the same age when A β plaques appear (Lachén-Montes *et al.*, 2016). Levels of cytoskeletal proteins as well as chaperones have in addition been suggested to be modulated by the presence of A β oligomers, as discovered in the APP Δ E693 mouse strain that accumulates oligomers rather than plaques (Takano *et al.*, 2013). Finally, the McGill mutant APP rat model displays changes in proteins related to stress response and synaptic function at a young age before the onset of A β plaques, whereas at later age metabolism (e.g. mitochondrial function and amino acid metabolism) becomes affected (Do Carmo *et al.*, 2018).

Analysis of the gene expression patterns of tau-based-transgenic models has revealed important insights into the cellular responses towards tau aggregation. Transcriptional profiling of JNPL3 mice, which express tau P301L, revealed a down-regulation

of anti-apoptotic factors, disturbances of microtubule-dependent intracellular trafficking and down-regulation of a number of immune factors (Ho *et al.*, 2001). A study on another tau P301L model used more stringent selection criteria to identify just one upregulated gene, encoding glyoxalase I (Chen *et al.*, 2004). This enzyme functions in a pathway that prevents glycation and the formation of advanced glycated end products, which are known to occur in ageing and neurodegenerative disease. Glyoxalase I is strongly expressed in neurons with flame-shaped tau aggregates both in the mice and in AD patient brains, suggesting that it may be upregulated as a protective mechanism in response to protein aggregation (Chen *et al.*, 2004).

Changes in proteins functioning in metabolism, including mitochondrial proteins of the respiratory chain complex and detoxifiers of ROS, as well as those with a role in synaptic transmission have furthermore been detected in a proteomic analysis of the tau P301L mouse model (David *et al.*, 2005). The injection of A β 42 fibrils into the amygdala of the tau P301L model has been found to result in further imbalances in components of the cytoskeleton, metabolism and synaptic transmission, as well as proteins functioning in stress response pathways and protein folding and degradation (David *et al.*, 2006). Many of these changes have also been found in SHSY-5Y cells expressing tau P301L upon injection with A β 42 fibrils, suggesting conserved and rapid responses, whereas perturbations in the cytoskeleton and synaptic function appear exclusively in the mouse model, pointing towards longer-term or indirect consequences (David *et al.*, 2006). Proteomic analysis of postsynaptic density isolated from tau P301S mice has shown a depletion of GTPase-regulatory proteins, which could be related to the actin cytoskeletal defects and loss of dendritic spines that are observed in this model (Dejanovic *et al.*, 2018). Consistent with many of these results, RNAseq of a more recent tau P301S mouse model has furthermore revealed perturbations in genes associated with presynaptic factors, glutamatergic signalling and protein scaffolding (Ke *et al.*, 2019).

An additive effect of A β and tau toxicity on mitochondrial dysfunction has been found in a triple mouse model, combining familial APP, PSEN2 and tau mutations (Rhein *et al.*, 2009). Mitochondrial proteins including Complex I and Complex IV subunits are downregulated at the protein level in the triple mouse model more strongly than in the parental APP/PSEN2 or tau strains. In support of these findings, the respiratory function of isolated mitochondria was found to be reduced more drastically and at earlier age in the triple model compared to the APP/PSEN2 strain or wild-type animals (Rhein *et al.*, 2009).

Proteomic approaches have moreover been used to assess the misfolding and aggregation of endogenous mouse proteins, which may be triggered by the pressure on the protein homeostasis machinery caused by the aggregation of transgenic A β and tau. Analysis of the SDS-insoluble fraction from mice expressing APP with Swedish mutations K670N/M671L and PSEN1 Δ exon9 has shown the presence of cytosolic proteins including glycolytic enzymes and members of the 14-3-3 family of chaperones (Xu *et al.*, 2013). In an independent study on the same mouse model using quantitative proteomics, mitochondrial proteins have been identified in the sarkosyl-insoluble fraction of aged mice (Thygesen *et al.*, 2018). Finally, recent work on the inducible tau model rTg4510 has led to a set of SDS-insoluble proteins including kinases, other enzymes and chaperones that partially overlaps with that of the APP/PSEN1 model established using the same methods (Pace *et al.*, 2018), suggesting a common

mechanism that triggers the misfolding of otherwise soluble proteins.

Biological pathways in Parkinson's disease models

Yeast models

The genetic toolkit available for screens in yeast has been applied extensively to α -synuclein models, leading to the discovery of conserved cellular pathways involved in its toxicity. Early screens have made use of a genome-wide deletion library, showing an enrichment for genes involved in lipid metabolism and vesicle-mediated transport (Willingham *et al.*, 2003). In accordance with the latter, a genome-wide overexpression screen for suppressors and enhancers of α -synuclein toxicity identified the yeast homologue of human Rab1, a GTPase involved in ER-to-Golgi vesicle trafficking, as an important suppressor of toxicity. This protein is often found to co-localise with α -synuclein cytoplasmic inclusions, and its overexpression restores the trafficking defects associated with α -synuclein toxicity (Cooper *et al.*, 2006). Yeast homologues of Rab3a and Rab8a, which function in synaptic and post-Golgi vesicle transport, respectively, have also been found associated with the inclusions in subsequent work, and overexpression of any one of these three Rab proteins rescues toxicity in *C. elegans* expressing α -synuclein in dopaminergic neurons (Gitler *et al.*, 2008).

In addition to defects in ER-to-Golgi trafficking, mitochondrial dysfunction has been identified in a transcriptional screen of a yeast strain with high α -synuclein expression levels, together with an increase in ROS (Su *et al.*, 2010). A library screen on this model has revealed a group of 1,2,3,4-tetrahydroquinolinones capable of reducing α -synuclein foci formation and toxicity, in addition to re-establishing ER-to-Golgi trafficking as well as mitochondrial function, suggesting that these processes are tightly linked. These compounds are furthermore active in a *C. elegans* model and in rat primary neuron cultures, providing evidence for a conserved mechanism (Su *et al.*, 2010).

The role of vesicle trafficking has been confirmed once more in a study that made use of the high sensitivity to hydrogen peroxide of the yeast strain expressing α -synuclein (Liang *et al.*, 2008). This study in addition reported ubiquitin-dependent protein degradation, protein biosynthesis and stress response genes as suppressors of toxicity. The *YPP1* gene was discovered as a suppressor specifically for the α -synuclein A30P mutant, and the encoded protein was shown to target α -synuclein to vacuoles for degradation (Flower *et al.*, 2007). Findings of another overexpression screen using a large library of 5500 strains, covering 85% of the yeast genome, are largely in agreement with the pathways previously mentioned, in addition pointing to a role for protein phosphorylation in the toxicity of α -synuclein (Yeager-Lotem *et al.*, 2009). In this screen, the yeast homologue of human *PARK9* was also identified, which encodes a lysosomal P-type ATPase that is mutated in a familial form of PD (Gitler *et al.*, 2009). Overexpression of this gene suppresses α -synuclein toxicity not only in yeast, but also in a *C. elegans* model and in rat primary dopaminergic neurons. It also protects yeast cells against manganese-induced toxicity, a known risk factor for PD, providing a potential link between genetic and environmental factors in initiating the disease process (Gitler *et al.*, 2009). Two other hits from this screen, an E3 ubiquitin ligase and a factor related to nitrosative stress, have later been validated in cortical neurons differentiated from patient-derived induced pluripotent stem cells (iPSCs) (Chung *et al.*, 2013a).

Building on the results from genetic screens, a computational approach was designed with the aim of creating a genetic network underlying the disease phenotype, and of transposing it to the human homologues. Thus, a genetic map of α -synuclein toxicity has been created which spans the pathways identified previously, ranging from ER quality control and vesicle trafficking to calcium signalling and mitochondrial function, in which known genetic causes of familial PD appear in different parts of the network (Khurana *et al.*, 2017). In an accompanying study that employs *in situ* peroxidase labelling coupled to mass spectrometry, the physical interaction partners of α -synuclein in rat primary cortical neurons were found to form a proteomic network that largely overlaps with the genetic network (Chung *et al.*, 2017). Altogether, the yeast α -synuclein models in combination with available genomic and proteomic tools have generated remarkable results in understanding the processes underlying PD.

Worm models

In agreement with the findings in yeast, several independent genetic screens in *C. elegans* found components of vesicle trafficking pathways to be involved in α -synuclein accumulation and toxicity. A genome-wide RNAi screen for suppressors of inclusion formation in the *C. elegans* model expressing α -synuclein in the body wall muscle cells revealed an enrichment of genes associated with the ER/Golgi and vesicle transport (van Ham *et al.*, 2008). In a study targeting a subset of genes known or hypothesised to be involved in PD, two vesicle trafficking genes were discovered that protect against α -synuclein-mediated degeneration of dopaminergic neurons, in addition to known genetic PD factors such as *DJ-1* and *PINK1* (Hamamichi *et al.*, 2008). Finally, another screen performed on a pan-neuronal α -synuclein model led to the detection of several genes involved in endocytosis that protected against α -synuclein neurotoxicity, including two subunits of the adapter protein complex AP2 (Kuwahara *et al.*, 2008).

A gene that seemingly operates in a separate pathway that promotes aggregation was originally discovered in a screen targeting polyglutamine aggregation in the body wall muscle cells of *C. elegans*. This gene, *moag-4* or *SERF* in humans, is similarly active in driving α -synuclein inclusion formation and toxicity, independently of known chaperone-dependent or insulin-signalling pathways (van Ham *et al.*, 2010). Although the function of *SERF* in humans is not known at present, this gene may represent a suitable target for therapeutic intervention in protein misfolding diseases.

Fly models

Drosophila models of α -synuclein toxicity have been used less extensively in the context of unbiased-genetic screens. Using the reduced levels of dopamine in the heads of flies expressing α -synuclein A53T as a read-out, in combination with a genome-wide deletion library, the mitochondrial chaperone TRAP1 has been identified as a potent suppressor of toxicity (Butler *et al.*, 2012). Overexpressing TRAP1 in the α -synuclein A53T flies partially restores their climbing ability and resistance to oxidative stress, and the latter has been confirmed in rat neurons as well as in human cell lines, pointing towards a conserved link between α -synuclein pathology and mitochondrial dysfunction that is thought to play an important role in PD (Vicario *et al.*, 2018).

Rodent models

Several transcriptomics and proteomics studies have been performed in recent years in rodent models for PD based on the transgenic expression of α -synuclein. One investigation was

carried out on a cross of an A53T α -synuclein overexpression model and a knock-out strain of the mitochondrial kinase PINK1, which causes an enhanced phenotype with ubiquitinated α -synuclein inclusions in the spinal cord and basal brain in mice from 1 year of age, and a reduced lifespan (Gispert *et al.*, 2015). Transcriptomic analysis of brain samples taken before the appearance of α -synuclein inclusions in this model has revealed perturbations in genes related to mitochondrial function, ubiquitin-mediated degradation, microtubule and vesicle trafficking, synaptic signalling and DNA-damage repair, compared to wild-type animals. At the protein level, analysis of ubiquitination patterns in 18-month-old mice has provided evidence for enhanced ubiquitination of α -synuclein itself upon the loss of PINK1, as well as reduced ubiquitination of the histone variant H2AX, a marker of sites of DNA damage (Gispert *et al.*, 2015).

A quantitative proteomics study on the model expressing α -synuclein with both A30P and A53T mutations has also highlighted the role of mitochondrial dysfunction and oxidative stress in the substantia nigra, as well as impairment of the ubiquitin-proteasome system, and ER stress (Yan *et al.*, 2017). A useful data source was furthermore established by a detailed analysis of the protein levels in distinct regions of the mouse brain, which has been performed for both non-transgenic mice and the line 61 PD mouse model (Jung *et al.*, 2017b). Comparison of the substantia nigra and an unaffected brain region in the PD model showed that membrane signalling proteins are upregulated specifically in the substantia nigra, whereas markers of extracellular matrix remodelling and inflammation are increased in aged PD mice compared to non-transgenic mice.

Further studies at the system level have mostly been performed on rodent models of PD induced by treatment with neurotoxins such as 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine, which have been remarkably successful in recapitulating phenotypic aspects of human PD (Tieu, 2011; Kasahara *et al.*, 2017). Although these studies have identified perturbations in common pathways including oxidative phosphorylation, oxidative stress, apoptosis and synaptic transmission (Smith, 2009), a comparative meta-analysis revealed that the gene expression signature of these models is relatively remote from that of human PD patients (Oerton and Bender, 2017). Overall, the current animal models of PD provide useful tools to study certain aspects of the multifactorial disease process, such as the interplay between mitochondrial dysfunction and α -synuclein aggregation, but need to be improved to recapitulate better the complexity of the disease in the human brain.

Methodological challenges

An essential aim of *in vivo* models of human disease is to enable preclinical studies on systems that recapitulate the pathological processes observed in patients. In this section, we discuss four major challenges, two biophysical and two biological, that remain to be addressed in order to reach this goal:

- (1) *To develop longitudinal detection methods for protein aggregates.* In AD and PD drug discovery pipelines, preclinical studies in mouse models often rely on the post-mortem quantification of protein deposits using methods based on immunohistochemistry or amyloid-sensitive fluorescent dyes (Sevigny *et al.*, 2016; Drummond and Wisniewski, 2017). Methods based on clearing techniques, including CLARITY (Chung and Deisseroth, 2013; Chung *et al.*, 2013b),

3DISCO (Ertürk *et al.*, 2012) and Scale (Hama *et al.*, 2011) (Table 1), enable more quantitative assessments of the total amounts and distributions of these deposits, thereby offering more accurate endpoints on the anti-aggregation activities of therapeutic candidate compounds.

One may expect even better readouts, however, by the further development of methods to monitor longitudinally the inhibitory effects of these compounds. Approaches based on magnetic resonance imaging (MRI) and PET (Table 1) (Weiner *et al.*, 2013; Sevigny *et al.*, 2016) already offer the possibility of carrying out this type of studies in mouse models. The development of PET tracers for α -synuclein will be particularly helpful in this direction (Merchant *et al.*, 2019).

To develop detection methods for protein oligomers. Both in AD and PD the most neurotoxic species are misfolded protein oligomers, and yet these species remain elusive and difficult to quantify, in particular in *in vivo* models (Table 1). Immunohistochemistry is typically used to probe the accumulation of A β , tau and α -synuclein in fixed samples, and conformation-specific antibodies are available for specific cases, including A11, which binds to β -sheet oligomers, and MC-1, which recognises misfolded tau. The recent introduction of enzyme-linked immunosorbent assay (ELISA)-based methods of greatly enhanced sensitivity, such as SIMOA (Kuhle *et al.*, 2016), subject to further development, is opening novel opportunities in this direction. In our laboratory we have recently demonstrated the design of antibodies targeting specific epitopes (Sormanni *et al.*, 2015; Aprile *et al.*, 2017; Aprile *et al.*, in press), and this approach holds promise for discriminating different populations of oligomeric species based on the accessibility of a range of epitopes.

Advances in microscopy are furthermore expected to contribute to an improved molecular understanding of misfolded protein species *in vivo*. Super-resolution microscopy has revealed the morphology and the growth of fibrils at the nanometre scale *in vitro* and in cells (Kaminski Schierle *et al.*, 2011b; Duim *et al.*, 2014; Pinotsi *et al.*, 2016), and continuing efforts to implement these technologies *in vivo* have already allowed the use of RESOLFT and STED nanoscopy to visualise organelles and the cytoskeleton in *C. elegans* (Dreier *et al.*, 2019), *Drosophila* (Schnorrenberg *et al.*, 2016) and the mouse brain (Berning *et al.*, 2012). Furthermore, the use of biosensors in combination with fluorescence microscopy may shed more light on the conformational state of protein aggregates *in vivo*, as demonstrated e.g. by the changes in the fluorescence lifetime of protein tags upon amyloid formation (Kaminski Schierle *et al.*, 2011a; Laine *et al.*, 2019).

Another approach that holds promise for the detection of oligomeric protein species in animal models and patient material relies on seeded amplification *in vitro*. Several assays, e.g. protein misfolding cyclic amplification (PMCA) (Saborio *et al.*, 2001), the amyloid-seeding assay (Colby *et al.*, 2007) and real-time quaking-induced conversion (RT-QuIC) (Atarashi *et al.*, 2011) (Table 1), were originally designed to detect low concentrations of misfolded PrP molecules, based on the conversion of PrP^C to PrP^{Sc} by the addition of seeds of the latter species. These assays have been adapted successfully to enable the detection of misfolded α -synuclein (Shahnawaz *et al.*, 2017; Jung *et al.*, 2017a) and tau (Meyer *et al.*, 2014) *in vitro*, from mice and from

human brain or cerebrospinal fluid. Further development of these approaches will undoubtedly help not only with the fundamental understanding of protein misfolding *in vivo*, but may also lead to diagnostic methods for patients.

- (1) *To create models to recapitulate the sporadic forms of AD and PD.* The overexpression of A β , tau and α -synuclein or their mutational variants provides models that recapitulate the familial forms of AD and PD. In the sporadic forms, more subtle perturbations of the protein homeostasis system are probably at play, which result in the aggregation of the disease-associated proteins starting from normal expression levels. The creation of specific animal models is becoming technically more feasible with developments in genetic-engineering techniques such as CRISPR/Cas9 (Knott and Doudna, 2018; Zhang, 2019), and by introducing additional genetic or environmental perturbations the need for the artificially high overexpression levels and non-physiological mutations on which current models often rely may be alleviated. For example, we have recently created a *C. elegans* model expressing A β 42 at single-copy levels in only two neurons, which displays small perturbations in neuronal signalling and may be useful to study the onset of disease symptoms triggered by additional sources of cellular stress (Sinnige et al., 2019).
- (2) *To create models to recapitulate the complexity of human AD and PD brain tissues.* The *in vivo* models that we have described in this review have the inevitable limitation that they are genetically non-identical to humans, leading to differences in neuronal anatomy as well as processes such as ageing and the action of the immune system, which are critical in the study of neurodegenerative diseases. The use of iPSC-derived neurons and direct reprogramming of patient fibroblasts are promising approaches to circumventing some of these issues (Xu et al., 2015; Engle et al., 2018). In combination with 2D and 3D culturing methods, such as neuronal rosettes and brain organoids, respectively (Choi et al., 2014; Otani et al., 2016; Lancaster et al., 2017; Gonzalez et al., 2018), these systems may increasingly provide avenues to elucidate disease mechanisms and to screen for therapeutics, in parallel to the use of traditional model organisms.

Conclusions

The use of *in vivo* models is instrumental in obtaining a detailed understanding of human diseases by connecting the observed phenotypes to the underlying molecular processes that give rise to them. In this review, we have described current *in vivo* models of AD and PD where the phenomenon of protein misfolding and aggregation plays a central role.

Studies based on these models have shown that mature amyloid fibrils or large aggregate inclusions are relatively inert and generally not associated with cytotoxicity, as they are limited in their ability to engage in aberrant interactions. By contrast, aggregation intermediates such as oligomeric species formed by A β , tau or α -synuclein are associated with the disturbance of cellular processes and with the onset and progress of neurodegenerative diseases. Some of the affected pathways appear to be disease-specific, as exemplified by the association of vesicle trafficking and mitochondrial function with the misfolding of α -synuclein, and of cytoskeletal transport and protein phosphorylation with that of tau. Other pathways may be perturbed more generally, most

notably the protein homeostasis network which becomes progressively impaired in ageing and neurodegeneration. Molecular chaperones have been shown to interact with all three proteins discussed in this paper, and to modulate their misfolding and aggregation (Sinnige et al., 2020).

However, the pathological mechanisms underlying these disorders are not yet fully understood, and no disease-modifying therapeutics have yet emerged. The creation of *in vivo* models that better recapitulate the sporadic forms of these diseases, as well as the development of biophysical methods for the accurate quantification of the aggregation process in living organisms, will be crucial in this regard.

References

- Ahmed M, Davis J, Aucoin D, Sato T, Ahuja S, Aimoto S, Elliott JJ, Van Nostrand WE and Smith SO (2010) Structural conversion of neurotoxic amyloid-beta(1-42) oligomers to fibrils. *Nature Structural & Molecular Biology* 17, 561–567.
- Alberti S and Dormann D (2019) Liquid–liquid phase separation in disease. *Annual Review of Genetics* 53, 171–194.
- Alexander AG, Marfil V and Li C (2014) Use of *Caenorhabditis elegans* as a model to study Alzheimer's disease and other neurodegenerative diseases. *Frontiers in Genetics* 5, 279.
- Ambadipudi S, Biernat J, Riedel D, Mandelkow E and Zweckstetter M (2017) Liquid–liquid phase separation of the microtubule-binding repeats of the Alzheimer-related protein tau. *Nature Communications* 8, 1–13.
- Ambadipudi S, Reddy JG, Biernat J, Mandelkow E and Zweckstetter M (2019) Residue-specific identification of phase separation hot spots of Alzheimer's-related protein tau. *Chemical Science* 10, 6503–6507.
- Ambegaokar SS and Jackson GR (2011) Functional genomic screen and network analysis reveal novel modifiers of tauopathy dissociated from tau phosphorylation. *Human Molecular Genetics* 20, 4947–4977.
- Amen T and Kaganovich D (2016) Yeast screening platform identifies FDA-approved drugs that reduce A β oligomerization. *Microbial Cell (Graz, Austria)* 3, 97–100.
- Andorfer C, Kress Y, Espinoza M, De Silva R, Tucker KL, Barde YA, Duff K and Davies P (2003) Hyperphosphorylation and aggregation of tau in mice expressing normal human tau isoforms. *Journal of Neurochemistry* 86, 582–590.
- Andorfer C, Acker CM, Kress Y, Hof PR, Duff K and Davies P (2005) Cell-cycle reentry and cell death in transgenic mice expressing nonmutant human tau isoforms. *Journal of Neuroscience* 25, 5446–5454.
- Aprile FA, Sormanni P, Perni M, Arosio P, Linse S, Knowles TPJ, Dobson CM and Vendruscolo M (2017) Selective targeting of primary and secondary nucleation pathways in A β 42 aggregation using a rational antibody scanning method. *Science Advances* 3, e1700488.
- Atarashi R, Satoh K, Sano K, Fuse T, Yamaguchi N, Ishibashi D, Matsubara T, Nakagaki T, Yamanaka H, Shirabe S, Yamada M, Mizusawa H, Kitamoto T, Klug G, McGlade A, Collins SJ and Nishida N (2011) Ultrasensitive human prion detection in cerebrospinal fluid by real-time quaking-induced conversion. *Nature Medicine* 17, 175–178.
- Auluck PK, Chan HYE, Trojanowski JQ, Lee VMY and Bonini NM (2002) Chaperone suppression of alpha-synuclein toxicity in a *Drosophila* model for Parkinson's disease. *Science (New York, N.Y.)* 295, 865–868.
- Baglioni S, Casamenti F, Bucciantini M, Luheshi LM, Taddei N, Chiti F, Dobson CM and Stefani M (2006) Prefibrillar amyloid aggregates could be generic toxins in higher organisms. *The Journal of Neuroscience* 26, 8160–8167.
- Bagriantsev S and Liebman S (2006) Modulation of A β 42 low-n oligomerization using a novel yeast reporter system. *BMC Biology* 4, 1–12.
- Balch WE, Morimoto RI, Dillin A and Kelly JW (2008) Adapting proteostasis for disease intervention. *Science (New York, N.Y.)* 319, 916–919.
- Bantscheff M, Lemeer S, Savitski MM and Kuster B (2012) Quantitative mass spectrometry in proteomics: critical review update from 2007 to the present. *Analytical and Bioanalytical Chemistry* 404, 939–965.

- Barabási A-L, Gulbahce N and Loscalzo J (2011) Network medicine: a network-based approach to human disease. *Nature Reviews Genetics* **12**, 56–68.
- Bardai FH, Wang L, Mutreja Y, Yenjerla M, Gamblin TC and Feany MB (2018) A conserved cytoskeletal signaling cascade mediates neurotoxicity of FTDP-17 tau mutations *in vivo*. *The Journal of Neuroscience* **38**, 108–119.
- Bartels T, Choi JG and Selkoe DJ (2011) α -Synuclein occurs physiologically as a helically folded tetramer that resists aggregation. *Nature* **477**, 107–110.
- Bateman RJ, Xiong C, Benzinger TLS, Fagan AM, Goate A, Fox NC, Marcus DS, Cairns NJ, Xie X, Blazey TM, Holtzman DM, Santacruz A, Buckles V, Oliver A, Moulder K, Aisen PS, Ghetti B, Klunck WE, McDade E, Martins RN, Masters CL, Mayeux R, Ringman JM, Rossor NM, Schofield PR, Sperling RA, Salloway S and Morris JC (2012) Clinical and biomarker changes in dominantly inherited Alzheimer's disease. *New England Journal of Medicine* **367**, 795–804.
- Bemporad F and Chiti F (2012) Protein misfolded oligomers: experimental approaches, mechanism of formation, and structure–toxicity relationships. *Chemistry and Biology* **19**, 315–327.
- Benilova I, Karran E and De Strooper B (2012) The toxic A β oligomer and Alzheimer's disease: an emperor in need of clothes. *Nature Neuroscience* **15**, 349–357.
- Berning S, Willig KI, Steffens H, Dibaj P and Hell SW (2012) Nanoscopy in a living mouse brain. *Science (New York, N.Y.)* **335**, 551 LP–551551.
- Bharadwaj PR, Verdile G, Barr RK, Gupta V, Steele JW, Lenard Lachenmayer M, Yue Z, Ehrlich ME, Petsko G, Ju S, Ringe D, Sankovich SE, Caine JM, MacRae IG, Gandy S and Martins RN (2012) Latrepirdine (Dimebon™) enhances autophagy and reduces intracellular GFP-A β 42 levels in yeast. *Journal of Alzheimer's Disease* **32**, 949–967.
- Bodner CR, Maltsev AS, Dobson CM and Bax A (2010) Differential phospholipid binding of α -synuclein variants implicated in Parkinson's disease revealed by solution NMR spectroscopy. *Biochemistry* **49**, 862–871.
- Borchelt DR, Thinakaran G, Eckman CB, Lee MK, Davenport F, Ratovitsky T, Prada CM, Kim G, Seekins S, Yager D, Slunt HH, Wang R, Seeger M, Levey AI, Gandy SE, Copeland NG, Jenkins NA, Price DL, Younkin SG and Sisodia SS (1996) Familial Alzheimer's disease-linked presenilin 1 variants elevate A β 1–42/1–40 ratio *in vitro* and *in vivo*. *Neuron* **17**, 1005–1013.
- Borchelt DR, Ratovitski T, Van Lare J, Lee MK, Gonzales V, Jenkins NA, Copeland NG, Price DL and Sisodia SS (1997) Accelerated amyloid deposition in the brains of transgenic mice coexpressing mutant presenilin 1 and amyloid precursor proteins. *Neuron* **19**, 939–945.
- Borst JW, Visser NV, Kouptsova O and Visser AJWG (2000) Oxidation of unsaturated phospholipids in membrane bilayer mixtures is accompanied by membrane fluidity changes. *Biochimica et Biophysica Acta (BBA) – Molecular and Cell Biology of Lipids* **1487**, 61–73.
- Botella JA, Bayersdorfer F and Schneuwly S (2008) Superoxide dismutase overexpression protects dopaminergic neurons in a *Drosophila* model of Parkinson's disease. *Neurobiology of Disease* **30**, 65–73.
- Boutajangout A, Authelat M, Blanchard V, Touchet N, Tremp G, Pradier L and Brion JP (2004) Characterisation of cytoskeletal abnormalities in mice transgenic for wild-type human tau and familial Alzheimer's disease mutants of APP and presenilin-1. *Neurobiology of Disease* **15**, 47–60.
- Bouter Y, Kacprowski T, Weissmann R, Dietrich K, Borgers H, Brauß A, Sperling C, Wirths O, Albrecht M, Jensen LR, Kuss AW and Bayer TA (2014) Deciphering the molecular profile of plaques, memory decline and neuron loss in two mouse models for Alzheimer's disease by deep sequencing. *Frontiers in Aging Neuroscience* **6**, 75.
- Boza-Serrano A, Yang Y, Paulus A and Deierborg T (2018) Innate immune alterations are elicited in microglial cells before plaque deposition in the Alzheimer's disease mouse model 5xFAD. *Scientific Reports* **8**, 1–16.
- Braak H and Braak E (1991) Neuropathological staging of Alzheimer-related changes. *Acta Neuropathologica* **82**, 239–259.
- Braak H, Del Tredici K, Rüb U, de Vos RAI, Jansen Steur ENH and Braak E (2003) Staging of brain pathology related to sporadic Parkinson's disease. *Neurobiology of Aging* **24**, 197–211.
- Brandt R, Gergou A, Wacker I, Fath T and Hutter H (2009) A *Caenorhabditis elegans* model of tau hyperphosphorylation: induction of developmental defects by transgenic overexpression of Alzheimer's disease-like modified tau. *Neurobiology of Aging* **30**, 22–33.
- Brehme M, Voisine C, Rolland T, Wachi S, Soper JH, Zhu Y, Orton K, Vilella A, Garza D, Vidal M, Ge H and Morimoto RI (2014) A chaperome subnetwork safeguards proteostasis in aging and neurodegenerative disease. *Cell Reports* **9**, 1–16.
- Breschi A, Gingeras TR and Guigó R (2017) Comparative transcriptomics in human and mouse. *Nature Reviews Genetics* **18**, 425.
- Brorsson AC, Bolognesi B, Tartaglia GG, Shammass SL, Favrin G, Watson I, Lomas DA, Chiti F, Vendruscolo M, Dobson CM, Crowther DC and Luheshi LM (2010) Intrinsic determinants of neurotoxic aggregate formation by the amyloid β peptide. *Biophysical Journal* **98**, 1677–1684.
- Bucciantini M, Giannoni E, Chiti F, Baroni F, Formigli L, Zurdo J, Taddei N, Ramponi G, Dobson CM and Stefani M (2002) Inherent toxicity of aggregates implies a common mechanism for protein misfolding diseases. *Nature* **416**, 507–511.
- Buell AK, Galvagnion C, Gaspar R, Sparr E, Vendruscolo M, Knowles TPJ, Linse S and Dobson CM (2014) Solution conditions determine the relative importance of nucleation and growth processes in α -synuclein aggregation. *Proceedings of the National Academy of Sciences of the United States of America* **111**, 7671–7676.
- Bundy JL, Vied C, Badger C and Nowakowski RS (2019) Sex-biased hippocampal pathology in the 5XFAD mouse model of Alzheimer's disease: a multi-omic analysis. *Journal of Comparative Neurology* **527**, 462–475.
- Burns TC, Li MD, Mehta S, Awad AJ and Morgan AA (2015) Mouse models rarely mimic the transcriptome of human neurodegenerative diseases: a systematic bioinformatics-based critique of preclinical models. *European Journal of Pharmacology* **759**, 101–117.
- Butler EK, Voigt A, Lutz AK, Toegel JP, Gerhardt E, Karsten P, Falkenburger B, Reinartz A, Winklhofer KF and Schulz JB (2012) The mitochondrial chaperone protein TRAP1 mitigates α -synuclein toxicity. *PLoS Genetics* **8**, e1002488.
- Butzlaff M, Hannan SB, Karsten P, Lenz S, Ng J, Voßfeldt H, Prüßing K, Pflanz R, Schulz JB, Rasse T and Voigt A (2015) Impaired retrograde transport by the Dynein/Dynactin complex contributes to Tau-induced toxicity. *Human Molecular Genetics* **24**, 3623–3637.
- Caine J, Sankovich S, Antony H, Waddington L, Macreadie P, Varghese J and Macreadie I (2007) Alzheimer's A β fused to green fluorescent protein induces growth stress and a heat shock response. *FEMS Yeast Research* **7**, 1230–1236.
- Cao S, Gelwix CC, Caldwell KA and Caldwell GA (2005) Torsin-mediated protection from cellular stress in the dopaminergic neurons of *Caenorhabditis elegans*. *The Journal of Neuroscience* **25**, 3801–3812.
- Cao W, Song HJ, Gangi T, Kelkar A, Antani I, Garza D and Konsolaki M (2008) Identification of novel genes that modify phenotypes induced by Alzheimer's β -amyloid overexpression in *Drosophila*. *Genetics* **178**, 1457–1471.
- Carmine-Simmen K, Proctor T, Tschäpe J, Poeck B, Triphan T, Strauss R and Kretschmar D (2009) Neurotoxic effects induced by the *Drosophila* amyloid-beta peptide suggest a conserved toxic function. *Neurobiology of Disease* **33**, 274–281.
- Casas C, Sergeant N, Itier JM, Blanchard V, Wirths O, van der Kolk N, Vingdeux V, van de Steeg E, Ret G, Canton T, Drobecq H, Clark A, Bonici B, Delacourte A, Benavides J, Schmitz C, Tremp G, Bayer TA, Benoit P and Pradier L (2004) Massive CA1/2 neuronal loss with intra-neuronal and N-terminal truncated A β 42 accumulation in a novel Alzheimer transgenic model. *The American Journal of Pathology* **165**, 1289–1300.
- Cerf E, Sarroukh R, Tamamizu-Kato S, Breydo L, Derclaye S, Dufrene YF, Narayanaswami V, Goormaghtigh E, Ruyschaert J-M and Raussens V (2009) Antiparallel β -sheet: a signature structure of the oligomeric amyloid β -peptide. *Biochemical Journal* **421**, 415–423.
- Chakraborty R, Vepuri V, Mhatre SD, Paddock BE, Miller S, Michelson SJ, Delvadia R, Desai A, Vinokur M, Melicharek DJ, Ureja S, Khandelwal P, Ansaloni S, Goldstein LE, Moir RD, Lee JC, Tabb LP, Saunders AJ and Marendra DR (2011) Characterization of a *Drosophila* Alzheimer's disease model: pharmacological rescue of cognitive defects. *PLoS ONE* **6**, e20799.
- Chen L and Feany MB (2005) α -Synuclein phosphorylation controls neurotoxicity and inclusion formation in a *Drosophila* model of Parkinson disease. *Nature Neuroscience* **8**, 657–663.

- Chen X and Petranovic D (2015) Amyloid- β peptide-induced cytotoxicity and mitochondrial dysfunction in yeast. *FEMS Yeast Research* 15, 1–10.
- Chen G, Chen KS, Knox JH, Inglis J, Bernard A, Martin SJ, Justice A, McConlogue L, Games D, Freedman SB and Morris RGM (2000) A learning deficit related to age and β -amyloid plaques in a mouse model of Alzheimer's disease. *Nature* 408, 975–979.
- Chen F, Wollmer MA, Hoerndli F, Mu G, Tsolaki M and Papassotiropoulos A (2004) Role for glyoxalase I in Alzheimer's disease. *Proceedings of the National Academy of Sciences of the United States of America* 101, 7687–7692.
- Chen L, Periquet M, Wang X, Negro A, McLean PJ, Hyman BT and Feany MB (2009) Tyrosine and serine phosphorylation of alpha-synuclein have opposing effects on neurotoxicity and soluble oligomer formation. *Journal of Clinical Investigation* 119, 3257–3265.
- Chen SW, Drakulic S, Deas E, Ouberai M, Aprile FA, Arranz R, Ness S, Roodveldt C, Williams T, De-Genst EJ, Klenerman D, Wood NW, Knowles TPJ, Alfonso C, Rivas G, Abramov AY, Valpuesta JM, Dobson CM and Cremades N (2015) Structural characterization of toxic oligomers that are kinetically trapped during α -synuclein fibril formation. *Proceedings of the National Academy of Sciences* 112, E1994–E2003.
- Chen X, Bisschops MMM, Agarwal NR, Ji B, Shanmugavel KP and Petranovic D (2017) Interplay of energetics and ER stress exacerbates Alzheimer's amyloid- β (A β) toxicity in yeast. *Frontiers in Molecular Neuroscience* 10, 232.
- Cheslet MF, Richter F, Zhu C, Magen I, Watson MB and Subramaniam SR (2012) A progressive mouse model of Parkinson's disease: the Thy1- α Syn ('line 61') mice. *Neurotherapeutics: The Journal of the American Society for Experimental Neurotherapeutics* 9, 297–314.
- Chiti F and Dobson CM (2017) Protein misfolding, amyloid formation, and human disease: a summary of progress over the last decade. *Annual Review of Biochemistry* 86, 27–68.
- Choi SH, Kim YH, Hebisch M, Sliwinski C, Lee S, D'Avanzo C, Chen H, Hooli B, Asselin C, Muffat J, Klee JB, Zhang C, Wainger BJ, Peitz M, Kovacs DM, Woolf CJ, Wagner SL, Tanzi RE and Kim DY (2014) A three-dimensional human neural cell culture model of Alzheimer's disease. *Nature* 515, 274–278.
- Chou JL, Shenoy DV, Thomas N, Choudhary PK, LaFerla FM, Goodman SR and Breen GAM (2011) Early dysregulation of the mitochondrial proteome in a mouse model of Alzheimer's disease. *Journal of Proteomics* 74, 466–479.
- Chung K and Deisseroth K (2013) CLARITY for mapping the nervous system. *Nature Methods* 10, 508–513.
- Chung CY, Koprach JB, Siddiqi H and Isacson O (2009) Dynamic changes in presynaptic and axonal transport proteins combined with striatal neuroinflammation precede dopaminergic neuronal loss in a rat model of AAV-synucleinopathy. *The Journal of Neuroscience* 29, 3365–3373.
- Chung CY, Khurana V, Auluck PK, Tardiff DF, Mazzuli JR, Soldner F, Baru V, Lou Y, Freyzon Y, Cho S, Mungenast AE, Muffat J, Mitalipova M, Pluth MD, Jui NT, Schüle B, Lippard SJ, Tsai LH, Krainc D, Buchwald SL, Jaenisch R and Lindquist S (2013a) Identification and rescue of α -synuclein toxicity in Parkinson patient-derived neurons. *Science (New York, N.Y.)* 342, 983–987.
- Chung K, Wallace J, Kim S-Y, Kalyanasundaram S, Andalman AS, Davidson TJ, Mirzabekov JJ, Zalocusky KA, Mattis J, Denisin AK, Pak S, Bernstein H, Ramakrishnan C, Grosenick L, Gradinaru V and Deisseroth K (2013b) Structural and molecular interrogation of intact biological systems. *Nature* 497, 332–337.
- Chung CY, Khurana V, Yi S, Sahni N, Loh KH, Auluck PK, Baru V, Udeshi ND, Freyzon Y, Carr SA, Hill DE, Vidal M, Ting AY and Lindquist S (2017) *In situ* peroxidase labeling and mass-spectrometry connects alpha-synuclein directly to endocytic trafficking and mRNA metabolism in neurons. *Cell Systems* 4, 242–250, e4.
- Citron M, Westaway D, Xia W, Carlson G, Diehl T, Levasque G, Johnson-Wood K, Lee M, Seubert P, Davis A, Kholodenko D, Motter R, Sherrington R, Perry B, Yao H, Strome R, Lieberburg I, Rommens J, Kim S, Schenk D, Fraser P, St George Hyslop P and Selkoe DJ (1997) Mutant presenilins of Alzheimer's disease increase production of 42-residue amyloid β -protein in both transfected cells and transgenic mice. *Nature Medicine* 3, 67–72.
- Cohen SIA, Vendruscolo M, Dobson CM and Knowles TPJ (2012) From macroscopic measurements to microscopic mechanisms of protein aggregation. *Journal of Molecular Biology* 421, 160–171.
- Cohen RM, Rezai-Zadeh K, Weitz TM, Rentsendorj A, Gate D, Spivak I, Bholat Y, Vasilevko V, Glabe CG, Breunig JJ, Rakic P, Davtyan H, Agadjanyan MG, Kepe V, Barrio JR, Bannykh S, Szekely CA, Pechnick RN and Town T (2013a) A transgenic Alzheimer rat with plaques, tau pathology, behavioral impairment, oligomeric A β , and frank neuronal loss. *Journal of Neuroscience* 33, 6245–6256.
- Cohen SIA, Linse S, Luheshi LM, Hellstrand E, White DA, Rajah L, Otzen DE, Vendruscolo M, Dobson CM and Knowles TPJ (2013b) Proliferation of amyloid- β 42 aggregates occurs through a secondary nucleation mechanism. *Proceedings of the National Academy of Sciences of the United States of America* 110, 9758–9763.
- Colby DW, Zhang Q, Wang S, Groth D, Legname G, Riesner D and Prusiner S (2007) Prion detection by an amyloid seeding assay. *Proceedings of the National Academy of Sciences of the United States of America* 104, 20914–20919.
- Colodner KJ and Feany MB (2010) Glial fibrillary tangles and JAK/STAT-mediated glial and neuronal cell death in a *Drosophila* model of glial tauopathy. *The Journal of Neuroscience* 30, 16102–16113.
- Colvin MT, Silvers R, Ni QZ, Can TV, Sergeev I, Rosay M, Donovan KJ, Michael B, Wall J, Linse S and Griffin RG (2016) Atomic resolution structure of monomeric A β 42 amyloid fibrils. *Journal of the American Chemical Society* 138, 9663–9674.
- Conway KA, Lee S-J, Rochet J-C, Ding TT, Williamson RE and Lansbury PT (2000) Acceleration of oligomerization, not fibrillization, is a shared property of both α -synuclein mutations linked to early-onset Parkinson's disease: implications for pathogenesis and therapy. *Proceedings of the National Academy of Sciences* 97, 571–576.
- Cooper AA, Gitler Cashikar AD, Cole A, Haynes M, Bhullar Liu B, Xu K, Strathearn K, Liu KE, Cao F, Caldwell S, Caldwell KA, Marsischky GA, Kolodner G, Labaer RD, Rochet J, Bonini J and Lindquist NM, S (2006) α -Synuclein blocks ER-Golgi traffic and Rab1 rescues neuron loss in Parkinson's models. *Science (New York, N.Y.)* 313, 324–329.
- Cremades N, Cohen SIA, Deas E, Abramov AY, Chen AY, Orte A, Sandal M, Clarke RW, Dunne P, Aprile FA, Bertocini CW, Wood NW, Knowles TPJ, Dobson CM and Klenerman D (2012) Direct observation of the interconversion of normal and toxic forms of α -synuclein. *Cell* 149, 1048–1059.
- Crowther RA, Jakes R, Spillantini MG and MG Y (1998) Synthetic filaments assembled from C-terminally truncated K-synuclein. *FEBS Letters* 436, 309–312.
- Crowther DC, Kinghorn KJ, Miranda E, Page R, Curry JA, Duthie FAI, Gubb DC and Lomas DA (2005) Intraneuronal A β , non-amyloid aggregates and neurodegeneration in a *Drosophila* model of Alzheimer's disease. *Neuroscience* 132, 123–135.
- Cukalevski R, Yang X, Meisl G, Weininger U, Bernfur K, Frohm B, Knowles TPJ and Linse S (2015) The A β 40 and A β 42 peptides self-assemble into separate homomolecular fibrils in binary mixtures but cross-react during primary nucleation. *Chemical Science* 6, 4215–4233.
- D'Angelo F, Vignaud H, Di Martino J, Salin B, Devin A, Cullin C and Marchal C (2013) A yeast model for amyloid-aggregation exemplifies the role of membrane trafficking and PICALM in cytotoxicity. *Disease Models & Mechanisms* 6, 206–216.
- da Silveira SA, Schneider BL, Cifuentes-Diaz C, Sage D, Abbas-Terki T, Iwatsubo T, Unser M and Aebischer P (2009) Phosphorylation does not prompt, nor prevent, the formation of α -synuclein toxic species in a rat model of Parkinson's disease. *Human Molecular Genetics* 18, 872–887.
- David DC, Hauptmann S, Scherping I, Schuessel K, Keil U, Rizzu P, Ravid R, Dröse S, Brandt U, Müller WE, Eckert A and Götz J (2005) Proteomic and functional analyses reveal a mitochondrial dysfunction in P301L tau transgenic mice. *Journal of Biological Chemistry* 280, 23802–23814.
- David DC, Ittner LM, Gehrig P, Nergensau D, Shepherd C, Halliday G and Götz J (2006) β -Amyloid treatment of two complementary P301L tau-expressing Alzheimer's disease models reveals similar deregulated cellular processes. *Proteomics* 6, 6566–6577.
- Dawson TM, Golde TE and Lagier-Tourenne C (2018) Animal models of neurodegenerative diseases. *Nature Neuroscience* 21, 1370–1379.

- De S, Wirthensohn DC, Flagmeier P, Hughes C, Aprile FA, Ruggeri FS, Whiten DR, Emin D, Xia Z, Varela JA, Sormanni P, Kundel F, Knowles TPJ, Dobson CM, Bryant C, Vendruscolo M and Klenerman D (2019) Different soluble aggregates of A β 42 can give rise to cellular toxicity through different mechanisms. *Nature Communications* **10**, 1541.
- de Calignon A, Polydoro M, Suárez-Calvet M, William C, Adamowicz DH, Kopeikina KJ, Pitstick R, Sahara N, Ashe KH, Carlson GA, Spire-Jones TL and Hyman BT (2012) Propagation of tau pathology in a model of early Alzheimer's disease. *Neuron* **73**, 685–697.
- Decker JM, Krüger L, Sydow A, Dennissen FJ, Siskova Z, Eckhard M and Eva-Maria M (2016) The Tau/A152T mutation, a risk factor for frontotemporal-spectrum disorders, leads to NR2B receptor-mediated excitotoxicity. *EMBO Reports* **17**, 552–569.
- Dehay B and Fernagut P-O (2016) Alpha-synuclein-based models of Parkinson's disease. *Revue Neurologique* **172**, 371–378.
- Dehay B, Vila M, Bezard E, Brundin P and Kordower JH (2016) Alpha-synuclein propagation: new insights from animal models. *Movement Disorders* **31**, 161–168.
- Dejanovic B, Huntley MA, De Mazière A, Meilandt WJ, Wu T, Srinivasan K, Jiang Z, Gandham V, Friedman BA, Ngu H, Foreman O, Carano RAD, Chih B, Klumperman J, Bakalarski C, Hanson JE and Sheng M (2018) Changes in the synaptic proteome in tauopathy and rescue of tau-induced synapse loss by C1q antibodies. *Neuron* **100**, 1322–1336, e7.
- del Alonso AC, Zaidi T, Novak M, Grundke-Iqbal I and Iqbal K (2001) Hyperphosphorylation induces self-assembly of τ Into tangles of paired helical filaments/straight filaments. *Proceedings of the National Academy of Sciences of the United States of America* **98**, 6923–6928.
- Dettmer U, Newman AJ, Soldner F, Luth ES, Kim NC, Von Saucken VE, Sanderson JB, Jaenisch R, Bartels T and Selkoe D (2015) Parkinson-causing α -synuclein missense mutations shift native tetramers to monomers as a mechanism for disease initiation. *Nature Communications* **6**, 7314.
- De Vos A, Anandhakumar J, Van den Brande J, Verduyck M, Franssens V, Winderickx J and Swinnen E (2011) Yeast as a model system to study tau biology. *International Journal of Alzheimer's Disease* **2011**, 428970.
- Di Scala C, Chahinian H, Yahi N, Garmy N and Fantini J (2014) Interaction of Alzheimer's β -amyloid peptides with cholesterol: mechanistic insights into amyloid pore formation. *Biochemistry* **53**, 4489–4502.
- Dixon C, Mathias N, Zweig RM, Davis DA and Gross DS (2005) Alpha-synuclein targets the plasma membrane via the secretory pathway and induces toxicity in yeast. *Genetics* **170**, 47–59.
- Do Carmo S and Cuello AC (2013) Modeling Alzheimer's disease in transgenic rats. *Molecular Neurodegeneration* **8**, 1–11.
- Do Carmo S, Crynen G, Paradis T, Reed J, Iulita MF, Ducatenzeiler A, Crawford F and Cuello AC (2018) Hippocampal proteomic analysis reveals distinct pathway deregulation profiles at early and late stages in a rat model of Alzheimer's-like amyloid pathology. *Molecular Neurobiology* **55**, 3451–3476.
- Dodart JC, Mathis C, Saura J, Bales KR, Paul SM and Ungerer A (2000) Neuroanatomical abnormalities in behaviorally characterized APP(V717F) transgenic mice. *Neurobiology of Disease* **7**, 71–85.
- Dosanjh LE, Brown MK, Rao G, Link CD and Luo Y (2010) Behavioral phenotyping of a transgenic *Caenorhabditis elegans* expressing neuronal amyloid-beta. *Journal of Alzheimer's Disease* **19**, 681–690.
- Dreier J, Castello M, Coccano G, Cáceres R, Plastino J, Vicidomini G and Testa I (2019) Smart scanning for low-illumination and fast RESOLFT nanoscopy *in vivo*. *Nature Communications* **10**, 556.
- Drummond E and Wisniewski T (2017) Alzheimer's disease: experimental models and reality. *Acta Neuropathologica* **133**, 155–175.
- Duff K, Eckman C, Zehr C, Yu X, Prada CM, Perez-Tur J, Hutton M, Buee L, Harigaya Y, Yager D, Morgan D, Gordon MN, Holcomb L, Refolo L, Zenk B, Hardy J and Younkin S (1996) Increased amyloid- β 42(43) in brains of mice expressing mutant presenilin 1. *Nature* **383**, 710–713.
- Duff K, Knight H, Refolo LM, Sanders S, Yu X, Picciano M, Malester B, Hutton M, Adamson J, Goedert M, Burki K and Davies P (2000) Characterization of pathology in transgenic mice over-expressing human genomic and cDNA tau transgenes. *Neurobiology of Disease* **7**, 87–98.
- Duim WC, Jiang Y, Shen K, Frydman J and Moerner WE (2014) Super-resolution fluorescence of Huntingtin reveals growth of globular species into short fibers and coexistence of distinct aggregates. *ACS Chemical Biology* **9**, 2767–2778.
- Eckermann K, Mocanu MM, Khlistunova I, Biernat J, Nissen A, Hofmann A, Schönig K, Bujard H, Haemisch A, Mandelkow E, Zhou L, Rune G and Mandelkow EM (2007) The β -propensity of tau determines aggregation and synaptic loss in inducible mouse models of tauopathy. *Journal of Biological Chemistry* **282**, 31755–31765.
- Eisenberg D and Jucker M (2012) The amyloid state of proteins in human diseases. *Cell* **148**, 1188–1203.
- Elder GA, Gama Sosa MA and De Gasperi R (2010) Transgenic mouse models of Alzheimer's disease. *Mount Sinai Journal of Medicine* **77**, 69–81.
- Engle SJ, Blaha L and Kleiman RJ (2018) Best practices for translational disease modeling using human iPSC-derived neurons. *Neuron* **100**, 783–797.
- Ertürk A, Becker K, Jähring N, Mauch CP, Hojer CD, Egen JG, Hellal F, Bradke F, Sheng M and Dodt H-U (2012) Three-dimensional imaging of solvent-cleared organs using 3DISCO. *Nature Protocols* **7**, 1983–1995.
- Eslamboli A, Romero-Ramos M, Burger C, Bjorklund T, Muzyczka N, Mandel RJ, Baker H, Ridley RM and Kirik D (2007) Long-term consequences of human alpha-synuclein overexpression in the primate ventral midbrain. *Brain* **130**, 799–815.
- Fatouros C, Pir GJ, Biernat J, Koushika SP, Mandelkow E, Mandelkow EM, Schmidt E and Baumeister R (2012) Inhibition of tau aggregation in a novel *Caenorhabditis elegans* model of tauopathy mitigates proteotoxicity. *Human Molecular Genetics* **21**, 3587–3603.
- Favrin G, Bean DM, Bilsland E, Boyer H, Fischer BE, Russell S, Crowther DC, Baylis HA, Oliver SG and Giannakou ME (2013) Identification of novel modifiers of A β toxicity by transcriptomic analysis in the fruit fly. *Scientific Reports* **3**, 1–10.
- Fay DS, Fluet A, Johnson CJ and Link CD (1998) *In vivo* aggregation of beta-amyloid peptide variants. *Journal of Neurochemistry* **71**, 1616–1625.
- Feany MB and Bender WW (2000) A *Drosophila* model of Parkinson's disease. *Nature* **404**, 394–398.
- Feng Y, Cappelletti V and Picotti P (2017) Quantitative proteomics of model organisms. *Current Opinion in Systems Biology* **6**, 58–66.
- Fernagut P-O and Chesselet M-F (2004) Alpha-synuclein and transgenic mouse models. *Neurobiology of Disease* **17**, 123–130.
- Filipcik P, Zilka N, Bugos O, Kucerek J, Koson P, Novak P and Novak M (2012) First transgenic rat model developing progressive cortical neurofibrillary tangles. *Neurobiology of Aging* **33**, 1448–1456.
- Finelli A, Kelkar A, Song HJ, Yang H and Konsolaki M (2004) A model for studying Alzheimer's Abeta42-induced toxicity in *Drosophila melanogaster*. *Molecular and Cellular Neuroscience* **26**, 365–375.
- Flagmeier P, Meisl G, Vendruscolo M, Knowles TPJ, Dobson CM, Buell AK and Galvagnion C (2016) Mutations associated with familial Parkinson's disease alter the initiation and amplification steps of α -synuclein aggregation. *Proceedings of the National Academy of Sciences of the United States of America* **113**, 201604645.
- Flower TR, Chesnokova LS, Froelich CA, Dixon C and Witt SN (2005) Heat shock prevents alpha-synuclein-induced apoptosis in a yeast model of Parkinson's disease. *Journal of Molecular Biology* **351**, 1081–1100.
- Flower TR, Clark-Dixon C, Metoyer C, Yang H, Shi R, Zhang Z and Witt SN (2007) YGR198w (YPP1) targets A30P α -synuclein to the vacuole for degradation. *Journal of Cell Biology* **177**, 1091–1104.
- Folwell J, Cowan CM, Ubhi KK, Shiab H, Newman TA, Shepherd D and Mudher A (2010) A β exacerbates the neuronal dysfunction caused by human tau expression in a *Drosophila* model of Alzheimer's disease. *Experimental Neurology* **223**, 401–409.
- Fong S, Teo E, Ng LF, Chen CB, Lakshmanan LN, Tsoi SY, Moore PK, Inoue T, Halliwell B and Gruber J (2016) Energy crisis precedes global metabolic failure in a novel *Caenorhabditis elegans* Alzheimer disease model. *Scientific Reports* **6**, 33781.
- Fonte V, Kapulkin WJ, Taft A, Fluet A, Friedman D and Link CD (2002) Interaction of intracellular beta amyloid peptide with chaperone proteins. *Proceedings of the National Academy of Sciences of the United States of America* **99**, 9439–9444.

- Fossgreen A, Brückner B, Czech C, Masters CL, Beyreuther K and Paro R (1998) Transgenic *Drosophila* expressing human amyloid precursor protein show gamma-secretase activity and a blistered-wing phenotype. *Proceedings of the National Academy of Sciences of the United States of America* **95**, 13703–13708.
- Freer R, Sormanni P, Vecchi G, Ciryam P, Dobson CM and Vendruscolo M (2016) A protein homeostasis signature in healthy brains recapitulates tissue vulnerability to Alzheimer's disease. *Science Advances* **2**, e1600947.
- Fu H, Possenti A, Freer R, Nakano Y, Hernandez Villegas NC, Tang M, Cahy PVM, Lassus BA, Chen S, Fowler SL, Figueroa HY, Huey ED, Johnson GVW, Vendruscolo M and Duff KE (2019) A tau homeostasis signature is linked with the cellular and regional vulnerability of excitatory neurons to tau pathology. *Nature Neuroscience* **22**, 47–56.
- Fujiwara H, Hasegawa M, Dohmae N, Kawashima A, Masliah E, Goldberg MS, Shen J, Takio K and Iwatsubo T (2002) Alpha-synuclein is phosphorylated in synucleinopathy lesions. *Nature Cell Biology* **4**, 160–164.
- Fusco G, Chen SW, Williamson PTF, Cascella R, Perni M, Jarvis JA, Cecchi C, Vendruscolo M, Chiti F, Cremades N and Ying L (2017) Structural basis of membrane disruption and cellular toxicity by alpha-synuclein oligomers. *Science (New York, N.Y.)* **358**, 1440–1443.
- Galvagnion C, Buell AK, Meisl G, Michaels TCT, Vendruscolo M, Knowles TPJ and Dobson CM (2015) Lipid vesicles trigger α -synuclein aggregation by stimulating primary nucleation. *Nature Chemical Biology* **11**, 229–234.
- Galvagnion C, Brown JW, Ouberai MM, Flagmeier P, Vendruscolo M, Buell AK, Sparr E and Dobson CM (2016) Chemical properties of lipids strongly affect the kinetics of the membrane-induced aggregation of α -synuclein. *Proceedings of the National Academy of Sciences* **113**, 7065–7070.
- Games D, Adams D, Alessandrini R, Barbour R, Berthelette P, Blackwell C, Carr T, Clemens J, Donaldson T and Gillespie F (1995) Alzheimer-type neuropathology in transgenic mice overexpressing V717F beta-amyloid precursor protein. *Nature* **373**, 523–527.
- Giasson BI, Duda JE, Quinn SM, Zhang B, Trojanowski JQ and Lee VM-Y (2002) Neuronal alpha-synucleinopathy with severe movement disorder in mice expressing A53T human alpha-synuclein. *Neuron* **34**, 521–533.
- Gispert S, Brehm N, Weil J, Seidel K, Rüb U, Kern B, Walter M, Roeper J and Auburger G (2015) Potentiation of neurotoxicity in double-mutant mice with Pink1 ablation and A53T-SNCA overexpression. *Human Molecular Genetics* **24**, 1061–1076.
- Gitler AD, Bevis BJ, Shorter J, Strathearn KE, Hamamichi S, Su LJ, Caldwell KA, Caldwell GA, Rochet J-C, McCaffery JM, Barlowe C and Lindquist S (2008) The Parkinson's disease protein alpha-synuclein disrupts cellular Rab homeostasis. *Proceedings of the National Academy of Sciences of the United States of America* **105**, 145–150.
- Gitler AD, Chesi A, Geddie ML, Strathearn KE, Hamamichi S, Hill KJ, Caldwell KA, Caldwell GA, Cooper AA, Rochet JC and Lindquist S (2009) α -Synuclein is part of a diverse and highly conserved interaction network that includes PARK9 and manganese toxicity. *Nature Genetics* **41**, 308–315.
- Goedert M, Spillantini MG, Jakes R, Rutherford D and Crowther RA (1989) Multiple isoforms of human microtubule-associated protein tau: sequences and localization in neurofibrillary tangles of Alzheimer's disease. *Neuron* **3**, 519–526.
- Goedert M, Eisenberg DS and Crowther RA (2017) Propagation of tau aggregates and neurodegeneration. *Annual Review of Neuroscience* **40**, 189–210.
- Goffeau A, Barrell G, Bussey H, Davis RW, Dujon B, Feldmann H, Galibert F, Hoheisel JD, Jacq C, Johnston M, Louis EJ, Mewes HW, Murakami Y, Philippsen P, Tettelin H and Oliver SG (1996) Life with 6000 genes. *Science (New York, N.Y.)* **274**, 546–567.
- Gonzalez C, Armijo E, Bravo-Alegria J, Becerra-Calixto A, Mays CE and Soto C (2018) Modeling amyloid beta and tau pathology in human cerebral organoids. *Molecular Psychiatry* **23**, 2363–2374.
- Gorbatyuk OS, Li S, Sullivan LF, Chen W, Kondrikova G, Manfredsson FP, Mandel RJ and Muzyczka N (2008) The phosphorylation state of Ser-129 in human alpha-synuclein determines neurodegeneration in a rat model of Parkinson disease. *Proceedings of the National Academy of Sciences of the United States of America* **105**, 763–768.
- Götz J, Chen F, Barmettler R and Nitsch RM (2001a) Tau filament formation in transgenic mice expressing P301L tau. *Journal of Biological Chemistry* **276**, 529–534.
- Götz J, Chen F, van Dorpe J and Nitsch RM (2001b) Formation of neurofibrillary tangles in P301L tau transgenic mice induced by abeta 42 fibrils. *Science (New York, N.Y.)* **293**, 1491–1495.
- Götz J, Bodea L-G and Goedert M (2018) Rodent models for Alzheimer disease. *Nature Reviews Neuroscience* **19**, 583–598.
- Greeve I, Kretschmar D, Tschape J, Beyn A, Brellinger C, Schweizer M, Nitsch R and Reifegerste R (2004) Age-dependent neurodegeneration and Alzheimer-amyloid plaque formation in transgenic *Drosophila*. *Journal of Neuroscience* **24**, 3899–3906.
- Gremer L, Schölzel D, Schenk C, Reinart E, Labahn J, Ravelli RBG, Tusche M, Lopez-Iglesias C, Hoyer W, Heise H, Willbold D and Schröder GF (2017) Fibril structure of amyloid- β (1–42) by cryo-electron microscopy. *Science (New York, N.Y.)* **358**, 116–119.
- Grueninger F, Bohrmann B, Czech C, Ballard TM, Frey JR, Weidensteiner C, von Kienlin M and Ozmen L (2010) Phosphorylation of tau at S422 is enhanced by A β in TauPS2APP triple transgenic mice. *Neurobiology of Disease* **37**, 294–306.
- Haass C and Selkoe DJ (2007) Soluble protein oligomers in neurodegeneration: lessons from the Alzheimer's amyloid β -peptide. *Nature Reviews Molecular Cell Biology* **8**, 101–112.
- Habchi J, Chia S, Limbocker R, Mannini B, Ahn M, Perni M, Hansson O, Arosio P, Kumita JR, Challa PK, Cohen SIA, Linse S, Dobson CM, Knowles TPJ and Vendruscolo M (2016) Systematic development of small molecules to inhibit specific microscopic steps of A β 42 aggregation in Alzheimer's disease. *Proceedings of the National Academy of Sciences of the United States of America* **114**, E200–E208.
- Habchi J, Chia S, Galvagnion C, Michaels TCT, Bellaiche MMJ, Ruggeri FS, Sanguanini M, Idini I, Kumita JR, Sparr E, Linse S, Dobson CM, Knowles TPJ and Vendruscolo M (2018) Cholesterol catalyses A β 42 aggregation through a heterogeneous nucleation pathway in the presence of lipid membranes. *Nature Chemistry* **10**, 673–683.
- Hall AM and Roberson ED (2012) Mouse models of Alzheimer's disease. *Brain Research Bulletin* **88**, 3–12.
- Hama H, Kurokawa H, Kawano H, Ando R, Shimogori T, Noda H, Fukami K, Sakaue-Sawano A and Miyawaki A (2011) Scale: a chemical approach for fluorescence imaging and reconstruction of transparent mouse brain. *Nature Neuroscience* **14**, 1481–1488.
- Hamamichi S, Rivas RN, Knight AL, Cao S, Caldwell KA and Caldwell GA (2008) Hypothesis-based RNAi screening identifies neuroprotective genes in a Parkinson's disease model. *Proceedings of the National Academy of Sciences of the United States of America* **105**, 728–733.
- Hanger DP, Anderton BH and Noble W (2009) Tau phosphorylation: the therapeutic challenge for neurodegenerative disease. *Trends in Molecular Medicine* **15**, 112–119.
- Hardy J and Selkoe DJ (2002) The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science (New York, N.Y.)* **297**, 353–356.
- Hargis KE and Blalock EM (2017) Transcriptional signatures of brain aging and Alzheimer's disease: what are our rodent models telling us? *Behavioural Brain Research* **322**, 311–328.
- Harigaya Y, Saido TC, Eckman CB, Prada C-M, Shoji M and Younkin SG (2000) Amyloid beta protein starting pyroglutamate at position 3 is a major component of the amyloid deposits in the Alzheimer's disease brain. *Biochemical and Biophysical Research Communications* **276**, 422–427.
- Hassan WM, Dostal V, Huemann BN, Yerg JE and Link CD (2015) Identifying A β -specific pathogenic mechanisms using a nematode model of Alzheimer's disease. *Neurobiology of Aging* **36**, 857–866.
- Heggland I, Storkaas IS, Soligard HT, Kbro-Flatmoen A and Witter MP (2015) Stereological estimation of neuron number and plaque load in the hippocampal region of a transgenic rat model of Alzheimer's disease. *European Journal of Neuroscience* **41**, 1245–1262.
- Hellstrand E, Sparr E and Linse S (2010) Retardation of A β fibril formation by phospholipid vesicles depends on membrane phase behavior. *Biophysical Journal* **98**, 2206–2214.

- Herdon LA, Schmeissner PJ, Dudaronek JM, Brown PA, Listner KM, Sakano Y, Paupard MC, Hall DH and Driscoll M** (2002) Stochastic and genetic factors influence tissue-specific decline in ageing *C. elegans*. *Nature* **419**, 808–814.
- Herrup K** (2015) The case for rejecting the amyloid cascade hypothesis. *Nature Neuroscience* **18**, 794–799.
- Ho L, Xiang Z, Mukherjee P, Zhang W, De Jesus N, Mirjany M, Yemul S and Pasinetti GM** (2001) Gene expression profiling of the tau mutant (P301L) transgenic mouse brain. *Neuroscience Letters* **310**, 16–19.
- Holcomb L, et al.** (1998) Accelerated Alzheimer-type phenotype in transgenic mice carrying both mutant amyloid precursor protein and presenilin 1 transgenes. *Nature Medicine* **4**, 97–100.
- Hong M, Zhukareva V, Vogelsberg-Ragaglia V, Wszolek Z, Reed L, Miller BI, Geschwind DH, Bird TD, McKeel D, Goate A, Morris JC, Wilhelmsen KC, Schellenberg GD, Trojanowski JQ and Lee VM-Y** (1998) Mutation-specific functional impairments in distinct tau isoforms of hereditary FTDP-17. *Science (New York, N.Y.)* **282**, 1914 LP–1911917.
- Hsiao K, Chapman P, Nilsen S, Eckman C, Harigaya Y, Younkin S, Yang FS and Cole G** (1996) Correlative memory deficits, Ab elevation, and amyloid plaques in transgenic mice. *Science (New York, N.Y.)* **274**, 99–102.
- Huang JK, Ma PL, Ji SY, Zhao XL, Tan JX, Sun XJ and De Huang F** (2013) Age-dependent alterations in the presynaptic active zone in a *Drosophila* model of Alzheimer's disease. *Neurobiology of Disease* **51**, 161–167.
- Huang D, Zimmerman MI, Martin PK, Nix AJ, Rosenberry TL and Paravastu AK** (2015) Antiparallel β -sheet structure within the C-terminal region of 42-residue Alzheimer's amyloid- β peptides when they form 150-kDa oligomers. *Journal of Molecular Biology* **427**, 2319–2328.
- Hurtado DE, Molina-Porcel L, Iba M, Aboagye AK, Paul SM, Trojanowski JQ and Lee VMY** (2010) A β accelerates the spatiotemporal progression of tau pathology and augments tau amyloidosis in an Alzheimer mouse model. *American Journal of Pathology* **177**, 1977–1988.
- Hutton M, Lewis J, Dickson D, Yen S-H and McGowan E** (2001) Analysis of tauopathies with transgenic mice. *Trends in Molecular Medicine* **7**, 467–470.
- Iijima K and Iijima-ando K** (2008) *Drosophila* models of Alzheimer's amyloidosis: the challenge of dissecting the complex mechanisms of toxicity of amyloid-beta 42. *Journal of Alzheimer's Disease* **15**, 523–540.
- Iijima K, Liu H-P, Chiang A-S, Hearn SA, Konsolaki M and Zhong Y** (2004) Dissecting the pathological effects of human Abeta40 and Abeta42 in *Drosophila*: a potential model for Alzheimer's disease. *Proceedings of the National Academy of Sciences of the United States of America* **101**, 6623–6628.
- Iijima K, Chiang HC, Hearn SA, Hakker I, Gatt A, Shenton C, Granger L, Leung A, Iijima-Ando K and Zhong Y** (2008) A β 42 mutants with different aggregation profiles induce distinct pathologies in *Drosophila*. *PLoS ONE* **3**, e1703.
- Ip CW, Klaus LC, Karikari AA, Visanji NP, Brotchie JM, Lang AE, Volkman J and Koprich JB** (2017) AAV1/2-induced overexpression of A53T- α -synuclein in the substantia nigra results in degeneration of the nigrostriatal system with Lewy-like pathology and motor impairment: a new mouse model for Parkinson's disease. *Acta Neuropathologica Communications* **5**, 11.
- Irizarry MC, Soriano F, McNamara M, Page KJ, Schenk D, Games D and Hyman BT** (1997) Abeta deposition is associated with neuropil changes, but not with overt neuronal loss in the human amyloid precursor protein V717F (PDAPP) transgenic mouse. *The Journal of Neuroscience* **17**, 7053–7059.
- Jack CR, Knopman DS, Jagust WJ, Petersen RC, Weiner MW, Aisen PS, Shaw LM, Vemuri P, Wiste HJ, Weigand SD, Lesnick TG, Pankratz VS, Donohue MC and Trojanowski JQ** (2013) Tracking pathophysiological processes in Alzheimer's disease: an updated hypothetical model of dynamic biomarkers. *The Lancet Neurology* **12**, 207–216.
- Jackson GR, Wiedau-Pazos M, Sang TK, Wagle N, Brown CA, Massachi S and Geschwind DH** (2002) Human wild-type tau interacts with wingless pathway components and produces neurofibrillary pathology in *Drosophila*. *Neuron* **34**, 509–519.
- Jackson RJ, Rudinskiy N, Herrmann AG, Croft S, Kim JSM, Petrova V, Ramos-Rodriguez JJ, Pitstick R, Wegmann S, Garcia-Alloza M, Carlson GA, Hyman BT and Spires-Jones TL** (2016) Human tau increases amyloid β plaque size but not amyloid β -mediated synapse loss in a novel mouse model of Alzheimer's disease. *European Journal of Neuroscience* **44**, 3056–3066.
- Janezic S, Threlfall S, Dodson PD, Dowie MJ, Taylor TN, Potgieter D, Parkkinen L, Senior SL, Anwar S, Ryan B, Deltheil T, Kosillo P, Cioroch M, Wagner K, Ansorge O, Bannerman DM, Bolam JP, Magill PJ, Cragg SJ and Wade-Martins R** (2013) Deficits in dopaminergic transmission precede neuron loss and dysfunction in a new Parkinson model. *Proceedings of the National Academy of Sciences of the United States of America* **110**, E4016–E4025.
- Jeganathan S, von Bergen M, Brutlach H, Steinhoff H-J and Mandelkow E** (2006) Global hairpin folding of tau in solution. *Biochemistry* **45**, 2283–2293.
- Jucker M** (2010) The benefits and limitations of animal models for translational research in neurodegenerative diseases. *Nature Medicine* **16**, 1210–1214.
- Jucker M and Walker LC** (2013) Self-propagation of pathogenic protein aggregates in neurodegenerative diseases. *Nature* **501**, 45–51.
- Jucker M and Walker LC** (2018) Propagation and spread of pathogenic protein assemblies in neurodegenerative diseases. *Nature Neuroscience* **21**, 1341–1349.
- Jung BC, Lim Y-J, Bae E-J, Lee JS, Choi MS, Lee MK, Lee H-J, Kim YS and Lee S-J** (2017a) Amplification of distinct α -synuclein fibril conformers through protein misfolding cyclic amplification. *Experimental & Molecular Medicine* **49**, e314.
- Jung SY, Choi JM, Rousseaux MWC, Malovannaya A, Kim JJ, Kutsera J, Wang Y, Huang Y, Zhu W, Maity S, Zoghbi HY and Qin J** (2017b) An anatomically resolved mouse brain proteome reveals Parkinson disease-relevant pathways. *Molecular & Cellular Proteomics* **16**, 581–593.
- Kalinderi K, Bostantjopoulou S and Fidani L** (2016) The genetic background of Parkinson's disease: current progress and future prospects. *Acta Neurologica Scandinavica* **134**, 314–326.
- Kaminski Schierle GS, Bertoncini CW, Chan FTS, van der Goot AT, Schwedler S, Skepper J, Schlachter S, van Ham T, Esposito A, Kumita JR, Nollen EAA, Dobson CM and Kaminski CF** (2011a) A FRET sensor for non-invasive imaging of amyloid formation *in vivo*. *ChemPhysChem* **12**, 673–680.
- Kaminski Schierle GS, Van De Linde S, Erdelyi M, Esbjörner EK, Klein T, Rees E, Bertoncini CW, Dobson CM, Sauer M and Kaminski CF** (2011b) *In situ* measurements of the formation and morphology of intracellular β -amyloid fibrils by super-resolution fluorescence imaging. *Journal of the American Chemical Society* **133**, 12902–12905.
- Karpinar DP, et al.** (2009) Pre-fibrillar alpha-synuclein variants with impaired beta-structure increase neurotoxicity in Parkinson's disease models. *The EMBO Journal* **28**, 3256–3268.
- Karran E and De Strooper B** (2016) The amyloid cascade hypothesis: are we poised for success or failure? *Journal of Neurochemistry* **139**, 237–252.
- Kasahara J, Choudhury ME, Nishikawa N, Tanabe A, Tsuji R, Zhou Y, Ogawa M, Yokoyama H, Tanaka J and Nomoto M** (2017) Neurotoxin 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-induced animal models of Parkinson's disease. In Conn PM (ed). *Animal Models for the Study of Human Disease*, 2nd Edn. pp. Cambridge MA: Academic Press, pp. 1087–1108.
- Kempf SJ, Metaxas A, Ibanez-Vea M, Darvesh S, Finsen B and Larsen MR** (2016) An integrated proteomics approach shows synaptic plasticity changes in an APP/PS1 Alzheimer's mouse model. *Oncotarget* **7**, 33627–33648.
- Ke YD, et al.** (2019) CNS cell type-specific gene profiling of P301S tau transgenic mice identifies genes dysregulated by progressive tau accumulation. *Journal of Biological Chemistry* **294**, 14149–14162.
- Khabirova E, Moloney A, Marciniak SJ, Williams J, Lomas DA, Oliver SG, Favrin G, Sattelle DB and Crowther DC** (2014) The TRiC/CCT chaperone is implicated in Alzheimer's disease based on patient GWAS and an RNAi screen in A β -expressing *Caenorhabditis elegans*. *PLoS ONE* **9**, 1–13.
- Khurana V and Lindquist S** (2010) Modelling neurodegeneration in *Saccharomyces cerevisiae*: why cook with baker's yeast? *Nature Reviews. Neuroscience* **11**, 436–449.
- Khurana V, et al.** (2017) Genome-scale networks link neurodegenerative disease genes to α -synuclein through specific molecular pathways. *Cell Systems* **4**, 157–170.

- Kim YE, Hipp MS, Bracher A, Hayer-Hartl M and Ulrich Hartl F (2013) Molecular chaperone functions in protein folding and proteostasis. *Annual Review of Biochemistry* **82**, 323–355.
- Kirik D, Rosenblad C, Burger C, Lundberg C, Johansen TE, Muzyczka N, Mandel RJ and Björklund A (2002) Parkinson-like neurodegeneration induced by targeted overexpression of α -synuclein in the nigrostriatal system. *The Journal of Neuroscience* **22**, 2780–2791.
- Kirik D, Annett LE, Burger C, Muzyczka N, Mandel RJ and Björklund A (2003) Nigrostriatal alpha-synucleinopathy induced by viral vector-mediated overexpression of human alpha-synuclein: a new primate model of Parkinson's disease. *Proceedings of the National Academy of Sciences of the United States of America* **100**, 2884–2889.
- Kjaergaard M, Dear AJ, Kundel F, Meisl G, Knowles TPJ and Klenerman D (2018) Oligomer diversity during the aggregation of the repeat-region of tau. *ACS Chemical Neuroscience* **9**, 3060–3071.
- Klein RL, King MA, Hamby ME and Meyer EM (2002) Dopaminergic cell loss induced by human A30P α -synuclein gene transfer to the rat substantia nigra. *Human Gene Therapy* **13**, 605–612.
- Knott GJ and Doudna JA (2018) CRISPR-Cas guides the future of genetic engineering. *Science (New York, N.Y.)* **361**, 866–869.
- Knowles TPJ, Waudby CA, Devlin GL, Cohen SIA, Aguzzi A, Vendruscolo M, Terentjev EM, Welland ME and Dobson CM (2009) An analytical solution to the kinetics of breakable filament assembly. *Science (New York, N.Y.)* **326**, 1533–1537.
- Knowles TPJ, Vendruscolo M and Dobson CM (2014) The amyloid state and its association with protein misfolding diseases. *Nature Reviews. Molecular Cell Biology* **15**, 384–396.
- Korhonen P, van Groen T, Thornell A, Kyrylenko S, Soininen M-L, Ojala J, Peltomaa E, Tanila H, Salminen A, Mandelkow EM and Soininen H (2011) Characterization of a novel transgenic rat carrying human tau with mutation P301L. *Neurobiology of Aging* **32**, 2314–2315.
- Koson P, Zilka N, Kovac A, Kovacech B, Korenova M, Filipcik P and Novak M (2008) Truncated tau expression levels determine life span of a rat model of tauopathy without causing neuronal loss or correlating with terminal neurofibrillary tangle load. *European Journal of Neuroscience* **28**, 239–246.
- Kraemer BC, Zhang B, Leverenz JB, Thomas JH, Trojanowski JQ and Schellenberg GD (2003) Neurodegeneration and defective neurotransmission in a *Caenorhabditis elegans* model of tauopathy. *Proceedings of the National Academy of Sciences of the United States of America* **100**, 9980–9985.
- Kraemer BC, Burgess JK, Chen JH, Thomas JH and Schellenberg GD (2006) Molecular pathways that influence human tau-induced pathology in *Caenorhabditis elegans*. *Human Molecular Genetics* **15**, 1483–1496.
- Kuhle J, Barro C, Andreasson U, Derfuss T, Lindberg R, Sandelius Å, Liman V, Norgren N, Blennow K and Zetterberg H (2016) Comparison of three analytical platforms for quantification of the neurofilament light chain in blood samples: ELISA, electrochemiluminescence immunoassay and Simoa. *Clinical Chemistry and Laboratory Medicine* **54**, 1655.
- Kumar S, Tepper K, Kaniyappan S, Biernat J, Wegmann S, Mandelkow EM, Müller DJ and Mandelkow E (2014) Stages and conformations of the tau repeat domain during aggregation and its effect on neuronal toxicity. *Journal of Biological Chemistry* **289**, 20318–20332.
- Kummer MP and Heneka MT (2014) Truncated and modified amyloid- β species. *Alzheimer's Research & Therapy* **6**, 28.
- Kuwahara T, Koyama A, Gengyo-Ando K, Masuda M, Kowa H, Tsunoda M, Mitani S and Iwatsubo T (2006) Familial Parkinson mutant α -synuclein causes dopamine neuron dysfunction in transgenic *Caenorhabditis elegans*. *Journal of Biological Chemistry* **281**, 334–340.
- Kuwahara T, Koyama A, Koyama S, Yoshina S, Ren CH, Kato T, Mitani S and Iwatsubo T (2008) A systematic RNAi screen reveals involvement of endocytic pathway in neuronal dysfunction in α -synuclein transgenic *C. elegans*. *Human Molecular Genetics* **17**, 2997–3009.
- Labbadia J and Morimoto RI (2015) The biology of proteostasis in aging and disease. *Annual Review of Biochemistry* **84**, 435–464.
- Lachén-Montes M, González-Morales A, de Morentin XM, Pérez-Valderrama E, Ausín K, Zelaya MV, Serna A, Aso E, Ferrer I, Fernández-Irigoyen J and Santamaría E (2016) An early dysregulation of FAK and MEK/ERK signaling pathways precedes the β -amyloid deposition in the olfactory bulb of APP/PS1 mouse model of Alzheimer's disease. *Journal of Proteomics* **148**, 149–158.
- LaFerla FM, Green KN and Oddo S (2007) Intracellular amyloid- β in Alzheimer's disease. *Nature Reviews Neuroscience* **8**, 499–509.
- Laine RF, Sinnige T, Ma KY, Haack AJ, Poudel C, Gaida P, Curry N, Perni M, Nollen EAA, Dobson CM, Vendruscolo M, Kaminski Schierle GS and Kaminski CF (2019) Fast fluorescence lifetime imaging reveals the aggregation processes of α -synuclein and polyglutamine in aging *Caenorhabditis elegans*. *ACS Chemical Biology* **14**, 1628–1636.
- Lakso M, Vartiainen S, Moilanen A-M, Sirviö J, Thomas JH, Nass R, Blakely RD and Wong G (2003) Dopaminergic neuronal loss and motor deficits in *Caenorhabditis elegans* overexpressing human α -synuclein. *Journal of Neurochemistry* **86**, 165–172.
- Lancaster MA, Corsini NS, Wolfinger S, Gustafson EH, Phillips AW, Burkard TR, Otani T, Livesey FJ and Knoblich JA (2017) Guided self-organization and cortical plate formation in human brain organoids. *Nature Biotechnology* **35**, 659.
- Landel V, Baranger K, Virard I, Loriod B, Khrestchatsky M, Rivera S, Benech P and Féron F (2014) Temporal gene profiling of the 5XFAD transgenic mouse model highlights the importance of microglial activation in Alzheimer's disease. *Molecular Neurodegeneration* **9**, 33.
- Lauwers E, Bequé D, Van Laere K, Nuyts J, Bormans G, Mortelmans L, Casteels C, Vercaemmen L, Bockstaal O, Nuttin B, Debysse Z and Baekelandt V (2007) Non-invasive imaging of neuropathology in a rat model of α -synuclein overexpression. *Neurobiology of Aging* **28**, 248–257.
- Lee VM-Y, Goedert M and Trojanowski JQ (2001) Neurodegenerative tauopathies. *Annual Review of Neuroscience* **24**, 1121–1159.
- Lee VM-Y, Kenyon TK and Trojanowski JQ (2005) Transgenic animal models of tauopathies. *Biochimica et Biophysica Acta (BBA) – Molecular Basis of Disease* **1739**, 251–259.
- Leon WC, Canneva F, Partridge V, Allard S, Ferretti MT, Dewilde A, Vercauteren F, Atifeh R, Ducatenzeiler A, Klein W, Szyf M, Alhonen L and Cuelllo AC (2010) A novel transgenic rat model with a full Alzheimer's-like amyloid pathology displays pre-plaque intracellular amyloid- β -associated cognitive impairment. *Journal of Alzheimer's Disease* **20**, 113–126.
- Leroy K, Ando K, Laporte V, Dedecker R, Suain V, Authélet M, Héraud C, Pierrot N, Yilmaz Z, Octave JN and Brion JP (2012) Lack of tau proteins rescues neuronal cell death and decreases amyloidogenic processing of APP in APP/PS1 mice. *American Journal of Pathology* **181**, 1928–1940.
- Lesné S, Ming TK, Kotilinek L, Kaye R, Glabe CG, Yang A, Gallagher M and Ashe KH (2006) A specific amyloid- β protein assembly in the brain impairs memory. *Nature* **440**, 352–357.
- Lewis J, McGowan E, Rockwood J, Melrose H, Nacharaju P, Van Slegtenhorst M, Gwinn-Hardy K, Paul Murphy M, Baker M, Yu X, Duff K, Hardy J, Corral A, Lin W-L, Yen S-H, Dickson DW, Davies P and Hutton M (2000) Neurofibrillary tangles, amyotrophy and progressive motor disturbance in mice expressing mutant (P301L) tau protein. *Nature Genetics* **25**, 402–405.
- Lewis J, Dickson DW, Lin W, Chisholm L, Corral A, Jones G, Yen S, Sahara N, Skipper L, Yager D, Eckman C, Hardy J, Hutton M and McGowan E (2001) Enhanced neurofibrillary degeneration in transgenic mice expressing mutant tau and APP. *Science (New York, N.Y.)* **293**, 1487–1491.
- Liang J, Clark-Dixon C, Wang S, Flower TR, Williams-Hart T, Zweig R, Robinson LC, Tatchell K and Witt SN (2008) Novel suppressors of α -synuclein toxicity identified using yeast. *Human Molecular Genetics* **17**, 3784–3795.
- Lin X, Parisiadou L, Sgobio C, Liu G, Yu J, Sun L, Shim H, Gu X-L, Luo J, Long C-X, Ding J, Mateo Y, Sullivan PH, Wu L-G, Goldstein DS, Lovinger D and Cai H (2012) Conditional expression of Parkinson's disease-related mutant alpha-synuclein in the midbrain dopaminergic neurons causes progressive neurodegeneration and degradation of transcription factor nuclear receptor related 1. *The Journal of Neuroscience* **32**, 9248–9264.
- Link CD (1995) Expression of human beta-amyloid peptide in transgenic *Caenorhabditis elegans*. *Proceedings of the National Academy of Sciences of the United States of America* **92**, 9368–9372.

- Link CD (2006) *C. elegans* models of age-associated neurodegenerative diseases: lessons from transgenic worm models of Alzheimer's disease. *Experimental Gerontology* **41**, 1007–1013.
- Link CD, Johnson CJ, Fonte V, Paupard MC, Hall DH, Styren S, Mathis CA and Klunk WE (2001) Visualization of fibrillar amyloid deposits in living, transgenic *Caenorhabditis elegans* animals using the sensitive amyloid dye, X-34. *Neurobiology of Aging* **22**, 217–226.
- Liu Y, Beyer A and Aebersold R (2016) On the dependency of cellular protein levels on mRNA abundance. *Cell* **165**, 535–550.
- Lo Bianco C, Ridet J-L, Schneider BL, Deglon N and Aebischer P (2002) Alpha-synucleinopathy and selective dopaminergic neuron loss in a rat lentiviral-based model of Parkinson's disease. *Proceedings of the National Academy of Sciences of the United States of America* **99**, 10813–10818.
- Luheshi LM, Tartaglia GG, Brorsson AC, Pawar AP, Watson IE, Chiti F, Vendruscolo M, Lomas DA, Dobson CM and Crowther DC (2007) Systematic *in vivo* analysis of the intrinsic determinants of amyloid β pathogenicity. *PLoS Biology* **5**, 2493–2500.
- Luheshi LM, Hoyer W, de Barros TP, van Hård ID, Brorsson AC, Macao B, Persson C, Crowther DC, Lomas DA, Ståhl S, Dobson CM and Hård T (2010) Sequestration of the A β peptide prevents toxicity and promotes degradation *in vivo*. *PLoS Biology* **8**, e1000334.
- Luna E, Decker SC, Riddle DM, Caputo A, Zhang B, Cole T, Caswell C, Xie SX, Lee VMY and Luk KC (2018) Differential α -synuclein expression contributes to selective vulnerability of hippocampal neuron subpopulations to fibril-induced toxicity. *Acta Neuropathologica* **135**, 855–875.
- Lüthi U, Schaerer-Brodbeck C, Tanner S, Middendorp O, Edler K and Barberis A (2003) Human β -secretase activity in yeast detected by a novel cellular growth selection system. *Biochimica et Biophysica Acta – General Subjects* **1620**, 167–178.
- Maeda S, Djukic B, Taneja P, Yu G, Lo I, Davis A, Craft R, Guo W, Wang X, Kim D, Ponnusamy R, Gill TM, Masliah E and Mucke L (2016) Expression of A152T human tau causes age-dependent neuronal dysfunction and loss in transgenic mice. *EMBO Reports* **17**, 530–551.
- Mahley RW (1988) Apolipoprotein E: cholesterol transport protein with expanding role in cell biology. *Science (New York, N.Y.)* **240**, 622–630.
- Mahul-Mellier A-L, Bartscher J, Maharjan N, Weerens L, Croisier M, Kuttler F, Leleu M, Knott GW and Lashuel HA (2020) The process of Lewy body formation, rather than simply α -synuclein fibrillization, is one of the major drivers of neurodegeneration. *Proceedings of the National Academy of Sciences* **117**, 4971–4982.
- Mannini B, Habchi J, Chia S, Ruggeri FS, Perni M, Knowles TPJ, Dobson CM and Vendruscolo M (2018) Stabilization and characterization of cytotoxic A β 40 oligomers isolated from an aggregation reaction in the presence of zinc ions. *ACS Chemical Neuroscience* **9**, 2959–2971.
- Masliah E, Rockenstein E, Veinbergs I, Mallory M, Hashimoto M, Takeda A, Sagara Y, Sisk A and Mucke L (2000) Dopaminergic loss and inclusion body formation in alpha-synuclein mice: implications for neurodegenerative disorders. *Science (New York, N.Y.)* **287**, 1265–1269.
- McColl G, Roberts BR, Gunn AP, Perez KA, Tew DJ, Masters CL, Barnham KJ, Cherny RA and Bush AI (2009) The *Caenorhabditis elegans* Abeta1-42 model of Alzheimer disease predominantly expresses Abeta3-42. *Journal of Biological Chemistry* **284**, 22697–22702.
- McColl G, Roberts BR, Pukala TL, Kenche VB, Roberts CM, Link CD, Ryan TM, Masters CL, Barnham KJ, Bush AI and Cherny RA (2012) Utility of an improved model of amyloid-beta (A β 1-42) toxicity in *Caenorhabditis elegans* for drug screening for Alzheimer's disease. *Molecular neurodegeneration* **7**, 57.
- McGowan E, Pickford F, Kim J, Onstead L, Eriksen J, Yu C, Skipper L, Murphy MP, Beard J, Das P, Jansen K, DeLucia M, Lin WL, Dolios G, Wang R, Eckmann CB, Dickson DW, Hutton M, Hardy J and Golde T (2005) A β 42 is essential for parenchymal and vascular amyloid deposition in mice. *Neuron* **47**, 191–199.
- McGowan E, Eriksen J and Hutton M (2006) A decade of modeling Alzheimer's disease in transgenic mice. *Trends in Genetics* **22**, 281–289.
- Meisl G, Yang X, Hellstrand E, Frohm B, Kirkegaard JB, Cohen SIA, Dobson CM, Linse S and Knowles TPJ (2014) Differences in nucleation behavior underlie the contrasting aggregation kinetics of the A β 40 and A β 42 peptides. *Proceedings of the National Academy of Sciences of the United States of America* **111**, 9384–9389.
- Meisl G, Kirkegaard JB, Arosio P, Michaels TCT, Vendruscolo M, Dobson CM, Linse S and Knowles TPJ (2016a) Molecular mechanisms of protein aggregation from global fitting of kinetic models. *Nature Protocols* **11**, 252–272.
- Meisl G, Yang X, Frohm B, Knowles TPJ and Linse S (2016b) Quantitative analysis of intrinsic and extrinsic factors in the aggregation mechanism of Alzheimer-associated A β -peptide. *Scientific Reports* **6**, 18728.
- Merchant KM, Cedarbaum JM, Brundin P, Dave KD, Eberling J, Espay AJ, Hutten SJ, Javidnia M, Luthman J, Maetzler W, Menalled L, Reimer AN, Stoessl AJ and Weiner DM (2019) A proposed roadmap for Parkinson's disease proof of concept clinical trials investigating compounds targeting alpha-synuclein. *Journal of Parkinson's Disease* **9**, 31–61.
- Meyer V, Dinkel PD, Rickman Hager E and Margittai M (2014) Amplification of tau fibrils from minute quantities of seeds. *Biochemistry* **53**, 5804–5809.
- Mhatre SD, Michelson SJ, Gomes J, Tabb LP, Saunders AJ and Marenda DR (2014) Development and characterization of an aged onset model of Alzheimer's disease in *Drosophila melanogaster*. *Experimental Neurology* **261**, 772–781.
- Michaels TCT, Liu LX, Meisl G and Knowles TPJ (2017) Physical principles of filamentous protein self-assembly kinetics. *Journal of Physics: Condensed Matter* **29**, 15.
- Michaels TCT, Šarić A, Curk S, Bernfur K, Arosio P, Meisl G, Dear AJ, Cohen SIA, Dobson CM, Vendruscolo M, Linse S and Knowles TPJ (2020) Dynamics of oligomer populations formed during the aggregation of Alzheimer's A β 42 peptide. *Nature Chemistry* **12**, 445–451.
- Middendorp O, Ortler C, Neumann U, Paganetti P, Lüthi U and Barberis A (2004) Yeast growth selection system for the identification of cell-active inhibitors of β -secretase. *Biochimica et Biophysica Acta – General Subjects* **1674**, 29–39.
- Minde DP, Dunker AK and Lilley KS (2017) Time, space, and disorder in the expanding proteome universe. *Proteomics* **17**, 1600399.
- Miyasaka T, Ding Z, Gengyo-Ando K, Oue M, Yamaguchi H, Mitani S and Ihara Y (2005) Progressive neurodegeneration in *C. elegans* model of tauopathy. *Neurobiology of Disease* **20**, 372–383.
- Mocanu M-M, Nissen A, Eckermann K, Khlistunova I, Biernat J, Drexler D, Petrova O, Schonig K, Bujard H, Mandelkow E, Zhou L, Rune G and Mandelkow E-M (2008) The potential for β -structure in the repeat domain of tau protein determines aggregation, synaptic decay, neuronal loss, and coassembly with endogenous tau in inducible mouse models of tauopathy. *The Journal of Neuroscience* **28**, 737–748.
- Mudhar A, Shepherd D, Newman TA, Mildren P, Jukes JP, Squire A, Mears A, Berg S, MacKay D, Asuni AA, Bhat R and Lovestone S (2004) GSK-3 β inhibition reverses axonal transport defects and behavioural phenotypes in *Drosophila*. *Molecular Psychiatry* **9**, 522–530.
- Neuner SM, Wilmott LA, Hoffmann BR, Mozhui K and Kaczorowski CC (2017) Hippocampal proteomics defines pathways associated with memory decline and resilience in normal aging and Alzheimer's disease mouse models. *Behavioural Brain Research* **322**, 288–298.
- Nishimura I, Yang Y and Lu B (2004) PAR-1 kinase plays an initiator role in a temporally ordered phosphorylation process that confers tau toxicity in *Drosophila*. *Cell* **116**, 671–682.
- Nuber S, Harmuth F, Kohl Z, Adame A, Trejo M, Schönig K, Zimmermann F, Bauer C, Casadei N, Giel C, Calaminus C, Pichler BJ, Jensen PH, Müller CP, Amato D, Kornhuber J, Teismann P, Yamakado H, Takahashi R, Winkler J, Masliah E and Riess O (2013) A progressive dopaminergic phenotype associated with neurotoxic conversion of α -synuclein in BAC-transgenic rats. *Brain* **136**, 412–432.
- Nuber S, Rajsoobath M, Minakaki G, Winkler J, Müller CP, Ericsson M, Caldaroni B, Dettmer U and Selkoe DJ (2018) Abrogating native α -synuclein tetramers in mice causes a L-DOPA-responsive motor syndrome closely resembling Parkinson's disease. *Neuron* **100**, 75–90.
- Oakley H, Cole SL, Logan S, Maus E, Shao P, Craft J, Guillozet-Bongaarts A, Ohno M, Disterhoft J, Van Eldik L, Berry R and Vassar R (2006) Intraneuronal beta-amyloid aggregates, neurodegeneration, and neuron loss in transgenic mice with five familial Alzheimer's disease mutations: potential factors in amyloid plaque formation. *Journal of Neuroscience* **26**, 10129–10140.

- Oddo S, Caccamo A, Shepherd JD, Murphy MP, Golde TE, Kaye R, Metherate R, Mattson MP, Akbari Y and LaFerla FM (2003) Triple-transgenic model of Alzheimer's disease with plaques and tangles intracellular A β and synaptic dysfunction. *Neuron* **39**, 409–421.
- Oerton E and Bender A (2017) Concordance analysis of microarray studies identifies representative gene expression changes in Parkinson's disease: a comparison of 33 human and animal studies. *BMC Neurology* **17**, 58.
- Oliveras-Salva M, Van Der Perren A, Casadei N, Stroobants S, Nuber S, D'Hooge R, Van Den Haute C and Baekelandt V (2013) RAAV2/7 vector-mediated overexpression of alpha-synuclein in mouse substantia nigra induces protein aggregation and progressive dose-dependent neurodegeneration. *Molecular Neurodegeneration* **8**, 44.
- Ordonez DG, Lee MK and Feany MB (2018) α -synuclein induces mitochondrial dysfunction through spectrin and the actin cytoskeleton. *Neuron* **97**, 108–124.
- Ossenkuppe R, et al. (2016) Tau PET patterns mirror clinical and neuroanatomical variability in Alzheimer's disease. *Brain* **139**, 1551–1567.
- Otani T, Marchetto MC, Gage FH, Simons BD and Livesey FJ (2016) 2D and 3D stem cell models of primate cortical development identify species-specific differences in progenitor behavior contributing to brain size. *Cell Stem Cell* **18**, 467–480.
- Outeiro TF and Lindquist S (2003) Yeast cells provide insight into alpha-synuclein biology and pathobiology. *Science (New York, N.Y.)* **302**, 1772–1775.
- Pace MC, Xu G, Fromholt S, Howard J, Crosby K, Giasson BI, Lewis J and Borchelt DR (2018) Changes in proteome solubility indicate widespread proteostatic disruption in mouse models of neurodegenerative disease. *Acta Neuropathologica* **136**, 919–938.
- Parthasarathy S, Inoue M, Xiao Y, Matsumura Y, Nabeshima Y, Hoshi M and Ishii Y (2015) Structural insight into an Alzheimer's brain-derived spherical assembly of amyloid β by solid-state NMR. *Journal of the American Chemical Society* **137**, 6480–6483.
- Pauwels K, Williams TL, Morris KL, Jonckheere W, Vandersteen A, Kelly G, Schymkowitz J, Rousseau F, Pastore A, Serpell LC and Broersen K (2012) Structural basis for increased toxicity of pathological A β 42:a β 40 ratios in Alzheimer disease. *Journal of Biological Chemistry* **287**, 5650–5660.
- Pereson S, Wils H, Kleinberger G, McGowan E, Vandewoestyne M, Van Broeck B, Joris G, Cuijt I, Deforce D, Hutton M, Van Broeckhoven C and Kumar-Singh S (2009) Progranulin expression correlates with dense-core amyloid plaque burden in Alzheimer disease mouse models. *The Journal of pathology* **219**, 173–181.
- Perez M, Ribe E, Rubio A, Lim F, Moran MA, Ramos PG, Ferrer I, Isla MTG and Avila J (2005) Characterization of a double (amyloid precursor protein-tau) transgenic: tau phosphorylation and aggregation. *Neuroscience* **130**, 339–347.
- Periquet M, Fulga T, Myllykangas L, Schlossmacher MG and Feany MB (2007) Aggregated alpha-synuclein mediates dopaminergic neurotoxicity in vivo. *Journal of Neuroscience* **27**, 3338–3346.
- Perni M, Challa PK, Kirkegaard JB, Limbocker R, Koopman M, Hardenberg MC, Sormanni P, Muller T, Saar KL, Roode LWY, Habchi J, Vecchi G, Fernando NW, Casford S, Nollen EAA, Vendruscolo M, Dobson CM and Knowles TPJ (2018) Massively parallel *C. elegans* tracking provides multi-dimensional fingerprints for phenotypic discovery. *Journal of Neuroscience Methods* **306**, 57–67.
- Pesah Y, Burgess H, Middlebrooks B, Ronningen K, Prosser J, Tirunaguru V, Zysk J and Mardon G (2005) Whole-mount analysis reveals normal numbers of dopaminergic neurons following misexpression of alpha-synuclein in *Drosophila*. *Genesis (New York, N.Y.: 2000)* **41**, 154–159.
- Pinotsi D, Michel CH, Buell AK, Laine RF, Mahou P, Dobson CM, Kaminski CF and Kaminski Schierle GS (2016) Nanoscopic insights into seeding mechanisms and toxicity of α -synuclein species in neurons. *Proceedings of the National Academy of Sciences of the United States of America* **113**, 3815–3819.
- Platt B, Drever B, Koss D, Stoppelkamp S, Jyoti A, Plano A, Utan A, Merrick G, Ryan D, Melis V, Wan H, Mingarelli M, Porcu E, Scrocchi L, Welch A and Riedel G (2011) Abnormal cognition, sleep, EEG and brain metabolism in a novel knock-in Alzheimer mouse, PLB1. *PLoS ONE* **6**, e27068.
- Polydoro M, Acker CM, Duff K, Castillo PE and Davies P (2009) Age-dependent impairment of cognitive and synaptic function in the htau mouse model of tau pathology. *The Journal of Neuroscience* **29**, 10741–10749.
- Pooler AM, Polydoro M, Maury EA, Nicholls SB, Reddy SM, Wegmann S, William C, Saqran L, Cagsal-Getkin O, Pitstick R, Beier DR, Carlson GA, Spires-Jones TL and Hyman BT (2015) Amyloid accelerates tau propagation and toxicity in a model of early Alzheimer's disease. *Acta Neuropathologica Communications* **3**, 1–11.
- Prufing K, Voigt A and Schulz JB (2013) *Drosophila melanogaster* as a model organism for Alzheimer's disease. *Molecular Neurodegeneration* **8**, 35.
- Radde R, Bolmont T, Kaeser SA, Coomaraswamy J, Lindau D, Stoltze L, Calhoun ME, Jaggi F, Wolburg H, Gengler S, Haass C, Ghetti B, Czech C, Holscher C, Mathews PM and Jucker M (2006) A β 42-driven cerebral amyloidosis in transgenic mice reveals early and robust pathology. *EMBO Reports* **7**, 940–946.
- Ramsden M, Kotilinek L, Forster C, Paulson J, McGowan E, SantaCruz K, Guimaraes A, Yue M, Lewis J, Carlson G, Hutton M and Ashe KH (2005) Age-dependent neurofibrillary tangle formation, neuron loss, and memory impairment in a mouse model of human tauopathy (P301L). *The Journal of Neuroscience* **25**, 10637–10647.
- Rencus-Lazar S, DeRowe Y, Adsi H, Gazit E and Laor D (2019) Yeast models for the study of amyloid-associated disorders and development of future therapy. *Frontiers in Molecular Biosciences* **6**, 15.
- Rhein V, Song X, Wiesner A, Ittner LM, Baysang G, Meier F, Ozmen L, Bluethmann H, Drose S, Brandt U, Savaskan E, Czech C, Gotz J and Eckert A (2009) Amyloid- β and tau synergistically impair the oxidative phosphorylation system in triple transgenic Alzheimer's disease mice. *Proceedings of the National Academy of Sciences of the United States of America* **106**, 20057–20062.
- Ribe EM, Perez M, Puig B, Gich I, Lim F, Cuadrado M, Sesma T, Catena S, Sanchez B, Nieto M, Gomez-Ramos P, Moran MA, Cabodevilla F, Samaranch L, Ortiz L, Perez A, Ferrer I, Avila J and Gomez-Isla T (2005) Accelerated amyloid deposition, neurofibrillary degeneration and neuronal loss in double mutant APP/tau transgenic mice. *Neurobiology of Disease* **20**, 814–822.
- Richards JG, Higgins GA, Ouagazzal AM, Ozmen L, Kew JN, Bohrmann B, Malherbe P, Brockhaus M, Loetscher H, Czech C, Huber G, Bluethmann H, Jacobsen H and Kemp JA (2003) PS2APP transgenic mice, coexpressing hPS2mut and hAPPswe, show age-related cognitive deficits associated with discrete brain amyloid deposition and inflammation. *Journal of Neuroscience* **23**, 8989–9003.
- Richfield EK, Thiruchelvam MJ, Cory-Slechta DA, Wuertzer C, Gainetdinov RR, Caron MG, Di Monte DA and Federoff HJ (2002) Behavioral and neurochemical effects of wild-type and mutated human α -synuclein in transgenic mice. *Experimental Neurology* **175**, 35–48.
- Rincon-Limas DE, Jensen K and Fernandez-Funez P (2012) *Drosophila* models of proteinopathies: the little fly that could. *Current Pharmaceutical Design* **18**, 1108–1122.
- Roberson ED, Scarce-Levie K, Palop JJ, Yan F, Chen IH, Wu T, Gerstein H, Yu G-Q and Mucke L (2007) Reducing endogenous tau ameliorates amyloid β -induced deficits in an Alzheimer's disease mouse model. *Science (New York, N.Y.)* **316**, 750–754.
- Roberts H and Brown D (2015) Seeking a mechanism for the toxicity of oligomeric α -synuclein. *Biomolecules* **5**, 282–305.
- Rockenstein E, Mallory M, Hashimoto M, Song D, Shults CW, Lang I and Masliah E (2002) Differential neuropathological alterations in transgenic mice expressing α -synuclein from the platelet-derived growth factor and Thy-1 promoters. *Journal of Neuroscience Research* **68**, 568–578.
- Saborio GP, Permanne B and Soto C (2001) Sensitive detection of pathological prion protein by cyclic amplification of protein misfolding. *Nature* **411**, 810–813.
- Saito T, Matsuba Y, Mihira N, Takano J, Nilsson P, Itoharu S, Iwata N and Saido TC (2014) Single App knock-in mouse models of Alzheimer's disease. *Nature Neuroscience* **17**, 661–663.
- Saito T, Mihira N, Matsuba Y, Sasaguri H, Hashimoto S, Narasimhan S, Zhang B, Murayama S, Higuchi M, Lee VMY, Trojanowski JQ and Saido TC (2019) Humanization of the entire murine Mapt gene provides a murine model of pathological human tau propagation. *Journal of Biological Chemistry* **294**, 12754–12765.

- SantaCruz K, Lewis J, Spires T, Paulson J, Kotilinek L, Ingelsson M, Guimaraes A, DeTure M, Ramsden M, McGowan E, Forster C, Yue M, Orne J, Janus C, Mariash A, Kushowski M, Hyman B, Hutton M and Ashe KH (2005) Tau suppression in a neurodegenerative mouse model improves memory function. *Science (New York, N.Y.)* **309**, 476–481.
- Sarantseva S, Timoshenko S, Bolshakova O, Karaseva E, Rodin D, Schwarzman AL and Vitek MP (2009) Apolipoprotein E-mimetics inhibit neurodegeneration and restore cognitive functions in a transgenic *Drosophila* model of Alzheimer's disease. *PLoS ONE* **4**, e8191.
- Sasaguri H, Nilsson P, Hashimoto S, Nagata K, Saito T, De Strooper B, Hardy J, Vassar R, Winblad B and Saido TC (2017) APP Mouse models for Alzheimer's disease preclinical studies. *The EMBO Journal* **36**, 2473–2487.
- Sato H, Kato T and Arawaka S (2013) The role of Ser129 phosphorylation of α -synuclein in neurodegeneration of Parkinson's disease: a review of *in vivo* models. *Reviews in the Neurosciences* **24**, 115.
- Saul A, Sprenger F, Bayer TA and Wirths O (2013) Accelerated tau pathology with synaptic and neuronal loss in a novel triple transgenic mouse model of Alzheimer's disease. *Neurobiology of Aging* **34**, 2564–2573.
- Savas JN, Wang YZ, DeNardo LA, Martinez-Bartolome S, McClatchy DB, Hark TJ, Shanks NF, Cozzolino KA, Lavallée-Adam M, Smukowski SN, Park SK, Kelly JW, Koo EH, Nakagawa T, Masliah E, Ghosh A and Yates JR (2017) Amyloid accumulation drives proteome-wide alterations in mouse models of Alzheimer's disease-like pathology. *Cell Reports* **21**, 2614–2627.
- Schnorrenberg S, Grotjohann T, Vorbrüggen G, Herzig A, Hell SW and Jakobs S (2016) *In vivo* super-resolution RESOLFT microscopy of *Drosophila melanogaster*. *eLife* **5**, e15567.
- Schubert OT, Röst HL, Collins BC, Rosenberger G and Aebersold R (2017) Quantitative proteomics: challenges and opportunities in basic and applied research. *Nature Protocols* **12**, 1289–1294.
- Selkoe DJ and Hardy J (2016) The amyloid hypothesis of Alzheimer's disease at 25 years. *EMBO Molecular Medicine* **8**, 595–608.
- Sevigny J, Chiao P, Bussière T, Weinreb PH, Williams L, Maier M, Dunstan R, Salloway S, Chen T, Ling Y, O'Gorman J, Qian F, Arastu M, Li M, Chollate S, Brennan MS, Quintero-Monzon O, Scannevin RH, Arnold HM, Engber T, Rhodes K, Ferrero J, Hang Y, Mikulskis A, Grimm J, Hock C, Nitsch RM and Sandrock A (2016) The antibody aducanumab reduces A β plaques in Alzheimer's disease. *Nature* **537**, 50–56.
- Seynnaeve D, Del Vecchio M, Fruhmann G, Verelst J, Cools M, Beckers J, Mulvihill DP, Winderickx J and Franssens V (2018) Recent insights on Alzheimer's disease originating from yeast models. *International Journal of Molecular Sciences* **19**, 1947.
- Shahmoradian SH, Lewis AJ, Genou C, Hench J, Moors TE, Navarro PP, Castaño-Diez D, Schweighauser G, Graff-Meyer A, Goldie KN, Sütterlin R, Huisman E, Ingrassia A, de Gier Y, Rozemuller AJM, Wang J, Paeppe A, De Erny J, Staempfli A, Hoernschemeyer J, Großberischkamp F, Niedieker D, El-Mashtoly SF, Quadri M, Van IJcken WJF, Bonifati V, Gerwert K, Bohrmann B, Frank S, Britschgi M, Stahlberg H, Van de Berg WDJ and Lauer ME (2019) Lewy pathology in Parkinson's disease consists of crowded organelles and lipid membranes. *Nature Neuroscience* **22**, 1099–1109.
- Shahnawaz M, Tokuda T, Waragai M, Mendez N, Ishii R, Trenkwalder C, Mollenhauer B and Soto C (2017) Development of a biochemical diagnosis of Parkinson disease by detection of α -synuclein misfolded aggregates in cerebrospinal fluid. *JAMA Neurology* **74**, 163–172.
- Shammas SL, Garcia GA, Kumar S, Kjaergaard M, Horrocks MH, Shivji N, Eva M, Knowles TPJ, Eckhard M and Klenerman D (2015) A mechanistic model of tau amyloid aggregation based on direct observation of oligomers. *Nature Communications* **6**, 7025.
- Shaye DD and Greenwald I (2011) Ortholist: a compendium of *C. elegans* genes with human orthologs. *PLoS ONE* **6**, e20085.
- Shin Y and Brangwynne CP (2017) Liquid phase condensation in cell physiology and disease. *Science (New York, N.Y.)* **357**, eaaf4382.
- Shulman JM and Feany MB (2003) Genetic modifiers of tauopathy in *Drosophila*. *Genetics* **165**, 1233–1242.
- Shulman JM, Chipendo P, Chibnik LB, Aubin C, Tran D, Keenan BT, Kramer PL, Schneider JA, Bennett DA, Feany MB and De Jager PL (2011) Functional screening of Alzheimer pathology genome-wide association signals in *Drosophila*. *American Journal of Human Genetics* **88**, 232–238.
- Shulman JM, Imboywa S, Giagtzoglou N, Powers MP, Hu Y, Devenport D, Chipendo P, Chibnik LB, Diamond A, Perrimon N, Brown NH, De Jager PL and Feany MB (2014) Functional screening in *Drosophila* identifies Alzheimer's disease susceptibility genes and implicates tau-mediated mechanisms. *Human Molecular Genetics* **23**, 870–877.
- Sin O, Michels H and Nollen EAA (2014) Genetic screens in *Caenorhabditis elegans* models for neurodegenerative diseases. *Biochimica et Biophysica Acta* **1842**, 1951–1959.
- Sinnige T, Ciryam P, Casford S, Dobson CM, De Bono M and Vendruscolo M (2019) Expression of the amyloid- β peptide in a single pair of *C. elegans* sensory neurons modulates the associated behavioural response. *PLoS ONE* **14**, e0217746.
- Sinnige T, Yu A and Morimoto RI (2020) Challenging proteostasis: role of the chaperone network to control aggregation-prone proteins in human disease. In Mendillo ML, Pincus D and Scherz-Shouval R (eds). *HSF1 and Molecular Chaperones in Biology and Cancer*. Cham: Springer, pp. 53–68.
- Smith DJ (2009) Mitochondrial dysfunction in mouse models of Parkinson's disease revealed by transcriptomics and proteomics. *Journal of Bioenergetics and Biomembranes* **41**, 487–491.
- Smith MG and Snyder M (2006) Yeast as a model for human disease. *Current Protocols in Human Genetics* **48**, 15.6.1–15.6.8.
- Sofola O, Kerr F, Rogers I, Killick R, Augustin H, Gandy C, Allen MJ, Hardy J, Lovestone S and Partridge L (2010) Inhibition of GSK-3 ameliorates A β pathology in an adult-onset *Drosophila* model of Alzheimer's disease. *PLoS Genetics* **6**, e1001087.
- Sormanni P, Aprile FA and Vendruscolo M (2015) Rational design of antibodies targeting specific epitopes within intrinsically disordered proteins. *Proceedings of the National Academy of Sciences* **2015**, 201422401.
- Speretta E, Jahn TR, Tartaglia GG, Favrin G, Barros TP, Imarisio S, Lomas DA, Luheshi LM, Crowther DC and Dobson CM (2012) Expression in *Drosophila* of tandem amyloid β peptides provides insights into links between aggregation and neurotoxicity. *Journal of Biological Chemistry* **287**, 20748–20754.
- Spillantini MG, Crowther RA, Jakes R, Cairns NJ, Lantos PL and Goedert M (1998a) Filamentous α -synuclein inclusions link multiple system atrophy with Parkinson's disease and dementia with Lewy bodies. *Neuroscience Letters* **251**, 205–208.
- Spillantini MG, Crowther RA, Kamphorst W, Heutink P and van Swieten JC (1998b) Tau pathology in two Dutch families with mutations in the microtubule-binding region of tau. *The American Journal of Pathology* **153**, 1359–1363.
- Spillantini MG, Crowther RA, Jakes R, Hasegawa M and Goedert M (1998c) α -Synuclein in filamentous inclusions of Lewy bodies from Parkinson's disease and dementia with Lewy bodies. *Proceedings of the National Academy of Sciences of the United States of America* **95**, 6469–6473.
- Spires-Jones TL and Hyman B (2014) The intersection of amyloid beta and tau at synapses in Alzheimer's disease. *Neuron* **82**, 756–771.
- Stroud JC, Liu C, Teng PK and Eisenberg D (2012) Toxic fibrillar oligomers of amyloid- β have cross- β structure. *Proceedings of the National Academy of Sciences* **109**, 7717–7722.
- Su LJ, Auluck PK, Outeiro TF, Yeger-Iotem E, Kritzer JA, Tardiff DF, Strathearn KE, Liu F, Cao S, Hamamichi S, Hill KJ, Caldwell KA, Bell GW, Fraenkel E, Cooper AA, Caldwell GA, Mccaffery JM, Rochet JC and Lindquist S (2010) Compounds from an unbiased chemical screen reverse both ER-to-Golgi trafficking defects and mitochondrial dysfunction in Parkinson's disease models. *Disease Models & Mechanisms* **208**, 194–208.
- Sun M and Chen L (2014) Studying tauopathies in *Drosophila*: a fruitful model. *Experimental Neurology* **274**, 52–57.
- Surmeier DJ, Obeso JA and Halliday GM (2017) Selective neuronal vulnerability in Parkinson disease. *Nature Reviews Neuroscience* **18**, 101–113.
- Sydow A, Hochgräfe K, Könen S, Cadinu D, Matenia D, Petrova O, Joseph M, Dennissen FJ and Mandelkow EM (2016) Age-dependent neuroinflammation and cognitive decline in a novel Ala152Thr-Tau transgenic mouse model of PSP and AD. *Acta Neuropathologica Communications* **4**, 17.

- Takano M, Yamashita T, Nagano K, Otani M, Maekura K, Kamada H, Tsunoda S, Tsutsumi Y, Tomiyama T, Mori H, Matsuura K and Matsuyama S (2013) Proteomic analysis of the hippocampus in Alzheimer's disease model mice by using two-dimensional fluorescence difference in gel electrophoresis. *Neuroscience Letters* **534**, 85–89.
- Tan L, Schedl P, Song H-J, Garza D and Konsolaki M (2008) The toll→NFκB signaling pathway mediates the neuropathological effects of the human Alzheimer's Aβ42 polypeptide in *Drosophila*. *PLoS ONE* **3**, e3966.
- Tenreiro S, Franssens V, Winderickx J and Outeiro TF (2017) Yeast models of Parkinson's disease-associated molecular pathologies. *Current Opinion in Genetics and Development* **44**, 74–83.
- Teo E, Ravi S, Barardo D, Kim H-S, Fong S, Cazenave-Gassiot A, Tan TY, Ching J, Kovalik J-P, Wenk MR, Gunawan R, Moore PK, Halliwell B, Tolwinski N and Gruber J (2019) Metabolic stress is a primary pathogenic event in transgenic *Caenorhabditis elegans* expressing pan-neuronal human amyloid beta. *eLife* **8**, e50069.
- The *C. elegans* Sequencing Consortium (1998) Genome sequence of the nematode *C. elegans*: a platform for investigating biology. *Science (New York, N.Y.)* **282**, 2012–2018.
- Thiruchelvam MJ, Powers JM, Cory-Slechta DA and Richfield EK (2004) Risk factors for dopaminergic neuron loss in human alpha-synuclein transgenic mice. *The European Journal of Neuroscience* **19**, 845–854.
- Thygesen C, Metaxas A, Larsen MR and Finsen B (2018) Age-dependent changes in the sarkosyl-insoluble proteome of APPSWE/PS1ΔE9 transgenic mice implicate dysfunctional mitochondria in the pathogenesis of Alzheimer's disease. *Journal of Alzheimer's Disease* **64**, 1247–1259.
- Tieu K (2011) A guide to neurotoxic animal models of Parkinson's disease. *Cold Spring Harbor Perspectives in Medicine* **1**, a009316.
- Tosun D, Landau S, Aisen PS, Petersen RC, Mintun M, Jagust W and Weiner MW (2017) Association between tau deposition and antecedent amyloid-β accumulation rates in normal and early symptomatic individuals. *Brain* **140**, 1499–1512.
- Tran J, Chang D, Hsu F, Wang H and Guo Z (2017) Cross-seeding between Aβ40 and Aβ42 in Alzheimer's disease. *FEBS Letters* **591**, 177–185.
- Treusch S, Hamamichi S, Goodman JL, Matlack KES, Chung CY, Baru V, Shulman JM, Parrado A, Bevis BJ, Valastyan JS, Han H, Lindhagen-persson M, Reiman EM, Evans DA, Bennett DA, Olofsson A, DeJager PL, Tanzi RE, Caldwell KA, Caldwell GA and Lindquist S (2011) Functional links between Abeta toxicity, endocytic trafficking, and Alzheimer's disease risk factors in yeast. *Science (New York, N.Y.)* **334**, 1241–1245.
- Umeda T, Maekawa S, Kimura T, Takashima A, Tomiyama T and Mori H (2014) Neurofibrillary tangle formation by introducing wild-type human tau into APP transgenic mice. *Acta Neuropathologica* **127**, 685–698.
- Vandebroek T, Vanhelmont T, Terwel D, Borghgraef P, Lemaire K, Snauwaert J, Wera S, Van Leuven F and Winderickx J (2005) Identification and isolation of a hyperphosphorylated, conformationally changed intermediate of human protein tau expressed in yeast. *Biochemistry* **44**, 11466–11475.
- Vandebroek T, Terwel D, Vanhelmont T, Gysemans M, Van Haesendonck C, Engelborghs Y, Winderickx J and Van Leuven F (2006) Microtubule binding and clustering of human Tau-4R and Tau-P301L proteins isolated from yeast deficient in orthologues of glycogen synthase kinase-3β or cdk5. *Journal of Biological Chemistry* **281**, 25388–25397.
- Van der Perren A, Toelen J, Casteels C, Macchi F, Van Rompuy AS, Sarre S, Casadei N, Nuber S, Himmelreich U, Osorio Garcia MI, Michotte Y, D'Hooge R, Bormans G, Van Laere K, Gijsbers R, Van den Haute C, Debyser Z and Baekelandt V (2015) Longitudinal follow-up and characterization of a robust rat model for Parkinson's disease based on overexpression of alpha-synuclein with adeno-associated viral vectors. *Neurobiology of Aging* **36**, 1543–1558.
- van Ham TJ, Thijssen KL, Breitling R, Hofstra RMW, Plasterk RHA and Nollen EAA (2008) *C. elegans* model identifies genetic modifiers of α-synuclein inclusion formation during aging. *PLoS Genetics* **4**, e1000027.
- van Ham TJ, Holmberg MA, van der Goot AT, Teuling E, Garcia-Arencibia M, Kim HE, Du D, Thijssen KL, Wiersma M, Burggraaff R, van Bergeijk P, van Rheenen J, Jerre van Veluw G, Hofstra RMW, Rubinsztein DC and Nollen EAA (2010) Identification of MOAG-4/SERF as a regulator of age-related proteotoxicity. *Cell* **142**, 601–612.
- Vanhelmont T, Vandebroek T, De Vos A, Terwel D, Lemaire K, Anandhakumar J, Franssens V, Swinnen E, Van Leuven F and Winderickx J (2010) Serine-409 phosphorylation and oxidative damage define aggregation of human protein tau in yeast. *FEMS Yeast Research* **10**, 992–1005.
- Verduyck M, Vignaud H, Bynens T, Van den Brande J, Franssens V, Cullin C and Winderickx J (2016) Yeast as a model for Alzheimer's disease: latest studies and advanced strategies. In Castrillo JI and Oliver SG (eds). *Systems Biology of Alzheimer's Disease*. pp. New York, NY: Humana Press, pp. 197–215.
- Vicario M, Cieri D, Brini M and Cali T (2018) The close encounter between alpha-synuclein and mitochondria. *Frontiers in Neuroscience* **12**, 388.
- Villemagne VL, Burnham S, Bourgeat P, Brown B, Ellis KA, Salvado O, Szoek C, Macaulay SL, Martins R, Maruff P, Ames D, Rowe CC and Masters CL (2013) Amyloid β deposition, neurodegeneration, and cognitive decline in sporadic Alzheimer's disease: a prospective cohort study. *The Lancet Neurology* **12**, 357–367.
- Visanji NP, Brotchie JM, Kalia LV, Koprach JB, Tandon A, Watts JC and Lang AE (2016) Alpha-synuclein-based animal models of Parkinson's disease: challenges and opportunities in a new era. *Trends in Neurosciences* **39**, 750–762.
- Völgyi K, Hádán K, Kis V, Gulyássy P, Badics K, Györfly BA, Simor A, Szabó Z, Janáky T, Drahos L, Dobolyi Á, Penke B, Juhász G and Kékesi KA (2017) Mitochondrial proteome changes correlating with β-amyloid accumulation. *Molecular Neurobiology* **54**, 2060–2078.
- Wacker J, Rönicker R, Westermann M, Wulff M, Reymann KG, Dobson CM, Horn U, Crowther DC, Luheshi LM and Fändrich M (2014) Oligomer-targeting with a conformational antibody fragment promotes toxicity in Aβ-expressing flies. *Acta Neuropathologica Communications* **2**, 43.
- Wakamatsu M, Ishii A, Iwata S, Sakagami J, Ukai Y, Ono M, Kanbe D, Muramatsu S, Kobayashi K, Iwatsubo T and Yoshimoto M (2008) Selective loss of nigral dopamine neurons induced by overexpression of truncated human alpha-synuclein in mice. *Neurobiology of Aging* **29**, 574–585.
- Wälti MA, Ravotti F, Arai H, Glabe CG, Wall JS, Böckmann A, Güntert P, Meier BH and Riek R (2016) Atomic-resolution structure of a disease-relevant Aβ(1–42) amyloid fibril. *Proceedings of the National Academy of Sciences* **113**, E4976–E4984.
- Wang Y and Mandelkow E (2016) Tau in physiology and pathology. *Nature Reviews Neuroscience* **17**, 5–21.
- Wang Z, Gerstein M and Snyder M (2009) RNA-Seq: a revolutionary tool for transcriptomics. *Nature Reviews Genetics* **10**, 57–63.
- Wang C, Saar V, Leung KL, Chen L and Wong G (2018) Human amyloid β peptide and tau co-expression impairs behavior and causes specific gene expression changes in *Caenorhabditis elegans*. *Neurobiology of Disease* **109**, 88–101.
- Wegmann S, Eftekharzadeh B, Tepper K, Zoltowska KM, Bennett RE, Dujardin S, Laskowski PR, MacKenzie D, Kamath T, Commins C, Vanderburg C, Roe AD, Fan Z, Molliex AM, Hernandez-Vega A, Muller D, Hyman AA, Mandelkow E, Taylor JP and Hyman BT (2018) Tau protein liquid–liquid phase separation can initiate tau aggregation. *The EMBO Journal* **37**, e98049.
- Weiner MW, Veitch DP, Aisen PS, Beckett LA, Cairns NJ, Green RC, Harvey D, Jack CR, Jagust W, Liu E, Morris JC, Petersen RC, Saykin AJ, Schmidt ME, Shaw L, Shen L, Siuciak JA, Soares H, Toga AW and Trojanowski JQ (2013) The Alzheimer's disease neuroimaging initiative: a review of papers published since its inception. *Alzheimer's & Dementia* **9**, e111–e194.
- Weingarten MD, Lockwood AH, Hwo SY and Kirschner MW (1975) A protein factor essential for microtubule assembly. *Proceedings of the National Academy of Sciences of the United States of America* **72**, 1858–1862.
- Wheeler JM, McMillan PJ, Hawk M, Iba M, Robinson L, Xu GJ, Dombroski BA, Jeong D, Dichter MA, Juul H, Loomis E, Raskind M, Leverenz JB, Trojanowski JQ, Lee VMY, Schellenberg GD and Kraemer BC (2015) High copy wild-type human 1N4R tau expression promotes early pathological tauopathy accompanied by cognitive deficits without progressive neurofibrillary degeneration. *Acta Neuropathologica Communications* **3**, 33.

- Williams DW, Tyrer M and Shepherd D (2000) Tau and tau reporters disrupt central projections of sensory neurons in *Drosophila*. *Journal of Comparative Neurology* **428**, 630–640.
- Willingham S, Outeiro TF, Devit MJ, Lindquist SL, Muchowski PJ, Willingham S, Outeiro TF, Devit MJ, Lindquist SL and Muchowski PJ (2003) Yeast genes that enhance the toxicity of a mutant Huntingtin fragment or α -synuclein. *Science (New York, N.Y.)* **302**, 1769–1773.
- Wirhth O, Breyhan H, Schäfer S, Roth C and Bayer TA (2008) Deficits in working memory and motor performance in the APP/PS1ki mouse model for Alzheimer's disease. *Neurobiology of Aging* **29**, 891–901.
- Wittmann CW, Wszolek MF, Shulman JM, Salvaterra PM, Lewis J, Hutton M and Feany MB (2001) Tauopathy in *Drosophila*: neurodegeneration without neurofibrillary tangles. *Science (New York, N.Y.)* **293**, 711–714.
- Wu Y, Wu Z, Butko P, Christen Y, Lambert MP, Klein WL, Link CD and Luo Y (2006) Amyloid-beta-induced pathological behaviors are suppressed by *Ginkgo biloba* extract EGb 761 and ginkgolides in transgenic *Caenorhabditis elegans*. *The Journal of neuroscience* **26**, 13102–13113.
- Xia C, Makarets SJ, Caso C, McGinnis S, Gomperts SN, Sepulcre J, Gomez-Isla T, Hyman BT, Schultz A, Vasdev N, Johnson KA and Dickerson BC (2017) Association of *in vivo* [18 F]AV-1451 tau PET imaging results with cortical atrophy and symptoms in typical and atypical Alzheimer disease. *JAMA Neurology* **74**, 427–436.
- Xiao Y, Ma B, McElheny D, Parthasarathy S, Long F, Hoshi M, Nussinov R and Ishii Y (2015) A β (1-42) fibril structure illuminates self-recognition and replication of amyloid in Alzheimer's disease. *Nature Structural & Molecular Biology* **22**, 499–505.
- Xu G, Stevens SM, Moore BD, McClung S and Borchelt DR (2013) Cytosolic proteins lose solubility as amyloid deposits in a transgenic mouse model of Alzheimer-type amyloidosis. *Human Molecular Genetics* **22**, 2765–2774.
- Xu J, Du Y and Deng H (2015) Direct lineage reprogramming: strategies, mechanisms, and applications. *Cell Stem Cell* **16**, 119–134.
- Yan J, Zhang P, Jiao F, Wang Q, He F, Zhang Q, Zhang Z, Lv Z, Peng X, Cai H and Tian B (2017) Quantitative proteomics in A30P/A53T α -synuclein transgenic mice reveals upregulation of Sell1. *PLoS ONE* **12**, e0182092.
- Yang X, Meisl G, Frohm B, Thulin E, Knowles TPJ and Linse S (2018) On the role of sidechain size and charge in the aggregation of A β 42 With familial mutations. *Proceedings of the National Academy of Sciences* **115**, E5849–E5858.
- Yeger-Lotem E, Riva L, Su LJ, Gitler AD, Cashikar AG, King OD, Auluck PK, Geddie ML, Valastyan JS, Karger DR, Lindquist S and Fraenkel E (2009) Bridging high-throughput genetic and transcriptional data reveals cellular responses to alpha-synuclein toxicity. *Nature Genetics* **41**, 316–323.
- Yoshiyama Y, Higuchi M, Zhang B, Huang SM, Iwata N, Saido TCC, Maeda J, Suhara T, Trojanowski JQ and Lee VMY (2007) Synapse loss and microglial activation precede tangles in a P301S tauopathy mouse model. *Neuron* **53**, 337–351.
- Yu-Jen C and Yun-Ru C (2014) The coexistence of an equal amount of Alzheimer's amyloid- β 40 and 42 forms structurally stable and toxic oligomers through a distinct pathway. *The FEBS Journal* **281**, 2674–2687.
- Zabrocki P, Pellens K, Vanhelmont T, Vandebroek T, Griffioen G, Wera S, Van Leuven F and Winderickx J (2005) Characterization of alpha-synuclein aggregation and synergistic toxicity with protein tau in yeast. *FEBS Journal* **272**, 1386–1400.
- Zhang F (2019) Development of CRISPR-Cas systems for genome editing and beyond. *Quarterly Reviews of Biophysics* **52**, e6.
- Zhang H, Komano H, Fuller RS, Gandy SE and Frail DE (1994) Proteolytic processing and secretion of human beta-amyloid precursor protein in yeast. Evidence for a yeast secretase activity. *The Journal of Biological Chemistry* **269**, 27799–27802.
- Zhang W, Espinoza D, Hines V, Innis M, Mehta P and Miller DL (1997) Characterization of β -amyloid peptide precursor processing by the yeast Yap3 and Mkc7 proteases. *Biochimica et Biophysica Acta – Molecular Cell Research* **1359**, 110–122.
- Zhao X-L, Wang W-A, Tan J-X, Huang J-K, Zhang X, Zhang B-Z, Wang Y-H, Yang Cheng H-Y, Zhu H-L, Sun X-J and Huang F-D (2010) Expression of beta-amyloid induced age-dependent presynaptic and axonal changes in *Drosophila*. *The Journal of Neuroscience* **30**, 1512–1522.
- Zhong Q, Congdon EE, Nagaraja HN and Kuret J (2012) Tau isoform composition influences rate and extent of filament formation. *Journal of Biological Chemistry* **287**, 20711–20719.
- Zilka N, Filipcik P, Koson P, Fialova L, Skrabana R, Zilkova M, Rolkova G, Kontsekkova E and Novak M (2006) Truncated tau from sporadic Alzheimer's disease suffices to drive neurofibrillary degeneration *in vivo*. *FEBS Letters* **580**, 3582–3588.